# EXHIBIT 1

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# (54) GUIDE RNA WITH CHEMICAL MODIFICATIONS

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# (58) Field of Classification Search

None

See application file for complete search history.

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# (57) ABSTRACT

The present invention relates to modified guide RNAs and their use in clustered, regularly interspaced, short palindromic repeats (CRISPR)/CRISPR-associated (Cas) systems.

# 33 Claims, 18 Drawing Sheets

Specification includes a Sequence Listing.

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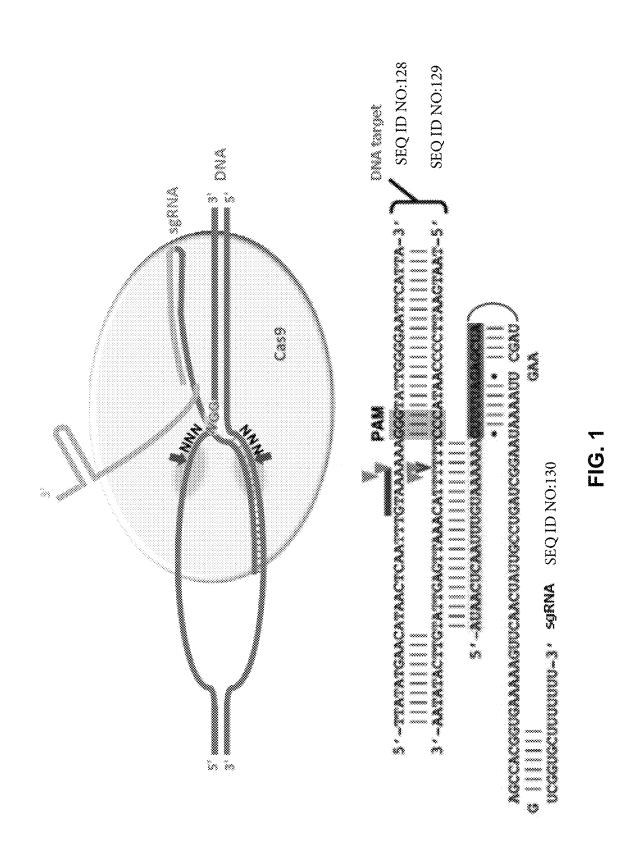
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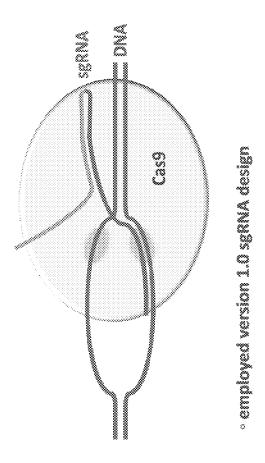
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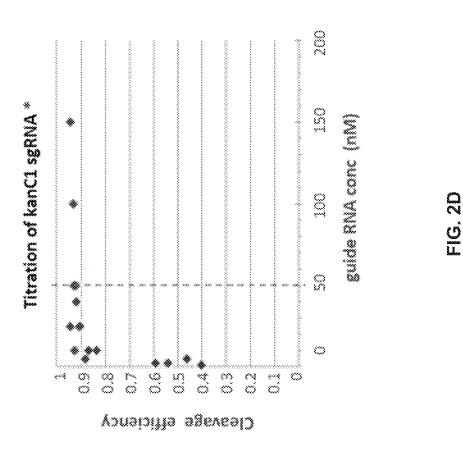
	8				
	40 nM (125 ng/20 ulman	2007	100 mM	2 0 0 5	30 mW
	\$ 0 8				
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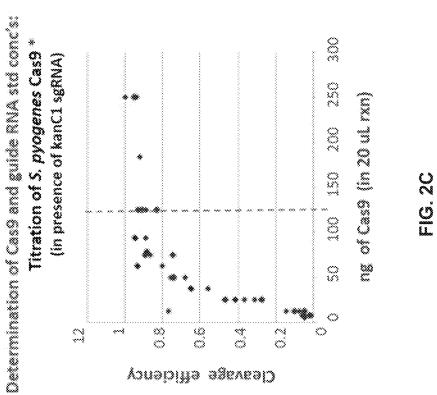
T. C. 2



HG. 2

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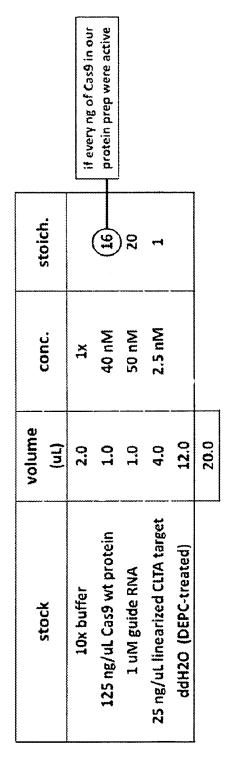


Standard cleavage conditions:

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In a pre-warmed SureCycler:

(i) incubate at 37 °C for 30 min

(ii) + RNase cocktail, incubate at 37 °C for 5 min, then at 70 °C for 15 min

(iii) + Proteinase K, incubate at 37 °C for 15 min

Analyze crude products on Bioanalyzer.

FIG. 3

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	C Cu	, on 0.45	% Cleaved	
Name of synthetic guide RNA	<u> </u>		target in vitro	Sequence $(5' \rightarrow 3')$
Chemical modifications tolerated		by Cas9	маличения	
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGG
CLTA1 (unmodified control)	42	(1) (1)	%56	CACCGAGUCGGUGCUUUUUU
				A*G*UCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAG
				CAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGU
2xOWePACE_CLTA1	6	~ ~ ~	95%	GGCACCGAGUCGGUGCUUUUUUU
				A*G*U*C*CUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAAC
				AGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAA
4xOWePACE_CLTA1	భ	(건 단 단	%88	guggcaccgagucggugcuuuuuu
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGG
CLTA1_4xOMePACE	800	~1 ~1 ~1	91%	CACCGAGUCGGUGCUUU*U*U*U*U
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGG
CLTA1_5xOWePACE	96	133	91%	CACCGAGUCGGUGCUU*U*U*U*U*U
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGG
CLTA1_2'OMe+20	67	(A)	%86	CACCGAGUCGGUGCUUUUUUU
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGG
CLTA1_2'OMe+19	88	67 (m) (m)	93%	CACCGAGUCGGUGCUUUUUU
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGG
CLTA1_2'OMe+18	9	(Y) (Y)	91%	CACCGAGUCGGUGCUUUUUU
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGG
CLTA1_2'OMe+17	70	~ ~ ~	93%	CACCGAGUCGGUGCUUUUUU

FIG. 4

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В применения применения в пр	<b>принципринципринципринципринципринципринципринципринципринципринципринципринципринципринципринципринципринципри</b>	**************************************	**************************************	A CELO TOPO TOPO TOPO TO BE A STATE OF THE S
				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGG
CLTA1_2'OMe+17,18	7	644 644	%06	cacceaeuceeuccuuuuuu
CLTA1 20 Deoxy				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACACACA UAGCAAGUUUAAAAAAAAGGCUAGUCGGUUAACAAAAAGUGG
	*~! (V)	64) (43)	7%	CACCGAGUCGGUGCUUUUUU
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
CLTA1_20_2'OMe				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGG
	4	የና ፫ጣ ፻ጣ	%68	cacceaeuceeuecuuuuuu
OTA1 37 3/08/3				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
	76	~~! ~~! ~~!	88%	CACCGAGUCGGUGCUUUUUUU
	7,		***************************************	AsGSUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGAAACAG
				CAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGU
CTLA1_5'SS	63	673 E-4 E-4	%96	GGCACCGAGUCGGUGCUUUUUU
				AsGSUSCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGAAACA
				GCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAG
CTLA1_5'SSS	64	773	94%	UGGCACCGAGUCGGUGCUUUUUU
				Asgsuscscucaucucccucaagcguunaagagcuaugcuggaaac
				AGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAA
CTLA1_5'SSSS	S	H 33	100%	GUGGCACCGAGUCGGUGCUUUUUU
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGAAACAGCA
				UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGG
CTLA1_3'SSSS	99	(주) (~)	94%	CACCGAGUCGGUGCUUUsUsUsU
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
	į	,		UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGG
3xOMe_CLTA1_3xOMe	77	(A) (-1) (-1)	%68	caccaagucaguacuuuuuu
				Ascsuscoucaucuccoucaaccouuaagaccuaugcuguaca
SACINIE CLI AL SACINIE III				GCAUAGCAAGUUUAAAUAAGCUAGUCCGUUAUCAACUUGAAAAG
0	107	የጣ የጣ የጣ	%26	uggcaccgagucggugcuuusususu
2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2				A*sG*sU*sCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUA
OMeThiopace_ce.fat_sxz	~	Ann Ann East (An	%68	ACAGCACCACCACCCACCCITIM*********************************
			27.55	

FIG. 4 (cont.

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				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
				UAGCAAGUUUAAAUAAGGCUAGUZZGUUAUCAACUUGAAAAAGUGG
CLTA1_22_70,71	122	(1) (m)	19%	CACCGAGUCGGUGCUUUUUU
				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA
CLTA1 22_95,96	132	e e	93%	UAGCAAGUUUAAAUAAGCUAGUCCGUUAUCAACUUGAAAAAGUGG
			*****	CANNGAGUCGGUGCUUUUUU
				(dmt) AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAA
CLTA1_QB3+GNRA_DMT-ON	ស្ល	۳ ۳	93%	CAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAA
-				AGUGGCACCGAGUCGGUGCUUUUUU
Chemical modifications NOT tolerated by Cas9	ins NOT to	lerated by Cas	තු	
CLTA1 37 Deoxy				AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGGG
	133	733	%0	CACCGAGUCGGUGCUUUUUU
**************************************	decement	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	, manusament and a second	одинивання винивання винив

LEGEND

 $\underline{\underline{\mathbf{M}}^*} = 2.0$ Me,3'PACE modification of nucleotide N

 $\underline{\underline{\mathbf{N}}}^* \mathbf{s} = 2'OMe, 3' \text{thioPACE modification of nucleotide N}$ 

 $\underline{\underline{\mathbf{N}}} = 2^{\circ}$ OMe modification of nucleotide N

 $\frac{N}{m}$  = 2'deoxyribonucleotide N

NsN = phosphorothioate linkage noted by s

2 = 2 nucleotide

dmt = dimethoxytrity!

FIG. 4 (cont.)

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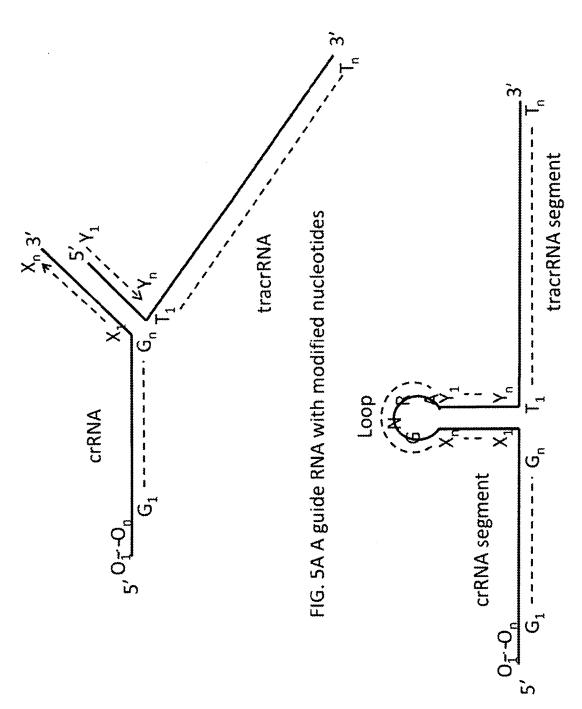
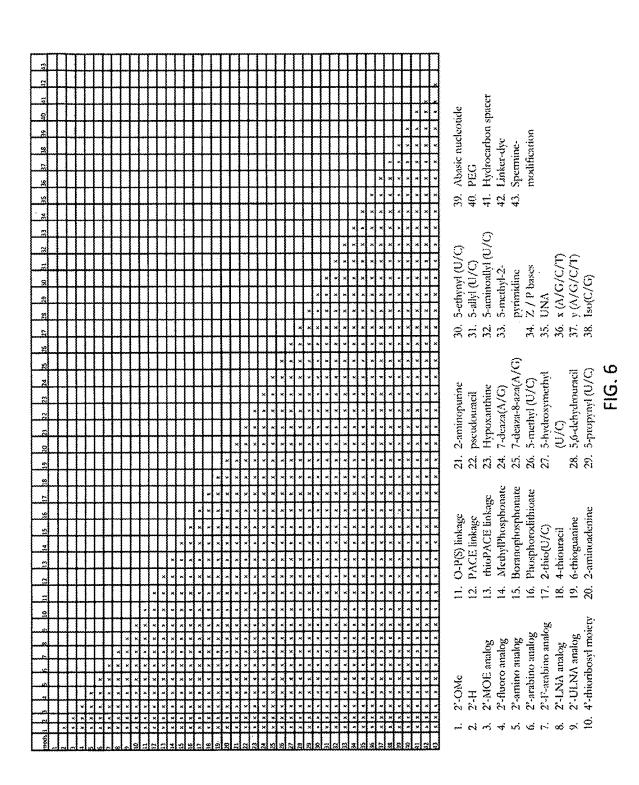


FIG. 5B A single guide RNA with modified nucleotides

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Single mod	Sugar	Phosphorus Linkage	Base modification*	Other
Double mod	3	3		
Sugar/Sugar	×	Х	×	×
Sugar/ P link	×	X	×	X
Sugar/Base	×	×	×	×
Sugar/other	X	X	×	X
P link/ Plink	×	X	×	×
P link/ base	×	×	×	×
P link/other	×	Х	×	×
Base/Base	Х	X	×	×
Base/other	Х	Х	X	×
other/other	×	×	×	×

<sup>\*</sup>Base modifications includes Base Pair Modifications

Sugar modifications ["Sugar"]: 2'-0-Methyl (=2'-0Me) (2'-0C<sub>1</sub>-C<sub>4</sub> alkyl), 2'-H, 2'-MOE (2'-0C<sub>1</sub>-C<sub>3</sub> alkyl-0C<sub>1</sub>-C<sub>3</sub> alkyl), 2'-F, 2'-amino, 2'arabino, 2'-f-arabino, 2'-LNA, 2'-UNLA, 4'-thioribosyl nucleotide.

phosphorothioate), -PACE (phosphonoacetate, phosphonocarboxylate), -thioPACE (thiophosphonoacetate, thiophosphonocarboxylate), Base modifications: 2-thiouracil, 2-thiocytosine, 4-thiouracil, 6-thioguanine, 2-aminoadenine, 2-aminopurine, pseudouracil, internucleotide linkage and 3' and/or 5' terminal nucleotide modifications ("Phosphorus Linkage" or "P link"); -P(S) -P(CH<sub>3</sub>) (methyiphosphonate, alkyiphosphonate), -P(BH<sub>3</sub>) (boranophosphonate), -P(S)<sub>2</sub> (phosphorodithioate)

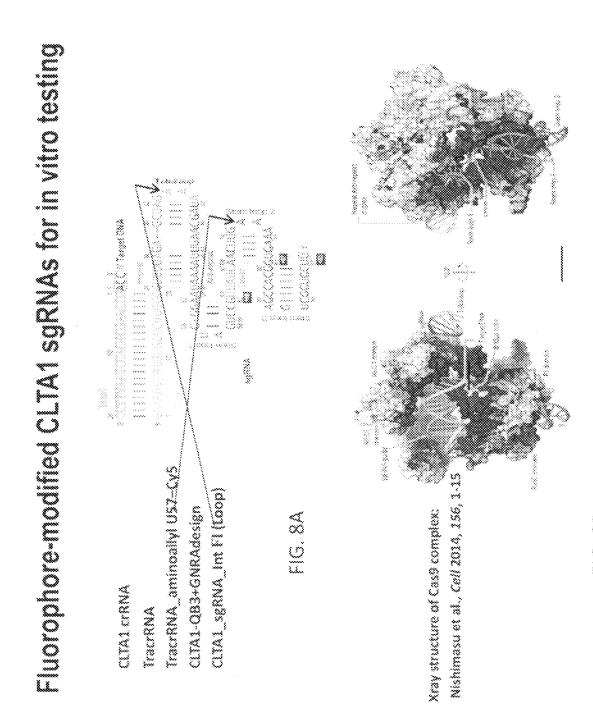
hypoxanthine, 7-deazaguanine, 7-deaza-8-azaguanine, 7-deazaadenine, 7-deaza-8-azaadenine, 5-methylcytosine, 5-methyluracil, 5hydroxymethylcytosine, 5-hydroxymethyluracil, 5,6-dehydrouracil, 5-propynylcytosine, 5-propynyluracil, 5-ethynylcytosine,

Base Pair modifications: Z/P nucleotides, UNA, isoC/isoG, 6-thioG/5-methyl-pyrimidine, x(A,G,C,T) and y(A,G,C,T) ethynyluracil, 5-allyluracil, 5-allylcytosine, 5-aminoallyl-uracil, 5-aminoallyl-cytosine and abasic nucleotides.

Other: End modifications and/or spacer/linker (ends or internal) modifications: PEG, hydrocarbon spacer, (including: heteroatom O,S,N)-substituted hydrocarbon spacers, halo-substituted-hydrocarbon spacers, keto, carboxy, amido, thionyl, carbamoyl, thionocarbama oyl)-containing hydrocarbon spacers), spermine, dyes linkers including: 6-·luorescein-phosphoramídite and the like, squarate conjugation, Diels-Alder conjugation, or "Click" chemistry conjugation.

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φ <u>ψ</u> <u>Ψ</u>

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lowers specificity. wobble pair that

# Using chemical modifications to improve specificity

Target Name	CLTA1 On- or Off-Target Site	Genomic Coordinates	COSMID Score	MIT Design Score
CLTA1 ON1	AGTCCTCATCTCCCTCAAGCAGG TCAGGAGTAGAGGGAGTTCGTCC	Chr9:36211735-36211757	0	100.0
CLTA1 OFF1	AGTCCTCAACTCCCTCAAGCAGG TCAGGAGTTGAGGGAGTTCGTCC	Chr8:15688928-15688950	0.35	61.1
CLTA1 OFF2	AGCCTCATTTCCCTCAAGCAGG TCGGGAGTAAAGGGAGTTCGTCC	Chr3:54189084-54189106	9.0	6.4
CLTA1 OFF3	actecteatececeteargeges tgaggagtaggggggggggggggggggggggggggggg	Chr15:88845439-88845461	68.0	4.5

cannot H-bond with Sulfur in 2-thioU G to form a

G

Example: • CLTA1\_2thioU+11 crRNA: 5' AGUCCUCAUC (2sU) CCCUCAAGCGUUUAAGAGCUAUGCUGUUUGA

AUGGUCCCAAAAC 3'

• CLTA1\_2thioU+9

• CLTA1\_2thioU+3

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2-thioU can increase target specificity of guide RNAs when off-target sites involve U-G wobble pairing

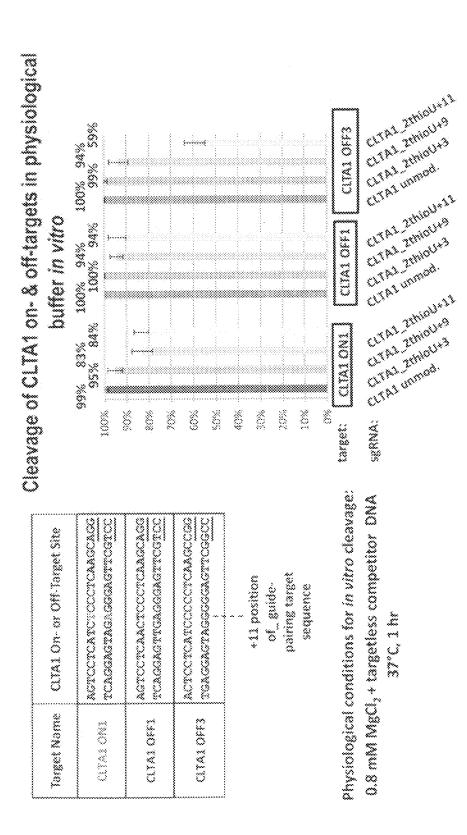
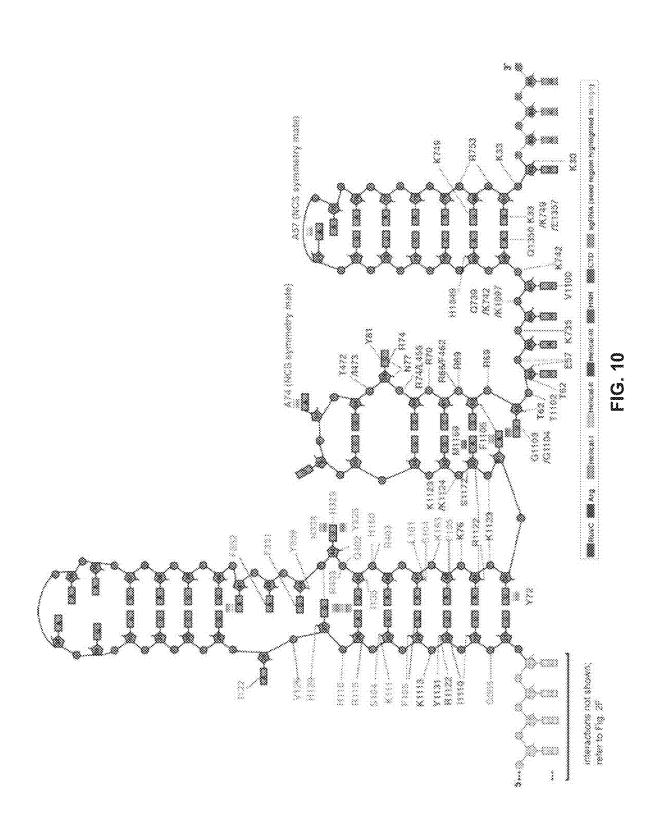


FIG. 98

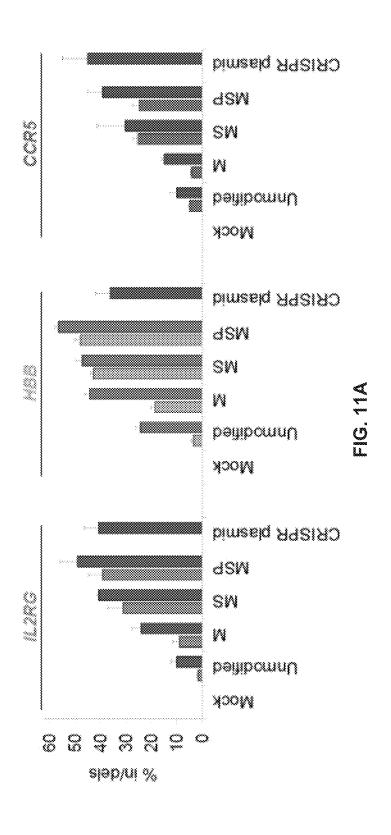
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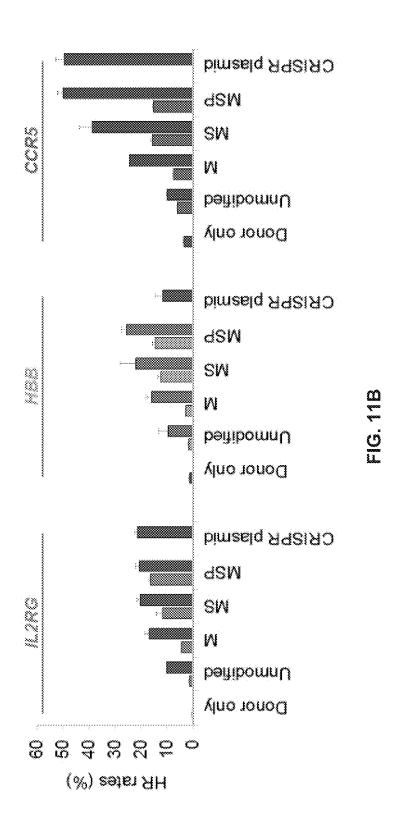
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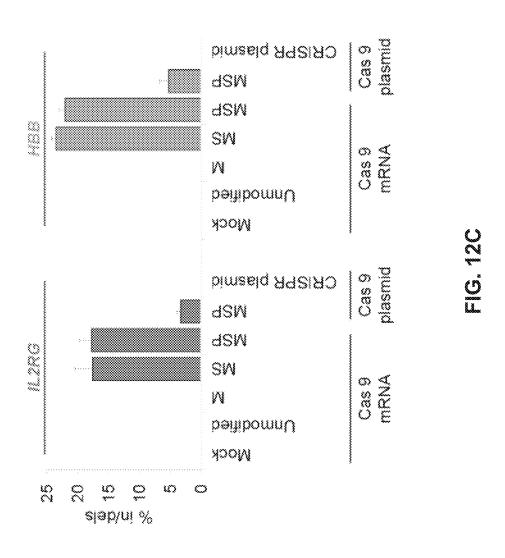


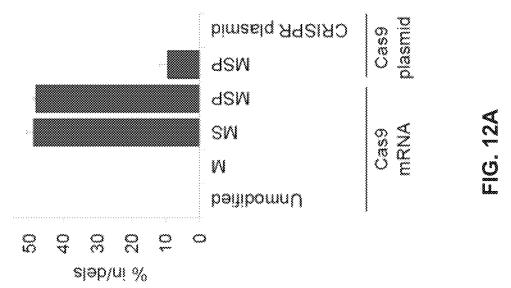
U.S. Patent

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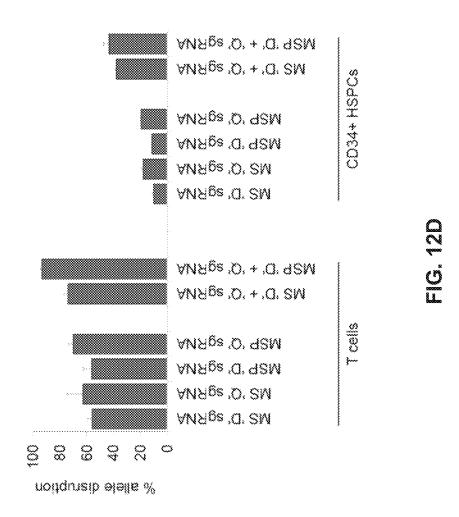
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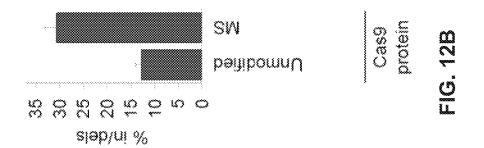




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# 1

# GUIDE RNA WITH CHEMICAL MODIFICATIONS

# CROSS REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Application No. 62/256,095, filed Nov. 16, 2015, U.S. Provisional Application No. 62/146,189, filed Apr. 10, 2015, and U.S. Provisional Application No. 62/087,211, filed Dec. 3, 2014, the contents of each of which is incorporated by reference in its entirety.

# SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted electronically in ASCII format and is hereby incorporated by reference in its entirety. Said ASCII copy, created on Aug. 5, 2020, is named 20160013\_ 04\_ SL.txt and is 114,989 bytes in size

## FIELD OF THE INVENTION

The present invention relates to the field of molecular biology. In particular, the present invention relates to the <sup>25</sup> clusters of regularly interspaced short palindromic repeats (CRISPR) technology.

## BACKGROUND OF THE INVENTION

The native prokaryotic CRISPR-Cas system comprises an array of short repeats with intervening variable sequences of constant length (i.e., clusters of regularly interspaced short palindromic repeats, or "CRISPR"), and CRISPR-associated ("Cas") proteins. The RNA of the transcribed CRISPR array 35 is processed by a subset of the Cas proteins into small guide RNAs, which generally have two components as discussed below. There are at least three different systems: Type I, Type II and Type III. The enzymes involved in the processing of the RNA into mature crRNA are different in the 3 40 systems. In the native prokaryotic system, the guide RNA ("gRNA") comprises two short, non-coding RNA species referred to as CRISPR RNA ("crRNA") and trans-acting RNA ("tracrRNA"). In an exemplary system, the gRNA forms a complex with a Cas nuclease. The gRNA:Cas 45 nuclease complex binds a target polynucleotide sequence having a protospacer adjacent motif ("PAM") and a protospacer, which is a sequence complementary to a portion of the gRNA. The recognition and binding of the target polynucleotide by the gRNA:Cas nuclease complex induces 50 cleavage of the target polynucleotide. The native CRISPR-Cas system functions as an immune system in prokaryotes, where gRNA:Cas nuclease complexes recognize and silence exogenous genetic elements in a manner analogous to RNAi in eukaryotic organisms, thereby conferring resistance to 55 exogenous genetic elements such as plasmids and phages.

It has been demonstrated that a single-guide RNA ("sgRNA") can replace the complex formed between the naturally-existing crRNA and tracrRNA.

Considerations relevant to developing a gRNA, including 60 a sgRNA, include specificity, stability, and functionality. Specificity refers to the ability of a particular gRNA:Cas nuclease complex to bind to and/or cleave a desired target sequence, whereas little or no binding and/or cleavage of polynucleotides different in sequence and/or location from 65 the desired target occurs. Thus, specificity refers to minimizing off-target effects of the gRNA:Cas nuclease com-

# 2

plex. Stability refers to the ability of the gRNA to resist degradation by enzymes, such as nucleases, and other substances that exist in intra-cellular and extra-cellular environments. Thus, there is a need for providing gRNA, including sgRNA, having increased resistance to nucleolytic degradation, increased binding affinity for the target polynucleotide, and/or reduced off-target effects while, nonetheless, having gRNA functionality. Further considerations relevant to developing a gRNA include transfectability and immunostimulatory properties. Thus, there is a need for providing gRNA, including sgRNA, having efficient and titratable transfectability into cells, especially into the nuclei of eukaryotic cells, and having minimal or no immunostimulatory properties in the transfected cells. Another impor-15 tant consideration for gRNA is to provide an effective means for delivering it into and maintaining it in the intended cell, tissue, bodily fluid or organism for a duration sufficient to allow the desired gRNA functionality.

### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a set of diagrams showing a schematic model of an exemplary CRISPR-Cas system. The exemplary system shown here is a Type II system having a Cas nuclease. In this particular example, the Cas nuclease is the Cas9 nuclease. The Cas9 nuclease recognizes a PAM sequence (here, the PAM sequence is a 3-nt sequence of NGG, where N is A, G, C or T, but other PAM sequences are known to exist). The sgRNA includes a guide sequence, a crRNA sequence or segment, and tracrRNA sequence or segment. The guide sequence of the sgRNA hybridizes with the DNA target directly upstream of the PAM sequence. In the example shown here, Cas9 mediates a double-stranded break upstream of the PAM sequence (arrows).

FIG. **2**A is a diagram showing an exemplary CRISPR-Cas9-mediated cleavage assay.

FIG. 2B is a table showing components and their concentrations for a biochemical cleavage assay used to generate the data in FIG. 4.

FIG. **2**C is a diagram showing titration of *Streptococcus pyogenes* Cas9 nuclease for the biochemical cleavage assay.

FIG. 2D is a diagram showing titration of an exemplary sgRNA for the biochemical cleavage assay. In this example a sgRNA named kanC1 is targeted to a complementary sequence in the kanamycin resistance gene.

FIG. 3 shows exemplary conditions and procedures for the biochemical cleavage assay which uses purified components in vitro.

FIG. 4 is a table showing the data obtained using exemplary modified guide RNAs in the cleavage assay.

FIG. 5A shows an exemplary guide RNA disclosed in the application.

FIG. 5B shows an exemplary single guide RNA (sgRNA) disclosed in the application.

FIG. 6 is a table showing exemplary guide RNAs having at least two chemical modifications (e.g., a first modification and a second modification). Each number represents a modification as indicated and each "x" indicates the combination of modifications in a guide RNA. In certain embodiments, the first and second modifications are present on a single nucleotide. In certain embodiments, the first and second modifications are present on separate nucleotides.

FIG. 7 shows exemplary types of guide RNAs having at least three chemical modifications. The lower part of FIG. 7 lists several types of modifications. The table in the upper part of FIG. 7 indicates how a double modification ("double mod," a combination of two types of modifications) can be

combined with a single modification ("single mod," one type of modification). An "x" indicates the presence of the corresponding double mod and single mod in a guide RNA.

3

FIGS. **8**A and **8**B show fluorophore-modified CLTA1 sgRNAs for in vitro testing. In FIG. **8**A, the RNA sequence (SEQ ID NO: 135) of a sgRNA for CLTA1 is shown, including a position where a fluorescent dye or a label could be attached to the sgRNA. Target DNA is SEQ ID NO: 134. FIG. **8**B shows a structure determined by Xray crystallography of a Cas9:sgRNA complex, as reported in Nishimasu et al., *Cell* 2014, 156, 1-15.

FIG. 9A shows CLTA1 sgRNAs modified with 2-thiouridine at certain locations (positions 3, 9 and 11) in an effort to improve specificity for the target CTLA1. Top strand and bottom strand sequences (respectively) of the CTLA1 targets are: ON1 (SEQ ID NOs: 136 and 137); OFF1 (SEQ ID NOs: 138 and 139); OFF2 (SEQ ID NOs: 140 and 141); and OFF3 (SEQ ID NOs: 142 and 143). FIG. 9B shows that gRNA modified with 2-thioU (SEQ ID NO: ###) can 20 increase target specificity of the gRNAs when off-target sites involve U-G wobble pairing. In particular, the CTLA1\_2thioU+11 had much lower cleavage of the offtarget sequence CLTA1 OFF3, which has a T to C mutation at the 11 position in the 5' strand. Top strand and bottom 25 strand sequences (respectively) of the CTLA1 targets are: ON1 (SEQ ID NOs: 136 and 137); OFF1 (SEQ ID NOs: 138 and 139); and OFF3 (SEQ ID NOs: 142 and 143).

FIG. 10 shows the guide RNA scaffold secondary structure, displaying noncovalent binding interactions with amino acids of Cas9, as reported in Jiang et al., Science (2015) 348:6242, 1477-81.

FIGS. 11A and 11B illustrate experimental results showing that gene disruption in human cell lines, with high frequencies of indels and homologous recombination (HR), can be achieved using synthesized and chemically modified sgRNAs disclosed herein, as reported in Hendel et al., *Nat. Biotechnol.* (2015) 33:9, 985-9.

FIGS. 12A, 12B, 12C and 12D illustrate experimental 40 results showing that chemically modified sgRNAs as described herein can be used to achieve high frequencies of gene disruption or targeted genome editing in stimulated primary human T cells as well as in CD34+ hematopoietic stem and progenitor cells (HSPCs), as reported in Hendel et 45 al., *Nat. Biotechnol.* (2015) 33:9, 985-9.

# DETAILED DESCRIPTION OF THE INVENTION

This invention is based, at least in part, on an unexpected discovery that certain chemical modifications to gRNA are tolerated by the CRISPR-Cas system. In particular, certain chemical modifications believed to increase the stability of the gRNA, to alter the thermostability of a gRNA hybridization interaction, and/or to decrease the off-target effects of Cas:gRNA complexation do not substantially compromise the efficacy of Cas:gRNA binding to, nicking of, and/or cleavage of the target polynucleotide. Furthermore, certain 60 chemical modifications are believed to provide gRNA, including sgRNA, having efficient and titratable transfectability into cells, especially into the nuclei of eukaryotic cells, and/or having minimal or no immunostimulatory properties in the transfected cells. Certain chemical modi- 65 fications are believed to provide gRNA, including sgRNA, which can be effectively delivered into and maintained in the

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intended cell, tissue, bodily fluid or organism for a duration sufficient to allow the desired gRNA functionality.

### I. Definitions

As used herein, the term "guide RNA" generally refers to an RNA molecule (or a group of RNA molecules collectively) that can bind to a Cas protein and aid in targeting the Cas protein to a specific location within a target polynucleotide (e.g., a DNA). A guide RNA can comprise a crRNA segment and a tracrRNA segment. As used herein, the term "crRNA" or "crRNA segment" refers to an RNA molecule or portion thereof that includes a polynucleotide-targeting guide sequence, a stem sequence, and, optionally, a 5'-overhang sequence. As used herein, the term "tracrRNA" or "tracrRNA segment" refers to an RNA molecule or portion thereof that includes a protein-binding segment (e.g., the protein-binding segment is capable of interacting with a CRISPR-associated protein, such as a Cas9). The term "guide RNA" encompasses a single guide RNA (sgRNA), where the crRNA segment and the tracrRNA segment are located in the same RNA molecule. The term "guide RNA" also encompasses, collectively, a group of two or more RNA molecules, where the crRNA segment and the tracrRNA segment are located in separate RNA molecules.

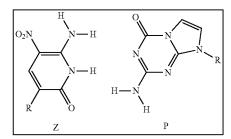
The term "scaffold" refers to the portions of guide RNA molecules comprising sequences which are substantially identical or are highly conserved across natural biological species. Scaffolds include the tracrRNA segment and the portion of the crRNA segment other than the polynucleotide-targeting guide sequence at or near the 5' end of the crRNA segment, excluding any unnatural portions comprising sequences not conserved in native crRNAs and tracrRNAs.

The term "nucleic acid", "polynucleotide" or "oligonucleotide" refers to a DNA molecule, an RNA molecule, or analogs thereof. As used herein, the terms "nucleic acid", "polynucleotide" and "oligonucleotide" include, but are not limited to DNA molecules such as cDNA, genomic DNA or synthetic DNA and RNA molecules such as a guide RNA, messenger RNA or synthetic RNA. Moreover, as used herein, the terms "nucleic acid" and "polynucleotide" include single-stranded and double-stranded forms.

The term "modification" in the context of an oligonucleotide or polynucleotide includes but is not limited to (a) end
modifications, e.g., 5' end modifications or 3' end modifi50 cations, (b) nucleobase (or "base") modifications, including
replacement or removal of bases, (c) sugar modifications,
including modifications at the 2', 3', and/or 4' positions, and
(d) backbone modifications, including modification or
replacement of the phosphodiester linkages. The term
55 "modified nucleotide" generally refers to a nucleotide having a modification to the chemical structure of one or more
of the base, the sugar, and the phosphodiester linkage or
backbone portions, including nucleotide phosphates.

The terms "Z" and "P" refer to the nucleotides, nucleobases, or nucleobase analogs developed by Steven Benner and colleagues as described for example in "Artificially expanded genetic information system: a new base pair with an alternative hydrogen bonding pattern" Yang, Z., Hutter, D., Sheng, P., Sismour, A. M. and Benner, S. A. (2006) *Nucleic Acids Res.*, 34, 6095-101, the contents of which is hereby incorporated by reference in its entirety.

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The terms "xA", "xG", "xC", "xT", or "x(A,G,C,T)" and "yA", "yG", "yC", "yT", or "y(A,G,C,T)" refer to nucleotides, nucleobases, or nucleobase analogs as described by Krueger et al in "Synthesis and Properties of Size-Expanded DNAs: Toward Designed, Functional Genetic Systems"; Andrew T. Krueger, Haige Lu, Alex H. F. Lee, and Eric T. Kool (2007) *Acc. Chem. Res.*, 40, 141-50, the contents of which is hereby incorporated by reference in its entirety.

The term "Unstructured Nucleic Acid" or "UNA" refers to nucleotides, nucleobases, or nucleobase analogs as described in U.S. Pat. No. 7,371,580, the contents of which is hereby incorporated by reference in its entirety. An 25 unstructured nucleic acid, or UNA, modification is also referred to as a "pseudo-complementary" nucleotide, nucleobase or nucleobase analog (see e.g., Lahoud et al. (1991) *Nucl. Acids Res.*, 36:10, 3409-19).

The terms "PACE" and "thioPACE" refer to internucle- 30 otide phosphodiester linkage analogs containing phosphonoacetate or thiophosphonoacetate groups, respectively. These modifications belong to a broad class of compounds comprising phosphonocarboxylate moiety, phosphonocarboxylate ester moiety, thiophosphonocarboxylate moiety 35 and thiophosphonocarboxylate ester moiety. These linkages can be described respectively by the general formulae  $P(CR_1R_2)_nCOOR$  and  $(S)-P(CR_1R_2)_nCOOR$  wherein n is an integer from 0 to 6 and each of R<sub>1</sub> and R<sub>2</sub> is independently selected from the group consisting of H, an alkyl and 40 substituted alkyl. Some of these modifications are described by Yamada et al. in "Synthesis and Biochemical Evaluation of Phosphonoformate Oligodeoxyribonucleotides" Christina M. Yamada, Douglas J. Dellinger and Marvin H. Caruthers (2006) J. Am. Chem. Soc. 128:15, 5251-61, the contents of 45 which is hereby incorporated by reference in its entirety.

As used herein, "modification" refers to a chemical moiety, or portion of a chemical structure, which differs from that found in unmodified ribonucleotides, namely adenosine, guanosine, cytidine, and uridine ribonucleotides. The term 50 "modification" may refer to type of modification. For example, "same modification" means same type of modification, and "the modified nucleotides are the same" means the modified nucleotides have the same type(s) of modification while the base (A, G, C, U, etc.) may be different. 55 Similarly, a guide RNA with "two modifications" is a guide RNA with two types of modifications, which may or may not be in the same nucleotide, and each type may appear in multiple nucleotides in the guide RNA. Similarly, a guide RNA with "three modifications" is a guide RNA with three 60 types of modifications, which may or may not be in the same nucleotide, and each type may appear in multiple nucleo-

As used herein, the term "target polynucleotide" or "target" refers to a polynucleotide containing a target nucleic 65 acid sequence. A target polynucleotide may be single-stranded or double-stranded, and, in certain embodiments, is

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double-stranded DNA. In certain embodiments, the target polynucleotide is single-stranded RNA. A "target nucleic acid sequence" or "target sequence," as used herein, means a specific sequence or the complement thereof that one wishes to bind to, nick, or cleave using a CRISPR system.

The term "hybridization" or "hybridizing" refers to a process where completely or partially complementary polynucleotide strands come together under suitable hybridization conditions to form a double-stranded structure or region in which the two constituent strands are joined by hydrogen bonds. As used herein, the term "partial hybridization" includes where the double-stranded structure or region contains one or more bulges or mismatches. Although hydrogen bonds typically form between adenine and thymine or adenine and uracil (A and T or A and U) or cytosine and guanine (C and G), other noncanonical base pairs may form (See e.g., Adams et al., "The Biochemistry of the Nucleic Acids," 11th ed., 1992). It is contemplated that modified nucleotides may form hydrogen bonds that allow or promote hybridization.

The term "cleavage" or "cleaving" refers to breaking of the covalent phosphodiester linkage in the ribosylphosphodiester backbone of a polynucleotide. The terms "cleavage" or "cleaving" encompass both single-stranded breaks and double-stranded breaks. Double-stranded cleavage can occur as a result of two distinct single-stranded cleavage events. Cleavage can result in the production of either blunt ends or staggered ends.

The term "CRISPR-associated protein" or "Cas protein" refers to a wild type Cas protein, a fragment thereof, or a mutant or variant thereof. The term "Cas mutant" or "Cas variant" refers to a protein or polypeptide derivative of a wild type Cas protein, e.g., a protein having one or more point mutations, insertions, deletions, truncations, a fusion protein, or a combination thereof. In certain embodiments, the "Cas mutant" or "Cas variant" substantially retains the nuclease activity of the Cas protein. In certain embodiments, the "Cas mutant" or "Cas variant" is mutated such that one or both nuclease domains are inactive. In certain embodiments, the "Cas mutant" or "Cas variant" has nuclease activity. In certain embodiments, the "Cas mutant" or "Cas variant" lacks some or all of the nuclease activity of its wild-type counterpart.

The term "nuclease domain" of a Cas protein refers to the polypeptide sequence or domain within the protein which possesses the catalytic activity for DNA cleavage. A nuclease domain can be contained in a single polypeptide chain, or cleavage activity can result from the association of two (or more) polypeptides. A single nuclease domain may consist of more than one isolated stretch of amino acids within a given polypeptide. Examples of these domains include RuvC-like motifs (amino acids 7-22, 759-766 and 982-989 in SEQ ID NO: 1) and HNH motif (aa 837-863). See Gasiunas et al. (2012) *Proc. Natl. Acad. Sci. USA*, 109:39, E2579-E2586 and WO2013176772.

A synthetic guide RNA that has "gRNA functionality" is one that has one or more of the functions of naturally occurring guide RNA, such as associating with a Cas protein, or a function performed by the guide RNA in association with a Cas protein. In certain embodiments, the functionality includes binding a target polynucleotide. In certain embodiments, the functionality includes targeting a Cas protein or a gRNA:Cas protein complex to a target polynucleotide. In certain embodiments, the functionality includes nicking a target polynucleotide. In certain embodiments, the functionality includes cleaving a target polynucleotide. In certain embodiments, the functionality includes cleaving a target polynucleotide. In certain embodiments, the functionality

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includes associating with or binding to a Cas protein. In certain embodiments, the functionality is any other known function of a guide RNA in a CRISPR-Cas system with a Cas protein, including an artificial CRISPR-Cas system with an engineered Cas protein. In certain embodiments, the 5 functionality is any other function of natural guide RNA. The synthetic guide RNA may have gRNA functionality to a greater or lesser extent than a naturally occurring guide RNA. In certain embodiments, a synthetic guide RNA may have greater functionality as to one property and lesser 10 functionality as to another property in comparison to a similar naturally occurring guide RNA.

A "Cas protein having a single-strand nicking activity" refers to a Cas protein, including a Cas mutant or Cas variant, that has reduced ability to cleave one of two strands 15 of a dsDNA as compared to a wild type Cas protein. For example, in certain embodiments, a Cas protein having a single-strand nicking activity has a mutation (e.g., amino acid substitution) that reduces the function of the RuvC domain (or the HNH domain) and as a result reduces the 20 ability to cleave one strand of the target DNA. Examples of such variants include the D10A, H839A/H840A, and/or N863A substitutions in *S. pyogenes* Cas9, and also include the same or similar substitutions at equivalent sites in Cas9 enzymes of other species.

As used herein, the term "portion" or "fragment" of a sequence refers to any portion of the sequence (e.g., a nucleotide subsequence or an amino acid subsequence) that is smaller than the complete sequence. Portions of polynucleotides can be any length, for example, at least 5, 10, 15, 30 20, 25, 30, 40, 50, 75, 100, 150, 200, 300 or 500 or more nucleotides in length. A portion of a guide sequence can be about 50%, 40%, 30%, 20%, 10% of the guide sequence, e.g., one-third of the guide sequence or shorter, e.g., 7, 6, 5, 4, 3, or 2 nucleotides in length.

The term "derived from" in the context of a molecule refers to a molecule isolated or made using a parent molecule or information from that parent molecule. For example, a Cas9 single mutant nickase and a Cas9 double mutant null-nuclease are derived from a wild-type Cas9 40 protein.

The term "substantially identical" in the context of two or more polynucleotides (or two or more polypeptides) refers to sequences or subsequences that have at least about 60%, at least about 70%, at least about 80%, at least about 90%, 45 about 90-95%, at least about 95%, at least about 98%, at least about 99% or more nucleotide (or amino acid) sequence identity, when compared and aligned for maximum correspondence using a sequence comparison algorithm or by visual inspection. Preferably, the "substantial 50 identity" between polynucleotides exists over a region of the polynucleotide at least about 50 nucleotides in length, at least about 100 nucleotides in length, at least about 200 nucleotides in length, at least about 300 nucleotides in length, at least about 500 nucleotides in length, or over the 55 entire length of the polynucleotide. Preferably, the "substantial identity" between polypeptides exists over a region of the polypeptide at least about 50 amino acid residues in length, at least about 100 amino acid residues in length, or over the entire length of the polypeptide.

As disclosed herein, a number of ranges of values are provided. It is understood that each intervening value, to the tenth of the unit of the lower limit, between the upper and lower limits of that range is also specifically contemplated. Each smaller range or intervening value encompassed by a 65 stated range is also specifically contemplated. The term "about" generally refers to plus or minus 10% of the

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indicated number. For example, "about 10%" may indicate a range of 9% to 11%, and "about 20" may mean from 18-22. Other meanings of "about" may be apparent from the context, such as rounding off, so, for example "about 1" may also mean from 0.5 to 1.4.

# II. CRISPR-Mediated Sequence-Specific Binding and/or Cleavage

Shown in FIG. 1 is a diagram of CRISPR-Cas9-mediated sequence-specific cleavage of DNA. The guide RNA is depicted as sgRNA with an exemplary 20-nucleotide (20-nt) guide sequence (other guide sequences may be, for example, from about 15 to about 30 nts in length) within the 5' domain, an internally positioned base-paired stem, and a 3' domain. The guide sequence is complementary to an exemplary 20-nt target sequence in a DNA target. The stem corresponds to a repeat sequence in crRNA and is complementary to a sequence in the tracrRNA. The 3' domain of the guide RNA corresponds to the 3' domain of the tracrRNA that binds a Cas9 nuclease. The Cas9:guide RNA complex binds and cleaves a target DNA sequence or protospacer directly upstream of a PAM sequence recognized by Cas9. In FIG. 1, a 3-nt PAM sequence is exemplified; however other PAM sequences, including 4-nt and 5-nt PAM sequences are known. In the system exemplified in FIG. 1, both strands of the target sequence in DNA are cleaved by Cas9 at the sites indicated by arrows.

## III. Guide RNAs

In at least one aspect, the present invention comprises a chemically modified guide RNA that has guide RNA functionality. A guide RNA that comprises any nucleotide other than the four canonical ribonucleotides, namely A, C, G, and U, whether unnatural or natural (e.g., a pseudouridine, inosine or a deoxynucleotide), is a chemically modified guide RNA. Likewise a guide RNA that comprises any backbone or internucleotide linkage other than a natural phosphodiester internucleotide linkage possesses a chemical modification and therefore is a chemically modified guide RNA. In certain embodiments, the retained functionality includes binding a Cas protein. In certain embodiments, the retained functionality includes binding a target polynucleotide. In certain embodiments, the retained functionality includes targeting a Cas protein or a gRNA:Cas protein complex to a target polynucleotide. In certain embodiments, the retained functionality includes nicking a target polynucleotide by a gRNA:Cas protein complex. In certain embodiments, the retained functionality includes cleaving a target polynucleotide by a gRNA:Cas protein complex. In certain embodiments, the retained functionality is any other known function of a guide RNA in a CRISPR-Cas system with a Cas protein, including an artificial CRISPR-Cas system with an engineered Cas protein. In certain embodiments, the retained functionality is any other function of a natural guide RNA.

# A. Exemplary Modifications

In certain embodiments, a nucleotide sugar modification incorporated into the guide RNA is selected from the group consisting of 2'-O—C<sub>1-4</sub>alkyl such as 2'-O-methyl (2'-OMe), 2'-deoxy (2'-H), 2'-O—C<sub>1-3</sub>alkyl-O—C<sub>1-3</sub>alkyl such as 2'-methoxyethyl ("2'-MOE"), 2'-fluoro ("2'-F"), 2'-amino ("2'-NH<sub>2</sub>"), 2'-arabinosyl ("2'-arabino") nucleotide, 2'-Farabinosyl ("2'-F-arabino") nucleotide, 2'-locked nucleic

acid ("LNA") nucleotide, 2'-unlocked nucleic acid ("ULNA") nucleotide, a sugar in L form ("L-sugar"), and 4'-thioribosyl nucleotide. In certain embodiments, an internucleotide linkage modification incorporated into the guide RNA is selected from the group consisting of: phosphorothioate "P(S)" (P(S)), phosphonocarboxylate (P(CH $_2$ ), COOR) such as phosphonoacetate "PACE" (P(CH $_2$ COO")), thiophosphonoacetate "thioPACE" ((S)P(CH $_2$ COO")), alkylphosphonate (P(C $_{1-3}$ alkyl) such as methylphosphonate —P(CH $_3$ ), boranophosphonate (P(BH $_3$ )), and phosphorodithioate (P(S) $_2$ ).

In certain embodiments, a nucleobase ("base") modification incorporated into the guide RNA is selected from the 15 group consisting of: 2-thiouracil ("2-thioU"), 2-thiocytosine ("2-thioC"), 4-thiouracil ("4-thioU"), 6-thioguanine ("6thioG"), 2-aminoadenine ("2-aminoA"), 2-aminopurine, pseudouracil, hypoxanthine, 7-deazaguanine, 7-deaza-8-azaguanine, 7-deazaadenine, 7-deaza-8-azaadenine, 5-methyl- 20 cytosine ("5-methylC"), 5-methyluracil ("5-methylU"), 5-hydroxymethylcytosine, 5-hydroxymethyluracil, 5,6-dehydrouracil, 5-propynylcytosine, 5-propynyluracil, 5-ethynyleytosine, 5-ethynyluracil, 5-allyluracil ("5-allylU"), 5-al-("5-allylC"), 5-aminoallyluracil lylcytosine aminoallylU"), 5-aminoallyl-cytosine ("5-aminoallylC"), an abasic nucleotide, Z base, P base, Unstructured Nucleic Acid ("UNA"), isoguanine ("isoG"), isocytosine ("isoC") [as described in "Enzymatic Incorporation of a New Base pair 30 into DNA and RNA Extends the Genetic Alphabet." Piccirilli, J. A.; Krauch, T.; Moroney, S. E.; Benner, S. A. (1990) Nature, 343, 33], 5-methyl-2-pyrimidine [as described in Rappaport, H. P. (1993) *Biochemistry*, 32, 3047], x(A,G,C, T) and y(A,G,C,T).

In certain embodiments, one or more isotopic modifications are introduced on the nucleotide sugar, the nucleobase, the phosphodiester linkage and/or the nucleotide phosphates. Such modifications include nucleotides comprising one or more <sup>15</sup>N, <sup>13</sup>C, <sup>14</sup>C, Deuterium, <sup>3</sup>H, <sup>32</sup>P, <sup>125</sup>I, <sup>131</sup>I atoms or other atoms or elements used as tracers.

In certain embodiments, an "end" modification incorporated into the guide RNA is selected from the group consisting of: PEG (polyethyleneglycol), hydrocarbon linkers 45 (including: heteroatom (O,S,N)-substituted hydrocarbon spacers; halo-substituted hydrocarbon spacers; keto-, carboxyl-, amido-, thionyl-, carbamoyl-, thionocarbamaoylcontaining hydrocarbon spacers), spermine linkers, dyes including fluorescent dyes (for example fluoresceins, rhodamines, cyanines) attached to linkers such as for example 6-fluorescein-hexyl, quenchers (for example dabcyl, BHQ) and other labels (for example biotin, digoxigenin, acridine, streptavidin, avidin, peptides and/or proteins). In certain embodiments, an "end" modification comprises a conjugation (or ligation) of the guide RNA to another molecule comprising an oligonucleotide (comprising deoxynucleotides and/or ribonucleotides), a peptide, a protein, a sugar, an oligosaccharide, a steroid, a lipid, a folic acid, a vitamin and/or other molecule. In certain embodiments, an "end" modification incorporated into the guide RNA is located internally in the guide RNA sequence via a linker such as for example 2-(4-butylamidofluorescein)propane-1,3-diol bis (phosphodiester) linker (depicted below), which is incorpo- 65 rated as a phosphodiester linkage and can be incorporated anywhere between two nucleotides in the guide RNA.

2-(4-butylamidofluorescein)propane-1,3-diol bis(phosphodiester) linker

Other linkers include for example by way of illustration, but are not limited to:

2-(3-(dye-amido)propanamido)propane-1,3-diol bis (phosphodiester) linker

In certain embodiments, the end modification comprises a terminal functional group such as an amine, a thiol (or sulfhydryl), a hydroxyl, a carboxyl, carbonyl, thionyl, thiocarbonyl, a carbamoyl, a thiocarbamoyl, a phoshoryl, an alkene, an alkyne, an halogen or a functional group-terminated linker, either of which can be subsequently conjugated to a desired moiety, for example a fluorescent dye or a non-fluorescent label or tag or any other molecule such as for example an oligonucleotide (comprising deoxynucleotides and/or ribonucleotides, including an aptamer), an amino acid, a peptide, a protein, a sugar, an oligosaccharide, a steroid, a lipid, a folic acid, a vitamin. The conjugation employs standard chemistry well-known in the art, including but not limited to coupling via N-hydroxysuccinimide, isothiocyanate, DCC (or DCI), and/or any other standart method as described in "Bioconjugate Techniques" by Greg T. Hermanson, Publisher Eslsevier Science, 3<sup>rd</sup> ed. (2013), the contents of which are incorporated herein by reference in their entireties.

In certain embodiments, the label or dye is attached or conjugated to a modified nucleotide in the gRNA. The conjugation of a fluorescent dye or other moiety such as a non-fluorescent label or tag (for example biotin, avidin, streptavidin, or moiety containing an isotopic label such as <sup>15</sup>N, <sup>13</sup>C, <sup>14</sup>C, Deuterium, <sup>3</sup>H, <sup>32</sup>P, <sup>125</sup>I and the like) or any other molecule such as for example an oligonucleotide (comprising deoxynucleotides and/or ribonucleotides

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including an aptamer), an amino acid, a peptide, a protein, a sugar, an oligosaccharide, a steroid, a lipid, a folic acid, a vitamin or other molecule can be effectuated using the so-called "click" chemistry or the so-called "squarate" conjugation chemistry. The "click" chemistry refers to the [3+2] cycloaddition of an alkyne moiety with an azide moiety, leading to a triazolo linkage between the two moieties as shown in the following scheme:

as described for example in El-Sagheer, A. H. and Brown, T. "Click chemistry with DNA", *Chem. Soc. Rev.*, 2010, 39, 1388-1405 and Mojibul, H. M. and XiaoHua, P., DNA-associated click chemistry, *Sci. China Chem.*, 2014, 57:2, 215-31, the contents of which are hereby incorporated by reference in their entirety.

In certain embodiments, the conjugation can be effectuated by alternative cycloaddition such as Diels-Alder [4+2] cycloaddition of a  $\pi$ -conjugated diene moiety with an alkene moiety.

The "squarate" conjugation chemistry links two moieties each having an amine via a squarate derivative to result in a squarate conjugate that contains a squarate moiety (see e.g., Tietze et al. (1991) *Chem. Ber.*, 124, 1215-21, the contents of which are hereby incorporated by reference in their entirety). For example, a fluorescein containing a linker amine is conjugated to an oligoribonucleotide containing an amine through a squarate linker as described in the scheme below. An example of the squarate linker is depicted in the following scheme:

Fluo 
$$\longrightarrow$$
 NH<sub>2</sub> + 45

R<sub>2</sub>O OR<sub>1</sub> 50

H<sub>2</sub>N Oligo Squarate

NH NH NH NH Oligo Squarate conjugate

In certain embodiments, a chemical modification incorporated into the guide RNA is selected from the group consisting of 2'-O—C<sub>1-4</sub>alkyl, 2'-H, 2'-O—C<sub>1-3</sub>alkyl-O—C<sub>1-3</sub>alkyl, 2'-F, 2'-NH<sub>2</sub>, 2'-arabino, 2'-F-arabin, 4'-thioribosyl, 2-thioU, 2-thioC, 4-thioU, 6-thioG, 2-aminoA, 2-aminopurine, pseudouracil, hypoxanthine, 7-deazaguanine, 7-deaza-8-azaguanine, 7-deaza-8-azaguanine, 7-deaza-8-azaguanine, 7-deaza-8-azaguanine, 7-deaza-8-azaguanine,

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nine, 5-methylC, 5-methylU, 5-hydroxymethylcytosine, 5-hydroxymethyluracil, 5,6-dehydrouracil, 5-propynylcytosine, 5-propynyluracil, 5-ethynylcytosine, 5-ethynyluracil, 5-allylU, 5-allylC, 5-aminoallyl-uracil, 5-aminoallyl-cytosine, an abasic nucleotide ("abN"), Z. P. UNA, isoC, isoG, 5-methyl-pyrimidine, x(A,G,C,T) and v(A,G,C,T), a phosphorothioate internucleotide linkage, a phosphonoacetate internucleotide linkage, a thiophosphonoacetate internucleotide linkage, a methylphosphonate internucleotide linkage, a boranophosphonate internucleotide linkage, a phosphorodithioate internucleotide linkage, 4'-thioribosyl nucleotide, a locked nucleic acid ("LNA") nucleotide, an unlocked nucleic acid ("ULNA") nucleotide, an alkyl spacer, a heteroalkyl (N, O, S) spacer, a 5'- and/or 3'-alkyl terminated nucleotide, a Unicap, a 5'-terminal cap known from nature, an xRNA base (analogous to "xDNA" base), an yRNA base (analogous to "yDNA" base), a PEG substituent, or a conjugated linker to a dye or non-fluorescent label (or tag) or other moiety as described above. Exemplary modified nucleotides are also depicted in Table 2.

# TABLE 2

Exemplary modified nucleotides contained in a synthetic guide sequence.

$$R_{1} = OH \ or \ 2'modificiation$$
 
$$R_{2} = OH \ or \ internucleotide \ linkage$$
 
$$B = base$$
 
$$\# \qquad R_{1} \qquad R_{2} \qquad B$$

.,	D.	n.	D
#	$R_1$	$R_2$	В
$\mathbf{A}1$	OH	OH	uridine
A2	OMe	OH	uridine
A3	F	OH	uridine
A4	Cl	OH	uridine
A5	$\operatorname{Br}$	OH	uridine
A6	I	OH	uridine
A7	$NH_2$	OH	uridine
A8	H	OH	uridine
A9	OH	phosphodiester	uridine
A10	OMe	phosphodiester	uridine
A11	F	phosphodiester	uridine
A12	Cl	phosphodiester	uridine
A13	$\operatorname{Br}$	phosphodiester	uridine
A14	I	phosphodiester	uridine
A15	$NH_2$	phosphodiester	uridine
A16	H	phosphodiester	uridine
A17	OH	phosphonoacetate	uridine
A18	OMe	phosphonoacetate	uridine
A19	F	phosphonoacetate	uridine
A20	Cl	phosphonoacetate	uridine
A21	$_{\mathrm{Br}}$	phosphonoacetate	uridine
A22	I	phosphonoacetate	uridine
A23	$NH_2$	phosphonoacetate	uridine
A24	Η	phosphonoacetate	uridine
A25	OH	thiophosphonoacetate	uridine
A26	OMe	thiophosphonoacetate	uridine
A27	F	thiophosphonoacetate	uridine
A28	Cl	thiophosphonoacetate	uridine
A29	$_{\mathrm{Br}}$	thiophosphonoacetate	uridine
A30	I	thiophosphonoacetate	uridine
A31	$NH_2$	thiophosphonoacetate	uridine
A32	Η	thiophosphonoacetate	uridine
A33	OH	phosphorothioate	uridine
A34	OMe	phosphorothioate	uridine
A35	F	phosphorothioate	uridine
A36	Cl	phosphorothioate	uridine
A37	Br	phosphorothioate	uridine
A38	I	phosphorothioate	uridine
A39	$NH_2$	phosphorothioate	uridine
A40	H	phosphorothioate	uridine

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TABLE 2-continued

# TABLE 2-continued

		TABLE 2-continued					TABLE 2-continued	1
Exempla	ıry modific	ed nucleotides contained in a sy	nthetic guide sequence.		Exempl	ary modifie	ed nucleotides contained in a s	ynthetic guide sequence.
mumm	$R_2$	$R_2 = OH \text{ or inter}$	2'modificiation mucleotide linkage - base	5	modom	$R_2$	$R_2 = OH \text{ or inte}$	2'modificiation rnucleotide linkage = base
#	$R_1$	$R_1$	В	10	#	$R_1$	$R_1$ $R_2$	В
A41	OH	phosphorodithioate	uridine		B43	F	phosphorodithioate	adenosine
A41 A42	OMe	phosphorodithioate	uridine		B44	Cl	phosphorodithioate	adenosine
A43	F	phosphorodithioate	uridine	15	B45	Br	phosphorodithioate	adenosine
A44 A45	Cl Br	phosphorodithioate phosphorodithioate	uridine uridine		B46 B47	$_{\mathrm{NH_{2}}}^{\mathrm{I}}$	phosphorodithioate phosphorodithioate	adenosine adenosine
A46	I	phosphorodithioate	uridine		B48	H	phosphorodithioate	adenosine
A47	$NH_2$	phosphorodithioate	uridine		B49	OH	methylphosphonate	adenosine
A48	Н	phosphorodithioate	uridine		B50	OMe	methylphosphonate	adenosine
A49 A50	OH OMe	methylphosphonate methylphosphonate	uridine uridine	20	B51 B52	F Cl	methylphosphonate methylphosphonate	adenosine adenosine
A50 A51	F	methylphosphonate	uridine		B52	Br	methylphosphonate	adenosine
A52	Cl	methylphosphonate	uridine		B54	I	methylphosphonate	adenosine
A53	$_{\mathrm{Br}}$	methylphosphonate	uridine		B55	$NH_2$	methylphosphonate	adenosine
A54	I	methylphosphonate	uridine		B56	Н	methylphosphonate	adenosine
A55 A56	$_{ m H}^{ m NH_2}$	methylphosphonate methylphosphonate	uridine uridine	25	B57 B58	OH OMe	boranophosphonate boranophosphonate	adenosine adenosine
A57	ОН	boranophosphonate	uridine		B59	F	boranophosphonate	adenosine
A58	OMe	boranophosphonate	uridine		<b>B</b> 60	Cl	boranophosphonate	adenosine
A59 A60	F	boranophosphonate	uridine		B61 B62	$_{ m I}^{ m Br}$	boranophosphonate	adenosine
A60 A61	Cl Br	boranophosphonate boranophosphonate	uridine uridine		B62	$^{1}_{ m NH_{2}}$	boranophosphonate boranophosphonate	adenosine adenosine
A62	I	boranophosphonate	uridine	30	B64	H	boranophosphonate	adenosine
A63	$NH_2$	boranophosphonate	uridine		C1	OH	ОН	cytidine
A64	Н	boranophosphonate	uridine		C2	OMe F	OH	cytidine
B1 B2	OH OMe	OH OH	adenosine adenosine		C3 C4	r Cl	OH OH	cytidine cytidine
B3	F	OH	adenosine		C5	Br	OH	cytidine
B4	Cl	ОН	adenosine	35	C6	I	ОН	cytidine
B5 B6	Br I	OH OH	adenosine adenosine		C7 C8	$^{ m NH_2}_{ m H}$	OH OH	cytidine cytidine
B7	$^{1}_{ m NH_{2}}$	OH	adenosine		C9	OH	phosphodiester	cytidine
В8	H	OH	adenosine		C10	OMe	phosphodiester	cytidine
В9	OH	phosphodiester	adenosine		C11	F	phosphodiester	cytidine
B10 B11	OMe F	phosphodiester phosphodiester	adenosine adenosine	40	C12 C13	Cl Br	phosphodiester phosphodiester	cytidine cytidine
B12	Cl	phosphodiester	adenosine		C14	I	phosphodiester	cytidine
B13	$\operatorname{Br}$	phosphodiester	adenosine		C15	$NH_2$	phosphodiester	cytidine
B14	I	phosphodiester	adenosine		C16	Н	phosphodiester	cytidine
B15 B16	$_{ m H}^{ m NH_2}$	phosphodiester phosphodiester	adenosine adenosine		C17 C18	OH OMe	phosphonoacetate phosphonoacetate	cytidine cytidine
B17	OH	phosphonoacetate	adenosine	45	C19	F	phosphonoacetate	cytidine
B18	OMe	phosphonoacetate	adenosine		C20	Cl	phosphonoacetate	cytidine
B19 B20	F Cl	phosphonoacetate phosphonoacetate	adenosine adenosine		C21 C22	Br I	phosphonoacetate phosphonoacetate	cytidine cytidine
B21	Br	phosphonoacetate	adenosine		C23	$^{1}_{ m NH_{2}}$	phosphonoacetate	cytidine
B22	I	phosphonoacetate	adenosine		C24	Н	phosphonoacetate	cytidine
B23	$NH_2$	phosphonoacetate	adenosine	50	C25	OH	thiophosphonoacetate	cytidine
B24 B25	H OH	phosphonoacetate thiophosphonoacetate	adenosine adenosine		C26 C27	OMe F	thiophosphonoacetate thiophosphonoacetate	cytidine cytidine
B26	OMe	thiophosphonoacetate	adenosine		C28	Cl	thiophosphonoacetate	cytidine
B27	F	thiophosphonoacetate	adenosine		C29	$_{\mathrm{Br}}$	thiophosphonoacetate	cytidine
B28	Cl	thiophosphonoacetate	adenosine		C30	I	thiophosphonoacetate	cytidine
B29 B30	Br I	thiophosphonoacetate thiophosphonoacetate	adenosine adenosine	55	C31 C32	$_{ m H}^{ m NH_2}$	thiophosphonoacetate thiophosphonoacetate	cytidine cytidine
B31	$NH_2$	thiophosphonoacetate	adenosine		C33	ОН	phosphorothioate	cytidine
B32	H	thiophosphonoacetate	adenosine		C34	OMe	phosphorothioate	cytidine
B33	ОН	phosphorothioate	adenosine		C35	F	phosphorothioate	cytidine
B34	OMe	phosphorothicate	adenosine	<b>CO</b>	C36	Cl	phosphorothioate	cytidine
B35 B36	F Cl	phosphorothioate phosphorothioate	adenosine adenosine	60	C37 C38	Br I	phosphorothioate phosphorothioate	cytidine cytidine
B37	Br	phosphorothioate	adenosine		C39	$^{1}$ NH $_{2}$	phosphorothioate	cytidine
B38	I	phosphorothioate	adenosine		C40	H	phosphorothioate	cytidine
B39	$NH_2$	phosphorothioate	adenosine		C41	ОН	phosphorodithioate	cytidine
B40	Н	phosphorothioate	adenosine	65	C42	OMe	phosphorodithioate	cytidine
B41 B42	OH OMe	phosphorodithioate phosphorodithioate	adenosine adenosine	65	C43 C44	F Cl	phosphorodithioate phosphorodithioate	cytidine cytidine
1542	Olvie	phosphorouninoate	agenosine		C44	Ci	phosphorourinoate	cyname

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TABLE 2-continues

# TABLE 2-continued

		TABLE 2-continued					TABLE 2-continued	1
Exempla	ry modifie	ed nucleotides contained in a synthe	etic guide sequence.		Exemple	ary modifie	ynthetic guide sequence.	
more	\ \	B $R_1 = OH \text{ or } 2'mo$ $R_2 = OH \text{ or internuc}$ $R = bas$	leotide linkage	5	more	\ \{\cdot\}	$R_1 = OH \text{ or}$ $R_2 = OH \text{ or integer}$	2'modificiation emucleotide linkage = base
			C			<i></i>		base
	$R_2'$	$R_1$		10		$R_2$	$R_1$	
#	$R_1$	$R_2$	В		#	$R_1$	$R_2$	В
C45	Br	phosphorodithioate	cytidine		D47	$\mathrm{NH}_2$	phosphorodithioate	guanosine
C46	I	phosphorodithioate	cytidine	1.5	D48 D49	H OH	phosphorodithioate methylphosphonate	guanosine
C47 C48	$_{ m H}^{ m NH_2}$	phosphorodithioate phosphorodithioate	cytidine cytidine	15	D50	OMe	methylphosphonate	guanosine guanosine
C49	ОН	methylphosphonate	cytidine		D51	F	methylphosphonate	guanosine
C50	OMe	methylphosphonate	cytidine		D52	Cl	methylphosphonate	guanosine
C51	F	methylphosphonate	cytidine		D53	$\operatorname{Br}$	methylphosphonate	guanosine
C52 C53	Cl Br	methylphosphonate methylphosphonate	cytidine cytidine	•	D54 D55	$_{\mathrm{NH}_{2}}^{\mathrm{I}}$	methylphosphonate methylphosphonate	guanosine guanosine
C54	I	methylphosphonate	cytidine	20	D56	H	methylphosphonate	guanosine
C55	$NH_2$	methylphosphonate	cytidine		D57	OH	boranophosphonate	guanosine
C56	Н	methylphosphonate	cytidine		D58	OMe	boranophosphonate	guanosine
C57	OH OMe	boranophosphonate boranophosphonate	cytidine		D59 D60	F	boranophosphonate	guanosine
C58 C59	F	boranophosphonate	cytidine cytidine		D60 D61	Cl Br	boranophosphonate boranophosphonate	guanosine guanosine
C60	Cl	boranophosphonate	cytidine	25	D62	I	boranophosphonate	guanosine
C61	$_{\mathrm{Br}}$	boranophosphonate	cytidine		D63	$\mathrm{NH}_2$	boranophosphonate	guanosine
C62	I	boranophosphonate	cytidine		D64	Н	boranophosphonate	guanosine
C63	$NH_2$	boranophosphonate	cytidine cytidine		E1 E2	OH OMe	OH OH	2-thiouridine
C64 D1	H OH	boranophosphonate OH	guanosine		E3	F	OH	2-thiouridine 2-thiouridine
D2	OMe	OH	guanosine	30	E4	Ĉl	OH	2-thiouridine
D3	F	OH	guanosine		E5	$_{\mathrm{Br}}$	ОН	2-thiouridine
D4	Cl	OH	guanosine		E6	I	OH	2-thiouridine
D5	$\operatorname{Br}$	OH	guanosine		E7 E8	$NH_2$	OH	2-thiouridine
D6 D7	$_{ m NH_2}$	OH OH	guanosine guanosine		E8 E9	H OH	OH phosphodiester	2-thiouridine 2-thiouridine
D8	Н	OH	guanosine	35	E10	OMe	phosphodiester	2-thiouridine
D9	OH	phosphodiester	guanosine		E11	F	phosphodiester	2-thiouridine
D10	OMe	phosphodiester	guanosine		E12	Cl	phosphodiester	2-thiouridine
D11 D12	F Cl	phosphodiester phosphodiester	guanosine guanosine		E13 E14	Br I	phosphodiester phosphodiester	2-thiouridine 2-thiouridine
D13	Br	phosphodiester	guanosine		E15	$^{1}_{ m NH_{2}}$	phosphodiester	2-thiouridine
D14	I	phosphodiester	guanosine	40	E16	H	phosphodiester	2-thiouridine
D15	$NH_2$	phosphodiester	guanosine		E17	OH	phosphonoacetate	2-thiouridine
D16 D17	H OH	phosphodiester phosphonoacetate	guanosine guanosine		E18 E19	OMe F	phosphonoacetate phosphonoacetate	2-thiouridine 2-thiouridine
D17	OMe	phosphonoacetate	guanosine		E20	Cl	phosphonoacetate	2-thiouridine
D19	F	phosphonoacetate	guanosine		E21	$\operatorname{Br}$	phosphonoacetate	2-thiouridine
D20	Cl	phosphonoacetate	guanosine	45	E22	I	phosphonoacetate	2-thiouridine
D21	$\operatorname{Br}$	phosphonoacetate	guanosine		E23	$NH_2$	phosphonoacetate	2-thiouridine
D22 D23	$_{ m NH_2}$	phosphonoacetate phosphonoacetate	guanosine guanosine		E24 E25	H OH	phosphonoacetate thiophosphonoacetate	2-thiouridine 2-thiouridine
D24	Н	phosphonoacetate	guanosine		E26	OMe	thiophosphonoacetate	2-thiouridine
D25	OH	thiophosphonoacetate	guanosine		E27	F	thiophosphonoacetate	2-thiouridine
D26	OMe	thiophosphonoacetate	guanosine	50	E28	Cl	thiophosphonoacetate	2-thiouridine
D27 D28	F Cl	thiophosphonoacetate thiophosphonoacetate	guanosine guanosine	30	E29 E30	Br I	thiophosphonoacetate thiophosphonoacetate	2-thiouridine 2-thiouridine
D29	Br	thiophosphonoacetate	guanosine		E31	$^{1}_{ m NH_{2}}$	thiophosphonoacetate	2-thiouridine
D30	I	thiophosphonoacetate	guanosine		E32	H	thiophosphonoacetate	2-thiouridine
D31	$NH_2$	thiophosphonoacetate	guanosine		E33	OH	phosphorothioate	2-thiouridine
D32	Н	thiophosphonoacetate	guanosine		E34	OMe	phosphorothioate	2-thiouridine
D33 D34	OH OMe	phosphorothioate phosphorothioate	guanosine guanosine	55	E35 E36	F Cl	phosphorothioate phosphorothioate	2-thiouridine 2-thiouridine
D35	F	phosphorothioate	guanosine		E37	Br	phosphorothioate	2-thiouridine
D36	Čl	phosphorothioate	guanosine		E38	I	phosphorothioate	2-thiouridine
D37	$_{\mathrm{Br}}$	phosphorothioate	guanosine		E39	$\mathrm{NH}_2$	phosphorothioate	2-thiouridine
D38	I	phosphorothioate	guanosine		E40	Η	phosphorothioate	2-thiouridine
D39	$NH_2$	phosphorothioate	guanosine	60	E41	OH	phosphorodithioate	2-thiouridine
D40	Н	phosphorothioate	guanosine		E42	OMe	phosphorodithioate	2-thiouridine
D41 D42	OH OMe	phosphorodithioate phosphorodithioate	guanosine		E43 E44	F Cl	phosphorodithioate phosphorodithioate	2-thiouridine 2-thiouridine
D42 D43	F F	phosphorodithioate phosphorodithioate	guanosine guanosine		E44 E45	Br	phosphorodithioate phosphorodithioate	2-thiouridine 2-thiouridine
D43	Cl	phosphorodithioate	guanosine		E46	I	phosphorodithioate	2-thiouridine
D45	Br	phosphorodithioate	guanosine	65	E47	$NH_2$	phosphorodithioate	2-thiouridine
D46	I	phosphorodithioate	guanosine		E48	Н	phosphorodithioate	2-thiouridine

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TABLE 2-continued					TABLE 2-continued				
Exemplary modified nucleotides contained in a synthetic guide sequence.					Exemplary modified nucleotides contained in a synthetic guide sequence				
<b>\</b>					ş				
www			2'modificiation	5	monomo			2'modificiation	
,	_	-	ernucleotide linkage = base		,	_		ernucleotide linkage = base	
	$R_2$	$R_1$		10		$R_2$	$R_1$		
#	$R_1$	$R_2$	В		#	$R_1$	$R_2$	В	
E49	ОН	methylphosphonate	2-thiouridine		F51	F	methylphosphonate	4-thiouridine	
E50 E51	OMe F	methylphosphonate methylphosphonate	2-thiouridine 2-thiouridine	1.5	F52 F53	Cl Br	methylphosphonate	4-thiouridine 4-thiouridine	
E51 E52	r Cl	methylphosphonate	2-thiouridine	15	F54	I	methylphosphonate methylphosphonate	4-thiouridine	
E53	Br	methylphosphonate	2-thiouridine		F55	$^{1}_{\mathrm{NH}_{2}}$	methylphosphonate	4-thiouridine	
E54	I	methylphosphonate	2-thiouridine		F56	H	methylphosphonate	4-thiouridine	
E55	$NH_2$	methylphosphonate	2-thiouridine		F57	OH	boranophosphonate	4-thiouridine	
E56	H	methylphosphonate	2-thiouridine		F58	OMe	boranophosphonate	4-thiouridine	
E57	OH	boranophosphonate	2-thiouridine	20	F59	F	boranophosphonate	4-thiouridine	
E58	OMe	boranophosphonate	2-thiouridine	20	F60	Cl	boranophosphonate	4-thiouridine	
E59	F	boranophosphonate	2-thiouridine		F61	$_{\mathrm{Br}}$	boranophosphonate	4-thiouridine	
E60	Cl	boranophosphonate	2-thiouridine		F62	I	boranophosphonate	4-thiouridine	
E61	$_{\mathrm{Br}}$	boranophosphonate	2-thiouridine		F63	$NH_2$	boranophosphonate	4-thiouridine	
E62	I	boranophosphonate	2-thiouridine		F64	Η	boranophosphonate	4-thiouridine	
E63	$NH_2$	boranophosphonate	2-thiouridine	25	G1	OH	OH	2-aminoadenosine	
E64	Н	boranophosphonate	2-thiouridine		G2	OMe	OH	2-aminoadenosine	
F1	OH	OH	4-thiouridine		G3	F	OH	2-aminoadenosine	
F2	OMe F	OH	4-thiouridine		G4	Cl D-	OH	2-aminoadenosine 2-aminoadenosine	
F3 F4	r Cl	OH OH	4-thiouridine 4-thiouridine		G5 G6	Br I	OH OH	2-aminoadenosine 2-aminoadenosine	
F5	Br	OH	4-thiouridine		G7	$^{1}_{ m NH_{2}}$	OH	2-aminoadenosine	
F6	I	OH	4-thiouridine	30	G8	H	OH	2-aminoadenosine	
F7	$NH_2$	OH	4-thiouridine		G9	OH	phosphodiester	2-aminoadenosine	
F8	H	OH	4-thiouridine		G10	OMe	phosphodiester	2-aminoadenosine	
F9	OH	phosphodiester	4-thiouridine		G11	F	phosphodiester	2-aminoadenosine	
F10	OMe	phosphodiester	4-thiouridine		G12	C1	phosphodiester	2-aminoadenosine	
F11	F	phosphodiester	4-thiouridine		G13	$\operatorname{Br}$	phosphodiester	2-aminoadenosine	
F12	Cl	phosphodiester	4-thiouridine	35	G14	I	phosphodiester	2-aminoadenosine	
F13	Br	phosphodiester	4-thiouridine		G15	$NH_2$	phosphodiester	2-aminoadenosine	
F14	I	phosphodiester	4-thiouridine		G16	Н	phosphodiester	2-aminoadenosine	
F15	$NH_2$	phosphodiester	4-thiouridine		G17	OMa	phosphonoacetate	2-aminoadenosine	
F16 F17	H OH	phosphodiester phosphonoacetate	4-thiouridine 4-thiouridine		G18 G19	OMe F	phosphonoacetate phosphonoacetate	2-aminoadenosine 2-aminoadenosine	
F17	ОМе	phosphonoacetate	4-thiouridine		G20	Cl	phosphonoacetate	2-aminoadenosine	
F19	F	phosphonoacetate	4-thiouridine	40	G20	Br	phosphonoacetate	2-aminoadenosine	
F20	Čl	phosphonoacetate	4-thiouridine		G22	I	phosphonoacetate	2-aminoadenosine	
F21	$_{\mathrm{Br}}$	phosphonoacetate	4-thiouridine		G23	$NH_2$	phosphonoacetate	2-aminoadenosine	
F22	I	phosphonoacetate	4-thiouridine		G24	Η	phosphonoacetate	2-aminoadenosine	
F23	$NH_2$	phosphonoacetate	4-thiouridine		G25	OH	thiophosphonoacetate	2-aminoadenosine	
F24	H	phosphonoacetate	4-thiouridine	45	G26	OMe	thiophosphonoacetate	2-aminoadenosine	
F25	OH	thiophosphonoacetate	4-thiouridine	75	G27	F	thiophosphonoacetate	2-aminoadenosine	
F26	OMe	thiophosphonoacetate	4-thiouridine		G28	Cl	thiophosphonoacetate	2-aminoadenosine	
F27	F	thiophosphonoacetate	4-thiouridine		G29	Br	thiophosphonoacetate	2-aminoadenosine	
F28 F29	Cl Br	thiophosphonoacetate thiophosphonoacetate	4-thiouridine 4-thiouridine		G30 G31	I	thiophosphonoacetate	2-aminoadenosine 2-aminoadenosine	
F30	I	thiophosphonoacetate	4-thiouridine		G32	$_{ m H}^{ m NH_2}$	thiophosphonoacetate thiophosphonoacetate	2-aminoadenosine 2-aminoadenosine	
F31	$^{1}_{\mathrm{NH_{2}}}$	thiophosphonoacetate	4-thiouridine	50	G32	OH	phosphorothioate	2-aminoadenosine	
F32	Н	thiophosphonoacetate	4-thiouridine		G34	OMe	phosphorothioate	2-aminoadenosine	
F33	ОН	phosphorothioate	4-thiouridine		G35	F	phosphorothioate	2-aminoadenosine	
F34	OMe	phosphorothioate	4-thiouridine		G36	Cl	phosphorothioate	2-aminoadenosine	
F35	F	phosphorothioate	4-thiouridine		G37	$_{\mathrm{Br}}$	phosphorothioate	2-aminoadenosine	
F36	Cl	phosphorothioate	4-thiouridine		G38	I	phosphorothioate	2-aminoadenosine	
F37	$_{\mathrm{Br}}$	phosphorothioate	4-thiouridine	55	G39	$NH_2$	phosphorothioate	2-aminoadenosine	
F38	I	phosphorothioate	4-thiouridine		G40	H	phosphorothioate	2-aminoadenosine	
F39	$NH_2$	phosphorothioate	4-thiouridine		G41	OH	phosphorodithioate	2-aminoadenosine	
F40	H	phosphorothioate	4-thiouridine		G42	OMe	phosphorodithioate	2-aminoadenosine	
F41	OH	phosphorodithioate	4-thiouridine		G43	F	phosphorodithioate	2-aminoadenosine	
F42	OMe	phosphorodithioate	4-thiouridine		G44	Cl	phosphorodithioate	2-aminoadenosine	
F43	F	phosphorodithioate	4-thiouridine	60	G45	$_{\mathrm{Br}}$	phosphorodithioate	2-aminoadenosine	
F44	Cl	phosphorodithioate	4-thiouridine		G46	I	phosphorodithioate	2-aminoadenosine	
F45	$_{\mathrm{Br}}$	phosphorodithioate	4-thiouridine		G47	$\mathrm{NH}_2$	phosphorodithioate	2-aminoadenosine	
F46	I	phosphorodithioate	4-thiouridine		G48	H	phosphorodithioate	2-aminoadenosine	
F47	$NH_2$	phosphorodithioate	4-thiouridine		G49	OH	methylphosphonate	2-aminoadenosine	
F48	Η	phosphorodithioate	4-thiouridine		G50	OMe	methylphosphonate	2-aminoadenosine	
F49	OH	methylphosphonate	4-thiouridine	65	G51	F	methylphosphonate	2-aminoadenosine	
F50	OMe	methylphosphonate	4-thiouridine		G52	Cl	methylphosphonate	2-aminoadenosine	

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TABLE 2-continued

# TABLE 2-continued

Exemplary modified nucleotides contained in a synthetic guide sequence.			•	Exemplary modified nucleotides contained in a synthetic guide sequence				
www		B = OHo		5	mon	<u> </u>	B = OH c	al was a
ξ		$R_2 = OH \text{ or int}$	r 2'modificiation ternucleotide linkage t = base		ξ	$\sum$	$R_1 = OH \text{ or } R_2 = OH \text{ or int}$	: 2'modificiation ernucleotide linkage = base
	$R_2$	$R_1$		10		$R_2$	$R_1$	
#	$R_1$	$R_2$	В		#	R <sub>1</sub>	$R_2$	В
G53 G54	Br I	methylphosphonate methylphosphonate	2-aminoadenosine 2-aminoadenosine		H55 H56	$_{\rm H}^{\rm NH_2}$	methylphosphonate methylphosphonate	7-deazaguanosine 7-deazaguanosine
G55	$NH_2$	methylphosphonate	2-aminoadenosine	15	H57	OH	boranophosphonate	7-deazaguanosine
G56	H	methylphosphonate	2-aminoadenosine		H58	OMe	boranophosphonate	7-deazaguanosine
G57	ОН	boranophosphonate	2-aminoadenosine		H59	F	boranophosphonate	7-deazaguanosine
G58	OMe	boranophosphonate	2-aminoadenosine		H60	Cl	boranophosphonate	7-deazaguanosine
G59 G60	F Cl	boranophosphonate boranophosphonate	2-aminoadenosine 2-aminoadenosine		H61 H62	Br I	boranophosphonate boranophosphonate	7-deazaguanosine 7-deazaguanosine
G61	Br	boranophosphonate	2-aminoadenosine	20	H63	$^{1}_{\mathrm{NH_{2}}}$	boranophosphonate	7-deazaguanosine
G62	I	boranophosphonate	2-aminoadenosine	20	H64	H	boranophosphonate	7-deazaguanosine
G63	$NH_2$	boranophosphonate	2-aminoadenosine		I1	OH	OH	inosine
G64	H	boranophosphonate	2-aminoadenosine		I2	OMe	OH	inosine
H1	OH	OH	7-deazaguanosine		I3	F	OH	inosine
H2	OMe	OH	7-deazaguanosine		I4	Cl	OH	inosine
H3	F	OH	7-deazaguanosine	25	I5	$\operatorname{Br}$	OH	inosine
H4 H5	Cl Br	OH OH	7-deazaguanosine 7-deazaguanosine		I6 I7	$_{ m NH_2}$	OH OH	inosine inosine
H6	I	OH	7-deazaguanosine 7-deazaguanosine		I8	H H	OH	inosine
H7	$_{ m NH_2}$	OH	7-deazaguanosine		I9	OH	phosphodiester	inosine
H8	H	OH	7-deazaguanosine		I10	OMe	phosphodiester	inosine
H9	OH	phosphodiester	7-deazaguanosine		I11	F	phosphodiester	inosine
H10	OMe	phosphodiester	7-deazaguanosine	30	I12	Cl	phosphodiester	inosine
H11	F	phosphodiester	7-deazaguanosine		I13	Br	phosphodiester	inosine
H12	Cl	phosphodiester	7-deazaguanosine		I14	I	phosphodiester	inosine
H13 H14	Br I	phosphodiester phosphodiester	7-deazaguanosine 7-deazaguanosine		I15 I16	$_{ m H}^{ m NH_2}$	phosphodiester phosphodiester	inosine inosine
H15	$^{1}_{ m NH_{2}}$	phosphodiester	7-deazaguanosine 7-deazaguanosine		I10 I17	ОН	phosphonoacetate	inosine
H16	Н	phosphodiester	7-deazaguanosine	35	I18	OMe	phosphonoacetate	inosine
H17	OH	phosphonoacetate	7-deazaguanosine		I19	F	phosphonoacetate	inosine
H18	OMe	phosphonoacetate	7-deazaguanosine		I20	Cl	phosphonoacetate	inosine
H19	F	phosphonoacetate	7-deazaguanosine		I21	Br	phosphonoacetate	inosine
H20	Cl	phosphonoacetate	7-deazaguanosine		I22	I	phosphonoacetate	inosine
H21 H22	Br I	phosphonoacetate phosphonoacetate	7-deazaguanosine 7-deazaguanosine		I23 I24	$_{ m H}^{ m NH_2}$	phosphonoacetate phosphonoacetate	inosine inosine
H23	$^{1}_{\mathrm{NH}_{2}}$	phosphonoacetate	7-deazaguanosine 7-deazaguanosine	40	I24 I25	ОН	thiophosphonoacetate	inosine
H24	Н	phosphonoacetate	7-deazaguanosine		I26	OMe	thiophosphonoacetate	inosine
H25	OH	thiophosphonoacetate	7-deazaguanosine		I27	F	thiophosphonoacetate	inosine
H26	OMe	thiophosphonoacetate	7-deazaguanosine		I28	Cl	thiophosphonoacetate	inosine
H27	F	thiophosphonoacetate	7-deazaguanosine		I29	Br	thiophosphonoacetate	inosine
H28	Cl	thiophosphonoacetate	7-deazaguanosine	45	I30	I	thiophosphonoacetate	inosine
H29 H30	Br I	thiophosphonoacetate thiophosphonoacetate	7-deazaguanosine 7-deazaguanosine		I31 I32	$_{ m H}^{ m NH_2}$	thiophosphonoacetate thiophosphonoacetate	inosine inosine
H31	$^{1}_{ m NH_{2}}$	thiophosphonoacetate	7-deazaguanosine 7-deazaguanosine		I33	OH	phosphorothioate	inosine
H32	Н	thiophosphonoacetate	7-deazaguanosine		I34	OMe	phosphorothioate	inosine
H33	OH	phosphorothioate	7-deazaguanosine		I35	F	phosphorothioate	inosine
H34	OMe	phosphorothioate	7-deazaguanosine	50	I36	Cl	phosphorothioate	inosine
H35	F	phosphorothioate	7-deazaguanosine	50	I37	$\operatorname{Br}$	phosphorothioate	inosine
H36	Cl Pr	phosphorothioate	7-deazaguanosine		I38	I	phosphorothioate	inosine inosine
H37 H38	Br I	phosphorothioate phosphorothioate	7-deazaguanosine 7-deazaguanosine		I39 I40	$_{ m H}^{ m NH_2}$	phosphorothioate phosphorothioate	inosine
H39	$NH_2$	phosphorothioate	7-deazaguanosine		I41	OH	phosphorodithioate	inosine
H40	H	phosphorothioate	7-deazaguanosine		I42	OMe	phosphorodithioate	inosine
H41	OH	phosphorodithioate	7-deazaguanosine	55	I43	F	phosphorodithioate	inosine
H42	OMe	phosphorodithioate	7-deazaguanosine		I44	C1	phosphorodithioate	inosine
H43	F	phosphorodithioate	7-deazaguanosine		I45	Br	phosphorodithioate	inosine
H44	Cl	phosphorodithioate	7-deazaguanosine		I46	I	phosphorodithioate	inosine
H45	Br	phosphorodithioate	7-deazaguanosine		I47	$NH_2$	phosphorodithioate	inosine
H46	I	phosphorodithioate	7-deazaguanosine		I48	H	phosphorodithioate	inosine
H47	$NH_2$	phosphorodithioate	7-deazaguanosine	60	I49	OH	methylphosphonate	inosine
H48	Н	phosphorodithioate	7-deazaguanosine		I50	OMe	methylphosphonate	inosine
H49	OH OMe	methylphosphonate	7-deazaguanosine		I51 I52	F Cl	methylphosphonate	inosine
H50 H51	OMe F	methylphosphonate methylphosphonate	7-deazaguanosine 7-deazaguanosine		152 153	Ci Br	methylphosphonate methylphosphonate	inosine inosine
H51 H52	r Cl	methylphosphonate	7-deazaguanosine 7-deazaguanosine		153 154	I I	methylphosphonate	inosine
H53	Br	methylphosphonate	7-deazaguanosine 7-deazaguanosine	65	I55	$^{1}$ $^{1}$	methylphosphonate	inosine
H54	I	methylphosphonate	7-deazaguanosine 7-deazaguanosine		I56	ИП <sub>2</sub> Н	methylphosphonate	inosine
	-		. Grazaguaroome		200	**		

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TABLE 2-continued

# 22 TABLE 2-continued

TABLE 2-continued					TABLE 2-continued			
Exemplary modified nucleotides contained in a synthetic guide sequence.					Exemplary modified nucleotides contained in a synthetic guide sequence			
www	_\	~	2'modificiation	5	modowo	_\		r 2'modificiation ternucleotide linkage
	\_	, –	= base			\_	-	B = base
	$R_2$	$R_1$		10		$R_2$	$R_1$	
#	$R_1$	$R_2$	В		#	$R_1$	$R_2$	В
157	ОН	boranophosphonate	inosine		J59	F	boranophosphonate	5-methylcytidine
I58 I59	OMe F	boranophosphonate boranophosphonate	inosine inosine	15	J60 J61	Cl Br	boranophosphonate boranophosphonate	5-methylcytidine 5-methylcytidine
I60	Cl	boranophosphonate	inosine	13	J62	I	boranophosphonate	5-methylcytidine
I61	Br	boranophosphonate	inosine		J63	$NH_2$	boranophosphonate	5-methylcytidine
162	I	boranophosphonate	inosine		J64	H	boranophosphonate	5-methylcytidine
I63	$NH_2$	boranophosphonate	inosine		K1	OH	ОН	5-aminoallyluridine
I64	Н	boranophosphonate	inosine		K2	OMe	OH	5-aminoallyluridine
J1 J2	OMa	OH	5-methylcytidine	20	K3 K4	F Cl	OH OH	5-aminoallyluridine
J2 J3	OMe F	OH OH	5-methylcytidine 5-methylcytidine		K5	Br	OH	5-aminoallyluridine 5-aminoallyluridine
J4	Cl	OH	5-methylcytidine		K6	I	OH	5-aminoallyluridine
J5	Br	OH	5-methylcytidine		K7	$NH_2$	OH	5-aminoallyluridine
J6	I	OH	5-methylcytidine		K8	Η	OH	5-aminoallyluridine
J7	$NH_2$	OH	5-methylcytidine	25	K9	OH	phosphodiester	5-aminoallyluridine
J8	Н	OH	5-methylcytidine	23	K10	OMe	phosphodiester	5-aminoallyluridine
J9	OM	phosphodiester	5-methylcytidine 5-methylcytidine		K11	F	phosphodiester	5-aminoallyluridine
J10 J11	OMe F	phosphodiester phosphodiester	5-methylcytidine		K12 K13	Cl Br	phosphodiester phosphodiester	5-aminoallyluridine 5-aminoallyluridine
J12	Čl	phosphodiester	5-methylcytidine		K14	I	phosphodiester	5-aminoallyluridine
J13	Br	phosphodiester	5-methylcytidine		K15	$NH_2$	phosphodiester	5-aminoallyluridine
J14	I	phosphodiester	5-methylcytidine	30	K16	H	phosphodiester	5-aminoallyluridine
J15	$NH_2$	phosphodiester	5-methylcytidine		K17	OH	phosphonoacetate	5-aminoallyluridine
J16	Н	phosphodiester	5-methylcytidine		K18	OMe	phosphonoacetate	5-aminoallyluridine
J17 J18	OH OMe	phosphonoacetate	5-methylcytidine		K19 K20	F	phosphonoacetate	5-aminoallyluridine
J19	F	phosphonoacetate phosphonoacetate	5-methylcytidine 5-methylcytidine		K20 K21	Cl Br	phosphonoacetate phosphonoacetate	5-aminoallyluridine 5-aminoallyluridine
J20	Ĉl	phosphonoacetate	5-methylcytidine	35	K22	I	phosphonoacetate	5-aminoallyluridine
J21	$_{\mathrm{Br}}$	phosphonoacetate	5-methylcytidine		K23	$NH_2$	phosphonoacetate	5-aminoallyluridine
J22	I	phosphonoacetate	5-methylcytidine		K24	H	phosphonoacetate	5-aminoallyluridine
J23	$NH_2$	phosphonoacetate	5-methylcytidine		K25	OH	thiophosphonoacetate	5-aminoallyluridine
J24	Н	phosphonoacetate	5-methylcytidine		K26	OMe	thiophosphonoacetate	5-aminoallyluridine
J25 J26	OH OMe	thiophosphonoacetate thiophosphonoacetate	5-methylcytidine 5-methylcytidine		K27 K28	F Cl	thiophosphonoacetate thiophosphonoacetate	5-aminoallyluridine 5-aminoallyluridine
J27	F	thiophosphonoacetate	5-methylcytidine	40	K29	Br	thiophosphonoacetate	5-aminoallyluridine
J28	Čl	thiophosphonoacetate	5-methylcytidine		K30	I	thiophosphonoacetate	5-aminoallyluridine
J29	$_{\mathrm{Br}}$	thiophosphonoacetate	5-methylcytidine		K31	$NH_2$	thiophosphonoacetate	5-aminoallyluridine
J30	I	thiophosphonoacetate	5-methylcytidine		K32	H	thiophosphonoacetate	5-aminoallyluridine
J31	$NH_2$	thiophosphonoacetate	5-methylcytidine		K33	OH	phosphorothioate	5-aminoallyluridine
J32 J33	H OH	thiophosphonoacetate phosphorothioate	5-methylcytidine 5-methylcytidine	45	K34 K35	OMe F	phosphorothioate phosphorothioate	5-aminoallyluridine 5-aminoallyluridine
J34	OMe	phosphorothioate	5-methylcytidine		K36	Cl	phosphorothioate	5-aminoallyluridine
J35	F	phosphorothioate	5-methylcytidine		K37	Br	phosphorothioate	5-aminoallyluridine
J36	Cl	phosphorothioate	5-methylcytidine		K38	I	phosphorothioate	5-aminoallyluridine
J37	$_{\mathrm{Br}}$	phosphorothioate	5-methylcytidine		K39	$\mathrm{NH}_2$	phosphorothioate	5-aminoallyluridine
J38	I	phosphorothioate	5-methylcytidine	50	K40	H	phosphorothioate	5-aminoallyluridine
J39	$NH_2$	phosphorothioate	5-methylcytidine	30	K41	OM	phosphorodithioate	5-aminoallyluridine
J40 J41	H OH	phosphorothioate phosphorodithioate	5-methylcytidine 5-methylcytidine		K42 K43	OMe F	phosphorodithioate phosphorodithioate	5-aminoallyluridine 5-aminoallyluridine
J42	OMe	phosphorodithioate	5-methylcytidine		K44	Cl	phosphorodithioate	5-aminoallyluridine
J43	F	phosphorodithioate	5-methylcytidine		K45	Br	phosphorodithioate	5-aminoallyluridine
J44	Cl	phosphorodithioate	5-methylcytidine		K46	I	phosphorodithioate	5-aminoallyluridine
J45	$_{\mathrm{Br}}$	phosphorodithioate	5-methylcytidine	55	K47	$NH_2$	phosphorodithioate	5-aminoallyluridine
J46	I	phosphorodithioate	5-methylcytidine		K48	H	phosphorodithioate	5-aminoallyluridine
J47	$NH_2$	phosphorodithioate	5-methylcytidine		K49	OH	methylphosphonate	5-aminoallyluridine
J48	Н	phosphorodithioate	5-methylcytidine		K50	OMe	methylphosphonate	5-aminoallyluridine
J49	OH OMe	methylphosphonate	5-methylcytidine		K51	F	methylphosphonate	5-aminoallyluridine
J50 I51	OMe F	methylphosphonate	5-methylcytidine	60	K52	Cl Br	methylphosphonate	5-aminoallyluridine
J51 J52	F Cl	methylphosphonate methylphosphonate	5-methylcytidine 5-methylcytidine	00	K53 K54	Br I	methylphosphonate methylphosphonate	5-aminoallyluridine 5-aminoallyluridine
J53	Br	methylphosphonate	5-methylcytidine		K55	$^{1}_{ m NH_{2}}$	methylphosphonate	5-aminoallyluridine
J54	I	methylphosphonate	5-methylcytidine		K56	H	methylphosphonate	5-aminoallyluridine
J55	$NH_2$	methylphosphonate	5-methylcytidine		K57	ОН	boranophosphonate	5-aminoallyluridine
J56	Н	methylphosphonate	5-methylcytidine		K58	OMe	boranophosphonate	5-aminoallyluridine
J57	OH	boranophosphonate	5-methylcytidine	65	K59	F	boranophosphonate	5-aminoallyluridine
J58	OMe	boranophosphonate	5-methylcytidine		<b>K</b> 60	Cl	boranophosphonate	5-aminoallyluridine

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TABLE 2-continued

# 24 TABLE 2-continued

Exemplary modified nucleotides contained in a synthetic guide sequence. Exemplary modified nucleotides contained in a synthetic guide sequence

B  $R_1 = OH \text{ or } 2' \text{modificiation}$   $R_2 = OH \text{ or internucleotide linkage}$  B = base

	$R_2$	$R_1$	
#	$R_1$	$R_2$	В
K61	Br	boranophosphonate	5-aminoallyluridine
K62	I	boranophosphonate	5-aminoallyluridine
K63	$NH_2$	boranophosphonate	5-aminoallyluridine
K64	Н	boranophosphonate	5-aminoallyluridine
L1	OH	OH	5-methyluridine
L2 L3	OMe F	OH OH	5-methyluridine 5-methyluridine
L3 L4	Cl	OH	5-methyluridine
L5	Br	OH	5-methyluridine
L6	I	ОН	5-methyluridine
L7	$NH_2$	ОН	5-methyluridine
L8	H	OH	5-methyluridine
L9	OH	phosphodiester	5-methyluridine
L10 L11	OMe F	phosphodiester	5-methyluridine 5-methyluridine
L11	r Cl	phosphodiester phosphodiester	5-methyluridine
L13	Br	phosphodiester	5-methyluridine
L14	I	phosphodiester	5-methyluridine
L15	$NH_2$	phosphodiester	5-methyluridine
L16	H	phosphodiester	5-methyluridine
L17	OH	phosphonoacetate	5-methyluridine
L18	OMe	phosphonoacetate	5-methyluridine
L19	F	phosphonoacetate	5-methyluridine
L20 L21	Cl Br	phosphonoacetate	5-methyluridine 5-methyluridine
L21	I	phosphonoacetate phosphonoacetate	5-methyluridine
L23	$^{ m NH}_2$	phosphonoacetate	5-methyluridine
L24	H	phosphonoacetate	5-methyluridine
L25	OH	thiophosphonoacetate	5-methyluridine
L26	OMe	thiophosphonoacetate	5-methyluridine
L27	F	thiophosphonoacetate	5-methyluridine
L28	Cl	thiophosphonoacetate	5-methyluridine
L29 L30	Br I	thiophosphonoacetate thiophosphonoacetate	5-methyluridine 5-methyluridine
L30	$^{1}_{ m NH_{2}}$	thiophosphonoacetate	5-methyluridine
L32	H	thiophosphonoacetate	5-methyluridine
L33	OH	phosphorothioate	5-methyluridine
L34	OMe	phosphorothioate	5-methyluridine
L35	F	phosphorothicate	5-methyluridine
L36	Cl	phosphorothioate	5-methyluridine
L37 L38	Br I	phosphorothioate	5-methyluridine
L38 L39	$^{1}_{ m NH_{2}}$	phosphorothioate phosphorothioate	5-methyluridine 5-methyluridine
L40	H	phosphorothioate	5-methyluridine
L41	OH	phosphorodithioate	5-methyluridine
L42	OMe	phosphorodithioate	5-methyluridine
L43	F	phosphorodithioate	5-methyluridine
L44	Cl	phosphorodithioate	5-methyluridine
L45	Br	phosphorodithioate	5-methyluridine
L46 L47	$_{ m NH_2}$	phosphorodithioate phosphorodithioate	5-methyluridine 5-methyluridine
L48	H H	phosphorodithioate	5-methyluridine
L49	OH	methylphosphonate	5-methyluridine
L50	OMe	methylphosphonate	5-methyluridine
L51	F	methylphosphonate	5-methyluridine
L52	Cl	methylphosphonate	5-methyluridine
L53	$_{\mathrm{Br}}$	methylphosphonate	5-methyluridine
L54	I	methylphosphonate	5-methyluridine
L55	$NH_2$	methylphosphonate	5-methyluridine
L56	Н	methylphosphonate	5-methyluridine
L57	OH	boranophosphonate	5-methyluridine
L58	OMe	boranophosphonate	5-methyluridine
L59 L60	F Cl	boranophosphonate boranophosphonate	5-methyluridine 5-methyluridine
L60 L61	Br	boranophosphonate	5-methyluridine
1.60	10	barran barrhanata	5-mearyruridine

5-methyluridine

L62

Ι

boranophosphonate

B  $R_1 = OH \text{ or } 2^t\text{modificiation}$   $R_2 = OH \text{ or internucleotide linkage}$  B = base

 $\begin{array}{c|ccccc} \# & R_1 & R_2 & B \\ \hline L63 & NH_2 & boranophosphonate & 5-methyluridine \\ L64 & H & boranophosphonate & 5-methyluridine \\ \end{array}$ 

As described herein, certain unnatural base pairs (e.g., isoG and isoC, Z base and P base; see Zhang et al. (2015) *J. Am. Chem. Soc.*) may be advantageous for affecting the thermostability of the guide RNA secondary structure. These modifications can be used to prevent misfolding of the guide RNA scaffold with other domains of a guide RNA sequence.

Recent guide RNA:Cas9 protein structural information (FIG. 10, as reported in Jiang et al. 2015, *Science*) and in vivo/in vitro functional mutation studies (see, e.g., Briner et al. 2014, *Mol. Cell*, 56, 333-9) indicate that the guide RNA scaffold is predominantly structurally conserved. This reinforces the importance of correct folding of the conserved domain of guide RNAs for functionality with Cas9. FIG. 10 shows the guide RNA scaffold secondary structure, displaying interactions with amino acids of Cas9. Most of the guide RNA nitrogenous bases are not involved in binding interactions with Cas9 protein.

The flanking sequences of the sgRNA scaffold increase 35 the likelihood of misfolding and hence misfunction. The 20 nt guide targeting sequence, 5' of the scaffold region, is user-specified for each target, thus the likelihood of misfolding is variable or target-specific. Also, many emerging CRISPR-Cas applications append functional sequences 3' of 40 the scaffold, such as CRISPRdisplay (Schechner et al., Nat. Methods 2015) and CRISPR-i/-a (Chen et al., Cell 2013), which are riboswitches or aptamers that also need to correctly and independently fold to function properly. To ensure that each of the functional domains (i.e. targeting guide, scaffold, aptamer) of a given sgRNA folds in a modular, independent manner, the structurally conserved scaffold base pairs can be substituted with unnatural, orthogonal base pairs (e.g., isoG and isoC; Z base and P base), and in some embodiments, substituted exclusively with unnatural, 50 orthogonal base pairs. This ensures that the sgRNA scaffold sequences will not stably interact in a secondary structure with elements of the target-pairing guide sequence or other non-native domains incorporated in the guide RNA such as any aptamer sequences or any non-native 5' or 3' overhangs on the guide RNA. Alternatively, the unnatural, orthogonal base pairs mentioned above could be incorporated in any non-native overhangs or aptamers that may be present, thus to prevent secondary structures involving misfolding of the scaffold sequence(s). 60

# B. Guide RNA with at Least One Modification

In one aspect, the present technology provides a guide RNA having at least one modification, constituting a modi-65 fied gRNA.

In certain embodiments, the modified gRNA comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19,

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20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 modified nucleotides. In other embodiments, the modified gRNA comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified nucleotides. In certain 5 embodiments, all nucleotides are modified. In certain embodiments, all the modifications are the same. In certain embodiments, all the modified nucleotides have the same type of modification. In certain embodiments, the modified gRNA comprises a combination of differently modified 10 nucleotides. In certain embodiments, the modified gRNA comprises two or more modified nucleotides. In certain embodiments, the modified gRNA comprises three or more modified nucleotides. In certain embodiments, the modified nucleotides are arranged contiguously. In certain embodi- 15 ments, the modified gRNA comprises at least one contiguous stretch of modified nucleotides. In certain embodiments, the modified gRNA comprises a contiguous stretch of at least 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 20 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 modified nucleotides. Each modified nucleotide may independently comprise one or more types of modifications. In certain embodiments, no modified nucleotides are contiguous, or some but not all are contiguous, in the sequence 25 of the modified gRNA.

In certain embodiments, the modification is within the 5' portion of the guide RNA. In certain embodiments, the modification is within the first five (5) nucleotides of the 5' portion of the guide RNA. In certain embodiments, the modification is within the first three (3) nucleotides of the 5' portion of the guide RNA. In certain embodiments, the modification is within the 3' portion of the guide RNA. In certain embodiments, the modification is within the last five (5) nucleotides of the 3' portion of the guide RNA. In certain embodiments, the modification is within the last three (3) nucleotides of the 3' portion of the guide RNA. In certain embodiments, the modification is within the internal region (i.e., between the 5' end and the 3' end) of the guide RNA.

In certain embodiments, the modification is incorporated 40 in the 5' portion or the 3' portion of the guide RNA, particularly within the first 5 or 10 nucleotides of the 5' portion or within the last 5 or 10 nucleotides of the 3' portion to, for example, protect the RNA from degradation by nucleases or for other purposes. In some other embodiments, 45 the modification is in both the 5' portion and the 3' portion of the guide RNA, particularly within the first 5 or 10 nucleotides of the 5' portion and within the last 5 or 10 nucleotides of the 3' portion to, for example, protect the RNA from degradation by nucleases or for other purposes. 50 In certain embodiments, more than one type of modification is present in both the 5' portion and the 3' portion of the guide RNA. In certain embodiments, the modifications are located at the 5' end, at the 3' end, and within the internal sequence of the guide RNA. In certain embodiments, a guide 55 RNA comprises 40 or fewer, alternatively 20 or fewer, alternatively 15 or fewer, alternatively 10 or fewer, alternatively 5 or fewer, alternatively 3 or fewer deoxyribonucleotide residues in the 5' or 3' portion of the guide RNA.

In certain embodiments, the modification is within the 60 crRNA segment of the guide RNA. In certain embodiments, the modification is within the guide sequence of the crRNA. In certain embodiments, the modification is within the first five (5) nucleotides of the crRNA segment. In certain embodiments, the modification is within the first three (3) 65 nucleotides of the crRNA segment. In certain embodiments, the modification is within a 5'-overhang on the crRNA

segment. In certain embodiments, the modification is within the tracrRNA segment of the guide RNA. In certain embodiments, the modification is within the last five (5) nucleotides of the tracrRNA segment of the guide RNA. In certain embodiments, the modification is within the last three (3) nucleotides of the tracrRNA segment of the guide RNA. In certain embodiments, when the guide RNA is a single guide RNA, the modification is located within the loop of the guide RNA. In certain embodiments, one or more modifications is within the loop L region. In certain embodiments, the modification comprises a dye, a non-fluorescent label, or a tag conjugated to a linker incorporated between two nucleotides as described above, for example by conjugation to a 2-(3-(dye/label/tag-amido)propanamido)propane-1,3diol bis(phosphodiester) linker or to a modified base of a nucleotide in the loop or L region.

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In certain embodiments, the modification comprises an end modification, such as a 5' end modification or a 3' end modification. Examples of end modifications include, but are not limited to phosphorylation (as natural phosphate or polyphosphate or as modified phosphohate groups such as for example, alkylphosphonate, phosphonocarboxylate, phosphonoacetate, boranophosphonate, phosphorothioate, phosphorodithioate and the like), biotinylation, conjugating or conjugated molecules, linkers, dyes, labels, tags, functional groups (such as for example but not limited to 5'-amino, 5'-thio, 5'-amido, 5'carboxy and the like), inverted linkages, or hydrocarbon moieties which may comprise ether, polyethylene glycol (PEG), ester, hydroxyl, aryl, halo, phosphodiester, bicyclic, heterocyclic or other organic functional group. In certain embodiments, the end modification comprises dimethoxytrityl.

In certain embodiments, the modification comprises a modified base. As used herein, "unmodified" bases include the purine bases adenine (A) and guanine (G), and the pyrimidine bases thymine (T), cytosine (C) and uracil (U). Examples of modified bases include, but are not limited to, synthetic and natural bases such as 2-thioU, 2-thioC, 4-thioU, 6-thioG, 2-aminoA, 2-aminoP, pseudouracil, hypoxanthine, 7-deazaguanine, 7-deaza-8-azaguanine, 7-deazaadenine. 7-deaza-8-azaadenine, 5-methylC, 5-methylU, 5-hydroxymethylcytosine, 5-hydroxymethyluracil, 5,6-dehydrouracil, 5-propynylcytosine, 5-propynyluracil, 5-ethynylcytosine, 5-ethynyluracil, 5-allylU, 5-allylC, 5-aminoallyl-uracil, and 5-aminoallyl-cytosine. In certain embodiments, the modification comprises an abasic nucleotide. In certain embodiments, the modification comprises a nonstandard purine or pyrimidine structure, such as Z or P, isoC or isoG, UNA, 5-methylpyrymidine, x(A,G,C,T) or y(A,G,C,T). In certain embodiments, the modified gRNA comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 modified bases. In other embodiments, the modified gRNA comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified bases. In certain embodiments, all bases in a gRNA are modified.

In certain embodiments, the modification comprises a modified sugar. Examples of modified sugars include, but are not limited to, sugars having modifications at the 2' position or modifications at the 4' position. For example, in certain embodiments, the sugar comprises 2'-O— $C_{1-4}$ alkyl, such as 2'-O-methyl (2'-OMe). In certain embodiments, the sugar comprises 2'-O— $C_{1-3}$ alkyl-O— $C_{1-3}$ alkyl, such as 2'-methoxyethoxy (2'-O— $CH_2CH_2OCH_3$ ) also known as 2'-O-(2-methoxyethyl) or 2'-MOE. In certain embodiments, the sugar comprises 2'-halo, such as 2'-F, 2'-Br, 2'-Cl, or 2'-I.

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In certain embodiments, the sugar comprises 2'-NH<sub>2</sub>. In certain embodiments, the sugar comprises 2'-H (e.g., a deoxynucleotide). In certain embodiments, the sugar comprises 2'-arabino or 2'-F-arabino. In certain embodiments, the sugar comprises 2'-LNA or 2'-ULNA. In certain embodiments, the sugar comprises a 4'-thioribosyl. In certain embodiments, the modified gRNA comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 modified sugars. In other embodiments, the modified gRNA comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified sugars. In certain embodiments, all sugars in a gRNA are modified.

modified backbone (i.e., an internucleotide linkage other than a natural phosphodiester). Examples of modified internucleotide linkages include, but are not limited to, a phosphorothioate internucleotide linkage, a chiral phosphorothioate internucleotide linkage, a phosphorodithioate 20 internucleotide linkage, a boranophosphonate internucleotide linkage, a C<sub>1-4</sub>alkyl phosphonate internucleotide linkage such as a methylphosphonate internucleotide linkage, a boranophosphonate internucleotide linkage, a phosphonocarboxylate internucleotide linkage such as a phosphonoac- 25 etate internucleotide linkage, a phosphonocarboxylate ester internucleotide linkage such as a phosphonoacetate ester internucleotide linkage, a thiophosphonocarboxylate internucleotide linkage such as for example a thiophosphonoacetate internucleotide linkage, a thiophosphonocarboxylate 30 ester internucleotide linkage such as a thiophosphonoacetate ester internucleotide linkage. Various salts, mixed salts and free acid forms are also included. In certain embodiments, the modified gRNA comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 35 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 modified internucleotide linkages. In other embodiments, the modified gRNA comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified internucleotide linkages. In certain 40 embodiments, all internucleotide linkages in a gRNA are modified.

In certain embodiments, the modification is a 2'-O-C<sub>1-</sub> 4alkyl, 2'-H, 2'-O— $C_{1-3}$ alkyl-O— $C_{1-3}$ alkyl, 2'-F, 2'-NH $_2$ , 2'-arabino, 2'-F-arabino, 2'-LNA, 2'-ULNA, 4'-thioribosyl, 45 2-thioU, 2-thioC, 4-thioU, 6-thioG, 2-aminoA, 2-aminoP, pseudouracil, hypoxanthine, 7-deazaguanine, 7-deaza-8-azaguanine, 7-deazaadenine, 7-deaza-8-azaadenine, 5-MeC, 5-MeU, 5-hydroxymethylcytosine, 5-hydroxymethyluracil, 5,6-dehydrouracil, 5-propynylcytosine, 5-propynyluracil, 50 5-ethynylcytosine, 5-ethynyluracil, 5-allylU, 5-allylC, 5-aminoallyl-uracil, 5-aminoallyl-cytosine, an abasic nucleotide, Z, P, UNA, isoC, isoG, 5-methyl-pyrimidine, x(A,G,C,T), y(A,G,C,T), a 3'-phosphorothioate group, a 3'-phosphonoacetate group, a 3'-phosphonoacetate ester 55 group, a 3'-thiophosphonoacetate group, a 3'-thiophosphonoacetate ester group, a 3'-methylphosphonate group, a 3'-boranophosphonate group, a 3'-phosphorodithioate group, or combinations thereof.

In certain embodiments, the modified nucleotide comprises a 2'-O-methyl-3'-phosphorothioate. In certain embodiments, the modified nucleotide comprises a 2'-O-methyl-3'-phosphonoacetate. In certain embodiments, the modified nucleotide comprises a 2'-O-methyl-3'-thio-phosphonoacetate. In certain embodiments, the modified comprises a Z base. In certain embodiments, the modified nucleotide comprises a Z'-halo-3'-phosphorothio-

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ate. In certain embodiments, the modified nucleotide comprises a 2'-halo-3'-phosphonoacetate. In certain embodiments, the modified nucleotide comprises a 2'-halo-3'-thiophosphonoacetate. In certain embodiments, the modified nucleotide comprises a 2'-fluoro-3'-phosphorothioate. In certain embodiments, the modified nucleotide comprises a 2'-fluoro-3'-phosphonoacetate. In certain embodiments, the modified nucleotide comprises a 2'-fluoro-3'-thiophosphonoacetate.

In certain embodiments, the guide RNA comprises an oligonucleotide represented by Formula (I):

$$W-Y \text{ or } Y-W$$
 (I)

a gRNA are modified.

In certain embodiments, the modification comprises a 15 tides of the oligonucleotide comprising at least one modified backbone (i.e., an internucleotide linkage other an a natural phosphodiester). Examples of modified inter-

In certain embodiments, W is within the 5' portion of the guide RNA. In certain embodiments, W is at least partially within the first five (5) nucleotides of the 5' portion of the guide RNA. In certain embodiments, W is at least partially within the first three (3) nucleotides of the 5' portion of the guide RNA. In certain embodiments, W is within the 3' portion of the guide RNA. In certain embodiments, W is at least partially within the last five (5) nucleotides of the 3' portion of the guide RNA. In certain embodiments, W is at least partially within the last three (3) nucleotides of the 3' portion of the guide RNA. In certain embodiments, W is within the internal region (i.e., between the 5' end and the 3' end) of the guide RNA.

In certain embodiments, W comprises an end modification, such as a 5' end modification or a 3' end modification as described above. In certain embodiments, the end modification comprises dimethoxytrityl.

In certain embodiments, W comprises a modified base as described above. In certain embodiments, W comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 modified bases. In other embodiments, W comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified bases. In certain embodiments, all bases in a gRNA are modified.

In certain embodiments, W comprises a modified sugar as described above. In certain embodiments, W comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 modified sugars. In other embodiments, W comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified sugars. In certain embodiments, all sugars in a gRNA are modified.

In certain embodiments, W comprises a modified backbone (i.e., an internucleotide linkage other than a phosphodiester) as described above. In certain embodiments, W comprises more than one, such as 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49 or 50 modified internucleotide linkages. In other embodiments, W comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified internucleotide linkages. In certain embodiments, all internucleotide linkages in a gRNA are modified.

In certain embodiments, W comprises a 2'-O—C<sub>1-4</sub>alkyl, 2'-H, 2'-O—C<sub>1-3</sub>alkyl-O—C<sub>1-3</sub>alkyl, 2'-F, 2'-NH<sub>2</sub>, 2'-arabino, 2'-F-arabino, 2'-LNA, 2'-ULNA, 4'-thioribosyl, 2-thioU, 2-thioU, 4-thioU, 6-thioG, 2-aminoA, 2-aminoP,

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pseudouracil, hypoxanthine, 7-deazaguanine, 7-deaza-8-azaguanine, 7-deazaadenine, 7-deaza-8-azaadenine, 5-MeC, 5-MeU, 5-hydroxymethylcytosine, 5-hydroxymethyluracil, 5,6-dehydrouracil, 5-propynylcytosine, 5-propynyluracil, 5-ethynylcytosine, 5-ethynyluracil, 5-allylU, 5-allylC, 5 5-aminoallyl-uracil, 5-aminoallyl-cytosine, abasic nucleotides, Z, P, UNA, isoC, isoG, 5-methyl-pyrimidine, x(A,G, C,T), y(A,G,C,T), a phosphorothioate internucleotide linkage, a phosphonoacetate internucleotide linkage, a phosphonoacetate ester internucleotide linkage, a thio- 10 phosphonoacetate internucleotide linkage, a thiophosphonoacetate ester internucleotide linkage a methylphosphonate internucleotide linkage, a boranophosphonate internucleotide linkage, a phosphorodithioate internucleotide linkage, or combinations thereof.

In certain embodiments, W comprises a 2'-O-methyl and a 3'-phosphorothioate group on the same nucleotide. In certain embodiments, W comprises a 2'-O-methyl and a 3'-phosphonoacetate group on the same nucleotide. In certain embodiments, W comprises a 2'-O-methyl and 3<sup>1</sup>-thio- 20 phosphonoacetate group on the same nucleotide. In certain embodiments, W comprises a 2'-F and a 3'-phosphorothioate group on the same nucleotide. In certain embodiments, W comprises a 2'-F and a 3'-phosphonoacetate group on the same nucleotide. In certain embodiments, W comprises a 25 2'-F and 3<sup>1</sup>-thiophosphonoacetate group on the same nucleotide.

In certain embodiments, W comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20 modified nucleotides. In certain embodiments, each of the modified 30 nucleotides comprises the same modification. In certain embodiments, W comprises a combination of variously modified nucleotides. In certain embodiments, W comprises two or more modified nucleotides. In certain embodiments, W comprises three or more modified nucleotides. In certain 35 embodiments, the modified nucleotides are not arranged contiguously in the sequence, or at least not entirely, as one or more unmodified nucleotides may intercede. In certain embodiments, the modified nucleotides are arranged concontiguous stretch of modified nucleotides. In certain embodiments, W comprises a contiguous stretch of at least three (3) modified nucleotides. In certain embodiments, W comprises a contiguous stretch of at least four (4) modified nucleotides. In certain embodiments, W comprises a con- 45 tiguous stretch of at least five (5) modified nucleotides.

In certain embodiments, the guide RNA comprises an oligonucleotide represented by Formula (II):

$$\mathbf{M}_{m}\mathbf{N}_{n}$$
 (II)

wherein each N independently represents an unmodified ribonucleotide;

wherein each M represents a modified nucleotide and is independently selected from the group consisting of a 2'-Omethyl ribonucleotide, a 3'-P(S) ribonucleotide, a 3'-PACE 55 ribonucleotide, a 3'-thioPACE ribonucleotide, a 2'-Omethyl-3'-P(S)-ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, a 2'-O-methyl-3'-thioPACE ribonucleotide, a Z nucleotide, and a 2'-deoxynucleotide;

wherein each M is at any position of the sequence of the 60 guide RNA;

wherein any given M is the same or different from any other M, and any given N is the same or different from any other N; and

wherein each of m and n are independently selected from 65 an integer between 0 and 219, provided that 50<m+n≤220, and m is not 0.

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In some embodiments, m+n<150.

In certain embodiments, each M is modified with one or more moieties independently selected from the group consisting of 2'-F, 2-thiouracil, 4-thiouracil, 2-aminoadenine, hypoxanthine, 5-methylcytosine, 5-methyluracil, 5-allylaminouracil, squarate linkage, a triazolo linkage, and a 2-(4-butylamidofluorescein)propane-1,3-diol phodiester) linkage. In some embodiments, M comprises a dye attached through a linker.

In certain embodiments, each M is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-P(S) ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, and a 2'-O-methyl-3'-thioPACE ribonucleotide. In certain embodiments, each M is independently selected from the group consisting of a 2'-O-methyl-3'-PACE ribonucleotide and a 2'-O-methyl-3'-thioPACE ribonucleotide.

In certain embodiments, where m>1, any given M is the same or different from any other M. In certain embodiments, where m>1, each M has the same modification.

In certain embodiments, each M is a 2'-O-methyl-3'-PACE ribonucleotide, m is selected from an integer between 1 and 10, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, each M is a 2'-O-methyl-3'-PACE ribonucleotide, m is selected from an integer between 1 and 5, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, each M is a 2'-O-methyl-3'-PACE ribonucleotide, m is selected from an integer between 2 and 5, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 148, provided 50<m+n≤150. In certain embodiments, m is 1. In certain embodiments, m is 2. In certain embodiments, m is 3. In certain embodiments, m is 4. In certain embodiments,

In certain embodiments, each M is a 2'-O-methyl-3'tiguously. In certain embodiments, W comprises at least one 40 thioPACE ribonucleotide, m is selected from an integer between 1 and 10, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, each M is a 2'-O-methyl-3'-thioPACE ribonucleotide, m is selected from an integer between 1 and 5, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, each M is a 2'-O-methyl-3'-thioPACE ribo- $^{
m (II)}$  50 nucleotide, m is selected from an integer between 2 and 5, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 148, provided 50<m+n≤150. In certain embodiments, m is 1. In certain embodiments, m is 2. In certain embodiments, m is 3. In certain embodiments, m is 4. In certain embodiments, m is 5.

> In certain embodiments, each M is a 2'-O-methyl ribonucleotide, m is selected from an integer between 1 and 40, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, each M is a 2'-O-methyl ribonucleotide, m is selected from an integer between 1 and 25, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, each M is a 2'-Omethyl ribonucleotide, m is selected from an integer

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between 1 and 20, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, m is 1. In certain embodiments, m is 2. In certain embodiments, m is 3. In certain embodiments, 5 m is 4. In certain embodiments, m is 5. In certain embodiments, m is 10. In certain embodiments, m is 15. In certain embodiments, m is 20. In certain embodiments, m is 30. In certain embodiments, m is 40.

In certain embodiments, each M is a 2'-deoxynucleotide, 10 m is selected from an integer between 1 and 30, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, each M is 2'-deoxynucleotide, m is selected from an integer between 1 15 and 20, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, m is 5. In certain embodiments, m is 10. In certain embodiments, m is 15. In certain embodi- 20 ments, m is 20. In certain embodiments, m is 30.

In certain embodiments, each M is a 2'-O-methyl-3'-P(S) ribonucleotide, m is selected from an integer between 1 and 10, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer 25 between 1 and 149, provided 50<m+n≤150. In certain embodiments, each M is a 2'-O-methyl-3'-P(S) ribonucleotide, m is selected from an integer between 1 and 5, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 30 149, provided 50<m+n≤150. In certain embodiments, m is 1. In certain embodiments, m is 2. In certain embodiments, m is 3. In certain embodiments, m is 4. In certain embodiments,

In certain embodiments, each M is a Z nucleotide, m is 35 selected from an integer between 1 and 10, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, each M is a Z nucleotide, m is selected from an integer between 1 and 40 5, each N is independently selected from the group consisting of A, U, C, and G, and n is selected from an integer between 1 and 149, provided 50<m+n≤150. In certain embodiments, m is 1. In certain embodiments, m is 2. In 4. In certain embodiments, m is 5.

In certain embodiments, the modification is a stabilityaltering modification. In certain embodiments, the modification increases nuclease resistance of the guide RNA relative to a guide RNA without the modification, thus it 50 enhances the guide RNA stability. In certain embodiments, the stability-altering modification is a stability-enhancing modification. For example, in certain embodiments, the stability-enhancing modification comprises a 2'-O-methyl or a 2'-O-C<sub>1-4</sub>alkyl nucleotide. In certain embodiments, the 55 stability-enhancing modification comprises a 2'-halo nucleotide, such as 2'-F, 2'-Br, 2'-Cl, or 2'-I. In certain embodiments, the stability-enhancing modification comprises a 2'MOE or a 2'-O—C<sub>1-3</sub>alkyl-O—C<sub>1-3</sub>alkyl. In certain embodiments, the stability-enhancing modification com- 60 prises a 2'-NH2 nucleotide. In certain embodiments, the stability-enhancing modification comprises a 2'-H (or 2'-deoxy) nucleotide. In certain embodiments, the stability-enhancing modification comprises a 2'-arabino or a 2'-Farabino. In certain embodiments, the stability-enhancing 65 modification comprises a 4'-thioribosyl sugar moiety. In certain embodiments, the stability-enhancing modification

comprises a 3'-phosphorothioate group. In certain embodiments, the stability-enhancing modification comprises a 3'-phosphonoacetate group. In certain embodiments, the stability-enhancing modification comprises a nucleotide containing a 3'-thiophosphonoacetate group. In certain embodiments, the stability-enhancing modification comprises a nucleotide containing a 3'-methylphosphonate group. In certain embodiments, the stability-enhancing modification comprises a nucleotide containing a 3'-boranophosphate group. In certain embodiments, the stabilityenhancing modification comprises a nucleotide containing a 3'-phosphorodithioate group. In certain embodiments, the stability-enhancing modification comprises a locked nucleic acid ("LNA") nucleotide. In certain embodiments, the stability-enhancing modification comprises an unlocked nucleic acid ("ULNA") nucleotide.

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In certain embodiments, the stability-enhancing modification comprises a 2'-O-methyl and a 3'-phosphorothioate group on the same nucleotide. In certain embodiments, the stability-enhancing modification comprises a 2'-O-methyl and a 3'-phosphonoacetate group on the same nucleotide. In certain embodiments, the stability-enhancing modification comprises a 2'-O-methyl and a 3'-thiophosphonoacetate group on the same nucleotide. In certain embodiments, the stability-enhancing modification comprises a 2'-fluoro and a 3'-phosphorothioate group on the same nucleotide. In certain embodiments, the stability-enhancing modification comprises a 2'-fluoro and a 3'-phosphonoacetate group on the same nucleotide. In certain embodiments, the stabilityenhancing modification comprises a 2'-fluoro and a 3'-thiophosphonoacetate group on the same nucleotide.

In certain embodiments, the modification is a specificityaltering modification. In some embodiments, specificity enhancement may be achieved by enhancing on-target binding and/or cleavage, or reducing off-target binding and/or cleavage, or a combination of both. In some other embodiments, specificity reduction may be achieved, for example, by reducing on-target binding and/or cleavage, or increasing off-target binding and/or cleavage, or a combination of both.

In certain embodiments, the specificity-altering modification comprises a 2'-O-methyl. In certain embodiments, the specificity-altering modification comprises a 2'-halo, such as 2'-fluoro.

In certain embodiments, the specificity-altering modificertain embodiments, m is 3. In certain embodiments, m is 45 cation comprises a 2-thiouracil base (2-thioU). In certain embodiments, the specificity-altering modification comprises 2-thioC. In certain embodiments, the specificityaltering modification comprises 4-thioU. In certain embodiments, the specificity-altering modification comprises 6-thioG. In certain embodiments, the specificity-altering modification comprises 2-aminoA. In certain embodiments, the specificity-altering modification comprises a 2-aminopurine. In certain embodiments, the specificity-altering modification comprises pseudouracil. In certain embodiments, the specificity-altering modification comprises hypoxanthine. In certain embodiments, the specificity-altering modification comprises 7-deazaguanine. In certain embodiments, the specificity-altering modification comprises 7-deaza-8azaguanine. In certain embodiments, the specificity-altering modification comprises 7-deazaadenine. In certain embodiments, the specificity-altering modification comprises 7-deaza-8-azaadenine. In certain embodiments, the specificity-altering modification comprises 5-methylC. In certain embodiments, the specificity-altering modification comprises 5-methylU. In certain embodiments, the specificityaltering modification comprises 5-hydroxymethylcytosine. In certain embodiments, the specificity-altering modification

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comprises 5-hydroxymethyluracil. In certain embodiments, the specificity-altering modification comprises 5,6-dehydrouracil. In certain embodiments, the specificity-altering modification comprises 5-propynylcytosine. In certain embodiments, the specificity-altering modification com- 5 prises 5-propynyluracil. In certain embodiments, the specificity-altering modification comprises 5-ethynylcytosine. In certain embodiments, the specificity-altering modification comprises 5-ethynyluracil. In certain embodiments, the specificity-altering modification comprises 5-allylU. In certain embodiments, the specificity-altering modification comprises 5-allylC. In certain embodiments, the specificityaltering modification comprises 5-aminoallylU. In certain embodiments, the specificity-altering modification comprises 5-aminoallylC. In certain embodiments, the specific- 15 ity-altering modification comprises an abasic nucleotide. In certain embodiments, the specificity-altering modification comprises a Z base. In certain embodiments, the specificityaltering modification comprises P base. In certain embodiments, the specificity-altering modification comprises a 20 UNA base. In certain embodiments, the specificity-altering modification comprises isoC. In certain embodiments, the specificity-altering modification comprises isoG. In certain embodiments, the specificity-altering modification comprises 5-methyl-pyrimidine. In certain embodiments, the 25 specificity-altering modification comprises x(A,G,C,T). In certain embodiments, the specificity-altering modification comprises y(A,G,C,T).

In certain embodiments, the specificity-altering modification comprises a phosphorothioate internucleotide link- 30 age. In certain embodiments, the specificity-altering modification comprises a phosphonoacetate internucleotide linkage. In certain embodiments, the specificity-altering modification comprises a thiophosphonoacetate internucleotide linkage. In certain embodiments, the specificity-alter- 35 ing modification comprises a methylphosphonate internucleotide linkage. In certain embodiments, the specificityaltering modification comprises a boranophosphate internucleotide linkage. In certain embodiments, the specificity-altering modification comprises a phosphorodithioate 40 internucleotide linkage. In certain embodiments, the specificity-altering modification comprises a ULNA. In certain embodiments, the specificity-altering modification comprises an LNA.

In certain embodiments, the modification alters RNA base 45 pairing by, for example, altering the melting temperature (Tm) of the guide RNA relative to a guide RNA without the modification. In certain embodiments, the modification lowers the Tm of the guide RNA relative to a guide RNA without the modification. In certain embodiments, the modification raises the Tm of the guide RNA relative to a guide RNA without the modification.

In certain embodiments, the specificity-altering modification lowers the Tm of a base pairing interaction. In certain embodiments, the modification that lowers the Tm of the 55 base pairing interaction is a 2'-deoxy, as it is well-known in the art that DNA/DNA base pairs have lower Tm than their respective counterpart in RNA/DNA duplexes. In certain embodiments, the modification that lowers the Tm of the base pairing interaction is 2-thiouracil, which slightly lowers 60 Tm of G-U wobble pair. In certain embodiments, the modification that lowers the Tm of the base pairing interaction is a phosphorothioate internucleotide linkage or a phosphorodithioate internucleotide linkage, which lower the Tm by ~0.5° C. per modification. In certain embodiments, the 65 modification that lowers the Tm of the base pairing interaction is a boranophosphonate internucleotide linkage,

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which lowers the Tm by ~0.5-0.8° C. per modification. In certain embodiments, the modification that lowers the Tm of the base pairing interaction is a phosphonoacetate internucleotide linkage, which lowers the Tm by ~1.3° C. per modification. In certain embodiments, the modification that lowers the Tm of the base pairing interaction is unlocked nucleic acid ("ULNA"), which lowers the Tm by ~5-8° C. per modification. In certain embodiments, the modification that lowers the Tm of the base pairing interaction is 2'-O-methyl-3'-methylphosphonate.

In certain embodiments, the specificity-altering modification raises the Tm of a base pairing interaction. In certain embodiments, the modification that raises the Tm of the base pairing interaction is a 2'-O-methyl, which raises Tm by ~0.5-0.7° C. per modification. In certain embodiments, the modification that raises the Tm of the base pairing interaction is a 2'-F, which raises Tm by ~1° C. per modification. In certain embodiments, the modification that raises the Tm of the base pairing interaction is a 2-thiouracil, which raises Tm of A-U pair (and, as noted above, slightly lowers Tm of G-U wobble pair). In certain embodiments, the modification that raises the Tm of the base pairing interaction is a 4-thiouracil, which raises Tm of G-U wobble pair and slightly raises Tm of A-U pair. In certain embodiments, the modification that raises the Tm of the base pairing interaction is a 2-amino-adenine, which raises Tm of its base pairing with U by ~1° C. per modification. In certain embodiments, the modification that raises the Tm of the base pairing interaction is a 5-methyl-uracil (5-methylU) (see, e.g., Wang & Kool (1995) Biochemistry, 34, 4125-32). In certain embodiments, the modification that raises the Tm of the base pairing interaction is a 5-methyl-cytosine (5-methylC). In certain embodiments, the modification that raises the Tm of the base pairing interaction is a locked nucleic acid ("LNA"), which raises Tm by 2-10° C. per modifica-

In certain embodiments, the modification alters transfection efficiency of the guide RNA relative to a guide RNA without the modification. In certain embodiments, the modification increases transfection efficiency of the guide RNA relative to a guide RNA without the modification. In certain embodiments, the modification decreases transfection efficiency of the guide RNA relative to a guide RNA without the modification. In certain embodiments, the modification neutralizes the anionic charge on phosphate to allow passive diffusion into cells. In certain embodiments, the charge-neutralizing modification comprises a phosphonoacetate alkyl ester internucleotide linkage, such as a phosphonoacetate methyl ester internucleotide linkage.

In certain embodiments, the modification alters the immunostimulatory effect of the guide RNA relative to a guide RNA without the modification. It was initially discovered that unmethylated bacterial DNA and synthetic analogs thereof are ligands for TLR9 (see Hemmi et al. (2000) Nature, 408, 740-5). The stimulation of TLR9 can be mitigated in the dinucleotide motif for example by modifying the C and G residues. The use of 5-methylcytosine, 2-aminocytosine, 2-thiocytosine, 5-methylisocytosine, P nucleobase (6-(β-D-2'-Deoxyribofuranosyl)-3,4-dihydro-8H-pyrimido[4,5-c][1,2]oxazin-7-one), and 2'-O-methylcytosine all result in loss or decrease in TLR9 stimulation. In certain embodiments, use of 6-thioguanine, 2,6-diaminopurine, 2-aminopurine, xanthosine, inosine, 7-deazaxanthosine, isoguanine, 8-oxoguanine, nebularine, 8-bromoguanine, K-nucleobase (2-amino-N-methoxyadenosine), and/or 2'-O-methylguanine can result in loss or decrease in TLR9 stimulation. In some embodiments, use of phosphodiester

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Typically, synthetically incorporated phosphorothioates can decrease the TLR9 response to a limited extent, as is thought to result from the presence of two stereoisomers of each phosphorothioate in synthetic RNA. However, it has been 5 shown that phosphorothioate-modified DNA lacking CpG motifs stimulate TLR9 to a rather small extent. The negative charge on the phosphorus is an important element for recognition by TLR9 and therefore removing the negative charge using alkylphosphonates can result in loss or 10 decrease in TLR9 stimulation. The use of phosphonoacetate (PACE) internucleotide linkages between deoxynucleosides in 5' and 3' terminal sequences can significantly increase the TLR9 response; however, the use of thiophosphonoacetate (thioPACE) internucleotide linkages between deoxynucleo- 15 sides in 5' and 3' terminal sequences can result in loss or decrease in TLR9 stimulation. In certain embodiments, use of sugar modifications that the favor C3'-endo conformation such as 2'-O-methyl modifications can be incorporated at 5' and 3' termini to decrease the TLR9 response. TLR 7 and 20 TLR8 can be stimulated by molecules containing 7-deazaguanine and by single-stranded RNA (see, e.g., Heil et al. (2004) Science, 303, 1526-9). TLR3 has been implicated in cellular immunoresponses to virus-derived double-stranded RNA. In certain embodiments, these TLR responses can be 25 mitigated for example by using 2'-O-methyl modifications, modified phosphodiester linkages containing sulfur, or modifications that decrease internucleotide negative charge such as methylphosphonate and/or phosphonoacetate internucleotide linkages.

modifications can lower or eliminate the TLR9 response.

In certain embodiments, the modification enhances stability and specificity of the guide RNA relative to a guide RNA without the modification. In certain embodiments, the modification enhances stability and transfection efficiency of the guide RNA relative to a guide RNA without the modification. In certain embodiments, the modification enhances specificity and transfection efficiency of the guide RNA relative to a guide RNA without the modification. In certain embodiments, the modification enhances the overall efficacy of the guide RNA relative to a guide RNA without 40 the modification.

# C. Guide RNA with a Combination of Modifications

In one aspect, the present technology provides a guide RNA having a combination of two or more modifications.

In certain embodiments, the two modifications are on the same nucleotide (for example, one nucleotide comprises a 2'-O-methyl and a 3'-thiophosphonoacetate moiety). In other 50 embodiments, the two modifications are on two different nucleotides (for example, one nucleotide has a 2-thioU base and another nucleotide has a 2'-O-methyl group).

In certain embodiments, each modification in the guide RNA is the same. In certain embodiments, at least one 55 modification in the guide RNA is different from at least one other modification in the guide RNA. In certain embodiments, a single nucleotide within the guide RNA possesses two or more modifications.

In certain embodiments, the guide RNA comprises a 60 combination of different types of modifications, and at least one type in the combination exists in multiple places in the guide RNA In certain embodiments, at least one type in the combination appears 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20 times in the guide RNA.

In certain embodiments, at least one type of the modifications in the combination appears in two or more modified 36

nucleotides. In certain embodiments, at least one type of the modifications in the combination appears in three or more modified nucleotides. In certain embodiments, the modified nucleotides are not arranged contiguously in the sequence, or at least not entirely, as one or more unmodified nucleotides may intercede. In certain embodiments, the modified nucleotides are arranged contiguously. In certain embodiments, the guide RNA comprises a stretch of contiguous modified nucleotides of the same type. In certain embodiments, the stretch has at least 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, or 40 modified nucleotides.

In certain embodiments, at least one type of the modifications in the combination is within the 5' portion of the guide RNA. In certain embodiments, at least one type of the modifications in the combination is within the first five (5) nucleotides of the 5' portion of the guide RNA. In certain embodiments, at least one type of the modifications in the combination is within the first three (3) nucleotides of the 5' portion of the guide RNA. In certain embodiments, at least one type of the modifications in the combination is within the 3' portion of the guide RNA. In certain embodiments, at least one type of the modifications in the combination is within the last five (5) nucleotides of the 3' portion of the guide RNA. In certain embodiments, at least one type of the modifications in the combination is within the last three (3) nucleotides of the 3' portion of the guide RNA. In certain embodiments, at least one type of the modifications in the combination is within the internal region (i.e., between the 5' end and the 3' end) of the guide RNA.

In certain embodiments, at least one type of the modifications in the combination is incorporated in the 5' portion or 3' portion of the guide RNA, particularly within the first 5 or 10 nucleotides of the 5' portion or within the last 5 or 10 nucleotides of the 3' portion to, for example, protect the RNA from degradation by nucleases or for other purposes. In certain embodiments, at least one type of the modifications in the combination is in the 5' portion and at least one type of the modifications in the combination is in the 3' portion of the guide RNA, particularly within the first 5 or 10 nucleotides of the 5' portion and within the last 5 or 10 nucleotides of the 3' portion to, for example, protect the RNA from degradation by nucleases or for other purposes. In certain embodiments, a guide RNA comprises 20 or fewer, alternatively 15 or fewer, alternatively 15 or fewer, alternatively 10 or fewer, alternatively 5 or fewer, alternatively 3 or fewer deoxyribonucleotide residues in the 5' portion of the guide RNA.

In certain embodiments, at least one type of the modifications in the combination is within the crRNA segment of the guide RNA. In certain embodiments, at least one type of the modifications in the combination is within the guide sequence of the crRNA. In certain embodiments, at least one type of the modifications in the combination is within the first five (5) nucleotides of the crRNA segment. In certain embodiments, at least one type of the modifications in the combination is within the first three (3) nucleotides of the crRNA segment. In certain embodiments, at least one type of the modifications in the combination is within the tracrRNA segment of the guide RNA. In certain embodiments, at least one type of the modifications in the combination is within the last five (5) nucleotides of the tracrRNA segment of the guide RNA. In certain embodiments, at least one type of the modifications in the combination is within the last three (3) nucleotides of the tracrRNA segment of the guide RNA.

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In certain embodiments, a first type of modification in the combination is within the 5' portion of the guide RNA and a second type of modification in the combination is within the internal region (i.e., between the 5' end and the 3' end) of the guide RNA. In certain embodiments, the first type of 5 modification is within the first five (5) nucleotides of the 5' portion of the guide RNA. In certain embodiments, the first type of modification is within the first three (3) nucleotides of the 5' portion of the guide RNA.

In certain embodiments, a first type of modification in the 10 combination is within the internal region (i.e., between the 5' end and the 3' end) of the guide RNA and a second type of modification in the combination is within the 3' portion of the guide RNA. In certain embodiments, the second type of modification is within the last five (5) nucleotides of the 3' 15 portion of the guide RNA. In certain embodiments, the second type of modification is within the last three (3) nucleotides of the 3' portion of the guide RNA.

In certain embodiments, a first type of modification in the combination is within the 5' portion of the guide RNA and 20 in the combination comprises an end modification, such as a second type of modification in the combination is within the 3' portion of the guide RNA. In certain embodiments, the first type of modification is within the first five (5) nucleotides of the 5' portion of the guide RNA. In certain embodiments, the first type of modification is within the first three 25 in the combination comprises a modified base. In certain (3) nucleotides of the 5' portion of the guide RNA. In certain embodiments, the second type of modification is within the last five (5) nucleotides of the 3' portion of the guide RNA. In certain embodiments, the second type of modification is within the last three (3) nucleotides of the 3' portion of the 30 guide RNA.

In certain embodiments, a first type of modification in the combination is within the 5' portion of the guide RNA, a second type of modification in the combination is within the internal region (i.e., between the 5' end and the 3' end) of the 35 guide RNA, and a third type of modification in the combination is within the 3' portion of the guide RNA. In certain embodiments, the first type of modification is within the first five (5) nucleotides of the 5' portion of the guide RNA. In certain embodiments, the first type of modification is within 40 the first three (3) nucleotides of the 5' portion of the guide RNA. In certain embodiments, the third type of modification is within the last five (5) nucleotides of the 3' portion of the guide RNA. In certain embodiments, the third type of modification is within the last three (3) nucleotides of the 3' 45 portion of the guide RNA.

In certain embodiments, a first type of modification in the combination is within the crRNA segment of the guide RNA and a second type of modification in the combination is within the tracr segment. In certain embodiments, the first 50 type of modification is within the guide sequence of the crRNA. In certain embodiments, the first type of modification is within the first five (5) nucleotides of the crRNA segment. In certain embodiments, the first type of modification is within the first three (3) nucleotides of the crRNA 55 segment. In certain embodiments, the second type of modification is within the last five (5) nucleotides of the tracrRNA segment of the guide RNA. In certain embodiments, the second type of modification is within the last three (3) nucleotides of the tracrRNA segment of the guide 60

In certain embodiments, a first type and a second type of modification in the combination are within the crRNA segment of the guide RNA. In certain embodiments, the first type of modification is within the guide sequence of the 65 crRNA. In certain embodiments, the first type of modification is within the first five (5) nucleotides of the crRNA

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segment. In certain embodiments, the first type of modification is within the first three (3) nucleotides of the crRNA

In certain embodiments, a first type and a second type of modification in the combination are within the crRNA segment of the guide RNA and a third type of modification in the combination is within the tracr segment. In certain embodiments, the first type of modification is within the guide sequence of the crRNA. In certain embodiments, the first type of modification is within the first five (5) nucleotides of the crRNA segment. In certain embodiments, the first type of modification is within the first three (3) nucleotides of the crRNA segment. In certain embodiments, the third type of modification is within the last five (5) nucleotides of the tracrRNA segment of the guide RNA. In certain embodiments, the third type of modification is within the last three (3) nucleotides of the tracrRNA segment of the guide RNA.

In certain embodiments, at least one of the modifications a 5' end modification or a 3' end modification as described above. In certain embodiments, the end modification comprises dimethoxytrityl.

In certain embodiments, at least one of the modifications embodiments, the modified gRNA comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, or 40 modified bases. In other embodiments, the modified gRNA comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified bases. In certain embodiments, all bases in a gRNA are modified.

In certain embodiments, at least one of the modifications in the combination comprises a modified sugar. In certain embodiments, the modified gRNA comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, or 40 modified sugars. In other embodiments, the modified gRNA comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified sugars. In certain embodiments, all sugars in a gRNA are modified.

In certain embodiments, at least one of the modifications in the combination comprises a modified backbone (i.e., an internucleotide linkage other than a natural phosphodiester). In certain embodiments, the modified gRNA comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, or 40 modified internucleotide linkages. In other embodiments, the modified gRNA comprises at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130 or 140 modified internucleotide linkages. In certain embodiments, all internucleotide linkages in a gRNA are modified.

In certain embodiments, at least one of the modifications in the combination comprises a 2'-O-methyl, a 2'-fluoro, a 2'-amino, a 2'-deoxy, a 2'-arabino, a 2'-F-arabino, a 2-thiouracil, a 2-aminoadenine, a 5-methylcytosine, a 5-aminoallyluracil, a Z base, a 3'-phosphorothioate, a 3'-phosphonoacetate, a 3'-phosphonoacetate ester, a 3'-thiophosphonoacetate, a 3'-thiophosphonoacetate ester, a 3'-methylphosphonate, a 3'-boranophosphonate, a 3'-phosphorodithioate, or combinations thereof. In certain embodiments, at least one of the modifications in the combination comprises a 2'-O-methyl, a 2'-deoxy, a Z base, a phosphorothioate internucleotide linkage, a phosphonoacetate internucleotide linkage, a thiophosphonoacetate internucleotide linkage, or combinations thereof. In certain embodiments, at least one of the modifications in the combination comprises a 2'-F, a

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2-thioU, a 4-thioU, a 2-aminoA, a 5-methylC, a 5-methylU, a 5-aminoallylU, or combinations thereof. In certain embodiments, at least one of the modifications in the combination is an "end" modification such as terminal phosphate, a PEG, a terminal amine, a terminal linker such as a hydrocarbon linker, a substituted hydrocarbon linker, a squarate linker, a triazolo linker, an internal linker such as 2-(4-butylamidofluorescein)propane-1,3-diol phodiester) linker, a linker conjugated to a dye, a linker conjugated to a non-fluorescent label, a linker conjugated to a tag or a linker conjugated to a solid support such as for example a bead or microarray. In certain embodiments, at least two of the modifications in the combination comprise a 2'-O-methyl nucleotide and phosphorothioate internucleotide linkage, a 2'-O-methyl nucleotide and phosphonoacetate internucleotide linkage, or a 2'-O-methyl nucleotide and thiophosphonoacetate internucleotide linkage. In certain embodiments, at least two of the modifications in the combination comprise a 2'-O-methyl nucleotide and phospho- 20 nocarboxylate internucleotide linkage, a 2'-O-methyl nucleotide and phosphonocarboxylate ester internucleotide linkage, a 2'-O-methyl nucleotide and thiophosphonocarboxylate internucleotide linkage, a 2'-O-methyl nucleotide and thiophosphonocarboxylate ester internucleotide linkage, 25 or combinations thereof. In other embodiments, the modifications in the combination further comprise a 2-thiouracil, 2-thiocytosine, 4-thiouracil, 6-thioguanine, 2-aminoadenine, 2-aminopurine, pseudouracil, inosine, 7-deazaguanine, 7-deaza-8-azaguanine, 7-deazaadenine, 7-deaza-8-azaade- 30 nine, 5-methylcytosine, 5-methyluracil, 5-hydroxymethylcytosine, 5-hydroxymethyluracil, 5,6-dehydrouracil, 5-pro-5-propynyluracil, pynylcytosine, 5-ethynylcytosine, 5-ethynyluracil, 5-allyluracil, 5-allylcytosine, 5-aminoallyluracil, 5-aminoallyl-cytosine, or an abasic nucleotide.

In certain embodiments, at least one of the modifications in the combination comprises a 2'-O-methyl-3'-phosphorothioate. In certain embodiments, at least one of the modifications in the combination comprises a 2'-O-methyl-3'phosphonoacetate. In certain embodiments, at least one of  $^{40}$ the modifications in the combination comprises a 2'-Omethyl-3'-thiophosphonoacetate. In certain embodiments, at least one of the modifications in the combination comprises a 2'-halo-3'-phosphorothioate. In certain embodiments, at least one of the modifications in the combination comprises a 2'-halo-3'-phosphonoacetate. In certain embodiments, at least one of the modifications in the combination comprises a 2'-halo-3'-thiophosphonoacetate. In certain embodiments, at least one of the modifications in the combination comprises a 2'-fluoro-3'-phosphorothioate. In certain embodiments, at least one of the modifications in the combination comprises a 2'-fluoro-3'-phosphonoacetate. In certain embodiments, at least one of the modifications in the combination comprises a 2'-fluoro-3'-thiophosphonoacetate. are represented in FIG. 6 and FIG. 7 respectively and are incorporated herein by reference.

In certain embodiments, the guide RNA comprises an oligonucleotide represented by Formula (III) or Formula (IV):

wherein Q and W each independently represent a nucleotide or a stretch of nucleotides of the oligonucleotide com40

prising at least one modification and Y and X each independently represent an unmodified portion of the oligonucleotide.

In certain embodiments, W is within the 5' portion of the guide RNA. In certain embodiments, W is at least partially within the first five (5) nucleotides of the 5' portion of the guide RNA. In certain embodiments, W is at least partially within the first three (3) nucleotides of the 5' portion of the guide RNA. In certain embodiments, W is within the internal region (i.e., between the 5' end and the 3' end) of the guide

In certain embodiments, Q is within the 3' portion of the guide RNA. In certain embodiments, Q is at least partially within the last five (5) nucleotides of the 3' portion of the guide RNA. In certain embodiments, Q is at least partially within the last three (3) nucleotides of the 3' portion of the guide RNA. In certain embodiments, Q is within the internal region (i.e., between the 5' end and the 3' end) of the guide RNA.

In certain embodiments, W comprises an end modification as described above, such as a 5' end or a 3' end modification. In certain embodiments, the end modification comprises dimethoxytrityl.

In certain embodiments, at least one of W or Q comprises a modified base as described above. In certain embodiments, at least one of W or Q comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50 modified bases. In certain embodiments, at least one of W or Q comprises more than one modified base, such as 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20 modified bases.

In certain embodiments, at least one of W or Q comprises 35 a modified sugar as described above. In certain embodiments, at least one of W or Q comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50 modified sugars. In certain embodiments, at least one of W or Q comprises more than one, such as 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20 modified sugars.

In certain embodiments, at least one of W or Q comprises a modified backbone (i.e., an internucleotide linkage other than a phosphodiester) as described above. In certain embodiments, at least one of W or Q comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50 modified internucleotide linkages. In certain embodiments, at least one of W or Q comprises more than one, such as 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20 modified internucleotide linkages.

In certain embodiments, at least one of W or Q comprises Possible combinations of at least two or three modifications 55 a 2'-O-methyl nucleotide, a 2'-F nucleotide, a 2'-amino nucleotide, a 2'-deoxy nucleotide, a 2-thiouridine nucleotide, a 2-aminoadeosine nucleotide, a 6-thioguanosine nucleotide, a 5-methylcytidine nucleotide, a 5-aminoallyluridine nucleotide, a Z nucleotide, a 3'-phosphorothioate internucleotide linkage, a 3'-phosphorothioate internucleotide linkage, a 3'-phosphonoacetate internucleotide linkage, a 3'-phosphonoacetate ester internucleotide linkage, a 3'-thiophosphonoacetate internucleotide linkage, a 3'-thiophosphonoacetate ester internucleotide linkage, a 3'-methylphosphonate internucleotide linkage, a 3'-boranophosphonate internucleotide linkage, a 3'-phosphorodithioate internucleotide linkage, or combinations thereof.

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In certain embodiments, at least one of W or Q comprises a 2'-O-methyl and a 3'-phosphorothioate group on the same nucleotide. In certain embodiments, at least one of W or Q comprises a 2'-O-methyl and a 3'-phosphonoacetate group linkage on the same nucleotide. In certain embodiments, at least one of W or Q comprises a 2'-O-methyl and 3'-thio-phosphonoacetate group on the same nucleotide. In certain embodiments, at least one of W or Q comprises a 2'-F and a 3'-phosphorothioate group on the same nucleotide. In certain embodiments, at least one of W or Q comprises a 2'-F and a 3'-phosphonoacetate group linkage on the same nucleotide. In certain embodiments, at least one of W or Q comprises a 2'-F and 3'-thiophosphonoacetate group on the same nucleotide.

In certain embodiments, at least one of W or Q comprises 15 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20 modified nucleotides. In certain embodiments, each of the modified nucleotides within at least one of W or Q comprises the same modification or modifications. In certain embodiments. W comprises a modified nucleotide that is 20 different than a modified nucleotide in Q. In certain embodiments, at least one of W or Q comprises two or more modified nucleotides. In certain embodiments, at least one of W or Q comprises three or more modified nucleotides. In certain embodiments, the modified nucleotides are not 25 arranged contiguously in the sequence, or at least not entirely, as one or more unmodified nucleotides may intercede. In certain embodiments, the modified nucleotides are arranged contiguously. In certain embodiments, at least one of W or Q comprises at least one contiguous stretch of 30 modified nucleotides. In certain embodiments, at least one of W or Q comprises a contiguous stretch of at least three (3) modified nucleotides. In certain embodiments, at least one of W or Q comprises a contiguous stretch of at least four (4) modified nucleotides. In certain embodiments, at least one of 35 W or Q comprises a contiguous stretch of at least five (5) modified nucleotides.

In certain embodiments, the guide RNA comprises a nucleotide sequence of Formula (V) or Formula (VI):

$$M_m N_n M'_m N'_{n'}$$
 (Formula V); or

$$\mathbf{M}_{m}\mathbf{N}_{n}\mathbf{M'}_{m'}\mathbf{N'}_{n}\mathbf{M''}_{m''} \tag{Formula VI}$$

wherein each M independently represents a modified ribonucleotide;

wherein each N independently represents an unmodified ribonucleotide:

wherein each M' independently represents a modified ribonucleotide;

wherein each N' independently represents an unmodified 50 ribonucleotide;

wherein each M" independently represents a modified ribonucleotide:

wherein m is an integer between 0 and 40, n is an integer between 0 and 130, m' is an integer between 0 and 10, n' is 55 an integer between 0 and 130, m" is an integer between 0 and 10, provided that m+m'+m" is greater than or equal to 1 and 50<m+n+m'+n'+m"≤150.

In certain embodiments, each M is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, 60 a 2'-O-methyl-3'-P(S) ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, a 2'-O-methyl-3'-thioPACE ribonucleotide, and a 2'-deoxynucleotide. In certain embodiments, each M is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, and a 2'-O-methyl-3'-PACE ribonucleotide, and a 2'-O-methyl-3'-thioPACE ribonucleotide. In

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certain embodiments, each M is independently selected from the group consisting of a 2'-O-methyl-3'-PACE ribonucle-otide and a 2'-O-methyl-3'-thioPACE ribonucleotide. In certain embodiments, where m>1, any given M is the same or different from any other M. In certain embodiments, where m>1, each M comprises the same modification or modifications.

In certain embodiments, each M' is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-P(S) ribonucleotide, a 2'-Omethyl-3'-PACE ribonucleotide, a 2'-O-methyl-3'-thioPACE ribonucleotide, and a 2'-deoxynucleotide. In certain embodiments, each M' is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-P(S) ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, and a 2'-O-methyl-3'-thioPACE ribonucleotide. In certain embodiments, each M' is independently selected from the group consisting of a 2'-O-methyl-3'-PACE ribonucleotide and a 2'-O-methyl-3'-thioPACE ribonucleotide. In certain embodiments, where m'>1, any given M' is the same or different from any other M'. In certain embodiments, where m'>1, each M' comprises the same modification or modifications.

In certain embodiments, each M" is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-P(S) ribonucleotide, a 2'-Omethyl-3'-PACE ribonucleotide, a 2'-O-methyl-3'-thioPACE ribonucleotide, and a 2'-deoxynucleotide. In certain embodiments, each M" is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-P(S) ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, and a 2'-O-methyl-3'-thioPACE ribonucleotide. In certain embodiments, each M" is independently selected from the group consisting of a 2'-O-methyl-3'-PACE ribonucleotide and a 2'-O-methyl-3'-thioPACE ribonucleotide. In certain embodiments, where m">1, any given M" is the same or different from any other M". In certain embodiments, where m'>1, each M" comprises the same modification or modifications.

In certain embodiments, each M is a 2'-O-methyl-3'-PACE ribonucleotide; m is selected from an integer between 1 and 10; each N is independently selected from the group consisting of A, U, C, and G; n is selected from an integer between 10 and 130; each M' is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-P(S)ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, a 2'-O-methyl-3'-thioPACE ribonucleotide, a 2'-deoxynucleotide, and a Z nucleotide; m' is selected from an integer between 1 and 10; each N is independently selected from the group consisting of A, U, C, and G; and n' is selected from an integer between 0 and 130. In certain embodiments, each M' is a 2'-O-methyl-3'-PACE ribonucleotide. In certain embodiments, each M' is a 2'-O-methyl-3'thioPACE ribonucleotide. In certain embodiments, each M' is a 2'-O-methyl ribonucleotide. In certain embodiments, each M' is a 2'-O-methyl-3'-P(S) ribonucleotide. In certain embodiments, each M' is a Z nucleotide.

In certain embodiments, each M is a 2'-O-methyl-3'-thioPACE ribonucleotide; m is selected from an integer between 1 and 10; each N is independently selected from the group consisting of A, U, C, and G; n is selected from an integer between 10 and 130; each M' is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, a 2'-O-methyl-3'-thioPACE ribonucleotide, a 2'-deoxynucleotide, and a Z nucleotide; m' is selected from an integer between 1 and 10; each N is

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independently selected from the group consisting of A, U, C, and G; and n' is selected from an integer between 0 and 130. In certain embodiments, each M' is a 2'-O-methyl-3'-PACE ribonucleotide. In certain embodiments, each M' is a 2'-O-methyl-3'-thioPACE ribonucleotide. In certain embodiments, each M' is a 2'-O-methyl ribonucleotide. In certain embodiments, each M' is a 2'-O-methyl-3'-P(S) ribonucleotide. In certain embodiments, each M' is a Z nucleotide.

In certain embodiments, each M is a 2'-O-methyl-3'-P(S) ribonucleotide; m is selected from an integer between 1 and 10 10; each N is independently selected from the group consisting of A, U, C, and G; n is selected from an integer between 10 and 130; each M' is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-P(S) ribonucleotide, a 2'-O-methyl-3'-PACE 15 ribonucleotide, a 2'-O-methyl-3'-thioPACE ribonucleotide, a 2'-deoxynucleotide, and a Z nucleotide; m' is selected from an integer between 1 and 10; each N is independently selected from the group consisting of A, U, C, and G; and n' is selected from an integer between 0 and 130. In certain 20 embodiments, each M' is a 2'-O-methyl-3'-PACE ribonucleotide. In certain embodiments, each M' is a 2'-O-methyl-3'thioPACE ribonucleotide. In certain embodiments, each M' is a 2'-O-methyl ribonucleotide. In certain embodiments, each M' is a 2'-O-methyl-3'-P(S)ribonucleotide. In certain 25 embodiments, each M' is a Z nucleotide.

In certain embodiments, each M is independently selected from the group consisting of a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-P(S) ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, a 2'-O-methyl-3'-thioPACE ribo- 30 nucleotide, a 2'-deoxynucleotide, and a Z nucleotide; m is selected from an integer between 0 and 10; each N is independently selected from the group consisting of A, U, C, and G; n is selected from an integer between 10 and 15; each M' is a 2'-O-methyl ribonucleotide; m' is selected from an 35 integer between 1 and 5; each N is independently selected from the group consisting of A, U, C, and G; and n' is selected from an integer between 0 and 130. In certain embodiments, each M is a 2'-O-methyl-3'-PACE ribonucleotide. In certain embodiments, each M is a 2'-O-methyl-3'- 40 thioPACE ribonucleotide. In certain embodiments, each M is a 2'-O-methyl ribonucleotide. In certain embodiments, each M is a 2'-O-methyl-3'-P(S)ribonucleotide. In certain embodiments, m is 0; n is selected from an integer between 10 and 15, m' is selected from an integer between 1 and 5; 45 and n' is selected from an integer between 0 and 130.

In certain embodiments, at least one of the modifications in the combination is a stability-altering modification. In certain embodiments, at least one of the modifications in the combination increases nuclease resistance of the guide RNA 50 relative to a guide RNA without the modification, thus it enhances the stability of the guide RNA.

In certain embodiments, at least one of the modifications in the combination is a stability-enhancing modification as described above.

In certain embodiments, at least one of the modifications in the combination is a specificity-altering modification as described above.

In certain embodiments, at least one of the modifications in the combination alters RNA base pairing. In certain 60 embodiments, at least one of the modifications in the combination lowers the Tm of a base pairing interaction as described above. In certain embodiments, at least one of the modifications in the combination raises the Tm of a base pairing interaction as described above.

In certain embodiments, at least one of the modifications in the combination alters transfection efficiency of the guide 44

RNA relative to a guide RNA without the modification. In certain embodiments, at least one of the modifications in the combination increases transfection efficiency of the guide RNA relative to a guide RNA without the modification. In certain embodiments, at least one of the modifications in the combination decreases transfection efficiency of the guide RNA relative to a guide RNA without the modification. In certain embodiments, at least one of the transfection-increasing modifications in the combination comprises a phosphonoacetate alkyl ester internucleotide linkage, such as a phosphonoacetate methyl ester internucleotide linkage.

In certain embodiments, at least one of the modifications in the combination enhances stability and specificity of the guide RNA relative to a guide RNA without the modification. In certain embodiments, at least one of the modifications in the combination enhances stability and transfection efficiency of the guide RNA relative to a guide RNA without the modification. In certain embodiments, at least one of the modifications in the combination enhances specificity and transfection efficiency of the guide RNA relative to a guide RNA without the modification.

In certain embodiments, at least one of the modifications in the combination alters the secondary structure of the guide RNA. This modification alters the base-pairing of any of the RNA/RNA internal duplexes in the guide RNA. Some of these modifications increase the base pairing of the RNA/RNA structure or alternatively increase the Tm of the RNA/RNA duplex, whereas other modifications decrease the base pairing (or Tm) of the RNA/RNA duplex or duplexes. Such modifications include base modified nucleotides, particularly UNA nucleotides such as the 2-thiouridine and 2-aminoadenosine pair, the Z/P nucleotide pair, the isoC/isoG pair, the 6-thioG/5-methylpyrimidine pair, and nucleotides with modifications on the sugar or the internucleotide linkages as discussed before.

In certain embodiments, the combination includes at least one modification or a set of modifications that increases nucleases resistance (i.e., stability) with at least one modification or a set of modifications that increases specificity (i.e., reduces off-target effects). In certain embodiments, the combination includes at least one modification or a set of modifications that increases nucleases resistance (i.e., stability) with at least one modification or a set of modifications that raises the Tm of some bases pairing in the guide RNA. In certain embodiments, the combination includes at least one modification or a set of modifications that increases nucleases resistance (i.e., stability) with at least one modification or a set of modifications that lowers the Tm of some bases pairing of the guide RNA. In certain embodiments, the combination includes at least one modification or a set of modifications that increases nuclease resistance (i.e., stability), at least one modification or a set of modifications that increases the Tm of some bases paring in the guide RNA, and at least one modification or a set of modifications that decreases the Tm of some base paring elsewhere in the guide RNA. In certain embodiments, the combination includes at least one modification or a set of modifications that increases nuclease resistance (i.e., stability) and at least one modification or a set of modifications that increases the binding of the guide RNA to Cas protein. In certain embodiments, the combination includes at least one modification or a set of modifications that increases nuclease resistance (i.e., stability) and at least one modification or a set of modifications that decreases the binding of the guide RNA to Cas protein.

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In certain embodiments, the guide RNA comprises a combination of the different types of modifications.

#### D. Guide RNA Structure

In certain embodiments, the guide RNA is able to form a complex with a CRISPR-associated-protein. In certain embodiments, the CRISPR-associated protein is provided by or is derived from a CRISPR-Cas type II system, which has an RNA-guided polynucleotide binding and/or nuclease 10 activity. In certain embodiments, the CRISPR-associated protein is Cas9, a Cas9 mutant, or a Cas9 variant. In certain embodiments, the CRISPR-associated protein is the Cas9 nuclease from Streptococcus pyogenes. In certain embodiments, the CRISPR-associated protein is the Cas9 nuclease 15 from Streptococcus thermophilus. In certain embodiments, the CRISPR-associated protein is the Cas9 nuclease from Staphylococcus aureus. In certain embodiments, the synthetic guide RNA or a synthetic guide RNA:CRISPR-associated protein complex maintains functionality of natural 20 guide RNA or a complex that does not have modified nucleotides. In certain embodiments, the functionality includes binding a target polynucleotide. In certain embodiments, the functionality includes nicking a target polynucleotide. In certain embodiments, the functionality includes 25 cleaving a target polynucleotide. In certain embodiments, the target polynucleotide is within a nucleic acid in vitro. In certain embodiments, the target polynucleotide is within the genome of a cell in vivo or in vitro (such as in cultured cells or cells isolated from an organism). In certain embodiments, 30 the target polynucleotide is a protospacer in DNA.

In certain embodiments, the crRNA segment comprises from 25 to 80 nucleotides. In certain embodiments, the crRNA segment comprises a guide sequence that is capable of hybridizing to a target sequence. In certain embodiments, 35 the guide sequence is complementary to the target sequence or a portion thereof. In certain embodiments, the guide sequence comprises from 15 to 30 nucleotides. In certain embodiments, the crRNA segment comprises a stem sequence. In certain embodiments, the stem sequence com- 40 prises from 10 to 50 nucleotides. In certain embodiments, the crRNA segment comprises a 5'-overhang sequence. In certain embodiments, the 5'-overhang sequence comprises from 1 to 10 nucleotides, alternatively 1 to 5 nucleotides, alternatively 1, 2 or 3 nucleotides. In certain embodiments, 45 the crRNA comprises both (i) a guide sequence that is capable of hybridizing to a target sequence and (ii) a stem sequence. In certain embodiments, the crRNA comprises (i) a 5'-overhang sequence, (ii) a guide sequence that is capable of hybridizing to a target sequence, and (iii) a stem 50 sequence. In certain embodiments wherein the crRNA segment comprises a stem sequence, the tracrRNA segment comprises a nucleotide sequence that is partially or completely complementary to the stem sequence of the crRNA segment. In certain embodiments, the tracrRNA segment 55 comprises at least one more duplex structure.

In certain embodiments, the guide RNA is a single guide RNA. In certain embodiments, the guide RNA is a single guide RNA, wherein the crRNA segment and the tracrRNA segment are linked through a loop L. In certain embodiments, the loop L comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 nucleotides. In certain embodiments, the loop L comprises a nucleotide sequence of GNRA, wherein N represents A, C, G, or U and R represents A or G. In certain embodiments, the loop L comprises a nucleotide sequence of GAAA. In certain embodiments, the guide RNA comprises more than one loop.

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The guide RNA comprises a 5' portion (i.e., the 5' half) and a 3' portion (i.e., the 3' half). In certain embodiments, the crRNA segment is 5' (i.e., upstream) of the tracrRNA segment. In certain embodiments, the tracrRNA segment is 5' relative to the crRNA segment.

In certain embodiments, the guide RNA comprises at least two separate RNA strands, for example, a crRNA strand and a separate tracrRNA strand. See, for example, FIG. 5A. In certain embodiments, each of the strands is a synthetic strand comprising one or more modifications. In certain embodiments, at least one of the strands is a synthetic strand comprising one or more modifications. In certain embodiments, the strands function together to guide binding, nicking, or cleaving of a target polynucleotide by a Cas protein, such as Cas9. In certain embodiments, the crRNA sequence and the tracrRNA sequence are on separate stands and hybridize to each other via two complementary sequences to form a stem or duplex.

In certain embodiments, the guide RNA is a single guide RNA comprising a crRNA sequence and a tracrRNA sequence. See, for example, FIG. 5B. In certain embodiments, the crRNA sequence and the tracrRNA sequence are connected by a loop sequence or "loop." In certain embodiments, a single guide RNA comprises a 5' portion and a 3' portion, wherein the crRNA sequence is upstream of the tracrRNA sequence.

In certain embodiments, the total length of the two RNA pieces can be about 50-220 (e.g., about 55-200, 60-190, 60-180, 60-170, 60-160, 60-150, 60-140, 60-130, and 60-120) nucleotides in length, such as about 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, or 220 nucleotides in length. Similarly, the single guide RNA (e.g., FIG. 5B) can be about 50-220 (e.g., about 55-200, 60-190, 60-180, 60-170, 60-160, 60-150, 60-140, 60-130, and 60-120) nucleotides in length, such as about 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, or 220 nucleotides in length.

As shown in FIGS. 5A and 5B, the synthetic guide RNA comprises (i) a crRNA sequence that comprises (a) a guide sequence (e.g., segment G<sub>1</sub>-G<sub>n</sub>, where each G represents a nucleotide in the guide sequence) capable of hybridizing to a target sequence in a nucleic acid, (b) a first stem sequence (e.g., segment  $X_1$ - $X_n$ , where each X represents a nucleotide in the first stem sequence) capable of hybridizing partially or completely to a second stem sequence, and, optionally (c) a 5'-overhang sequence (e.g., segment O<sub>1</sub>-O<sub>2</sub>, where each 0 represents a nucleotide in the overhang sequence), and (ii) a tracrRNA sequence that comprises the second stem sequence (e.g., segment  $Y_1$ - $Y_n$ , where each Y represents a nucleotide in the second stem sequence). The tracrRNA sequence further comprises segment  $T_1$ - $T_n$ , where each T represents a nucleotide in the tracrRNA sequence. The synthetic guide RNA shown in FIG. 5A includes one or more modifications. Likewise, the synthetic guide RNA shown in FIG. 5B includes one or more modifications. In certain embodiments, the modification is located at any point along the length of the crRNA, the tracrRNA, or the single guide RNA comprising a crRNA segment, a tracrRNA segment, and, optionally, a loop. In certain embodiments, any nucleotide represented by O, G, X, Y, or T in the synthetic guide RNA shown in FIGS. 5A and 5B may be a modified nucleotide. The guide RNA shown in FIG. 5B represents a single guide RNA (sgRNA) where the crRNA segment and the tracrRNA segment are connected by a loop having the sequence GNRA, wherein N represents A, C, G, or U, and R represents A or G.

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In certain embodiments, the crRNA segment of the guide RNA is 25-70 (e.g., 30-60, 35-50, or 40-45) nucleotides in length. In certain embodiments, the guide sequence is 12-30 (e.g., 16-25, 17-20, or 15-18) nucleotides in length. In some embodiments, a 5' portion of the crRNA does not hybridize or only partially hybridizes with the target sequence. For example, there can be a 5'-overhang on the crRNA segment.

In certain embodiments, the single guide RNA comprises a central portion including the stem sequence of the crRNA segment, the stem sequence of the tracrRNA segment, and, optionally, a loop that covalently connects the crRNA segment to the tracrRNA segment. In certain embodiments, the central segment of the single guide RNA is 8-60 (e.g., 10-55, 10-50, or 20-40) nucleotides in length.

In certain embodiments, the tracrRNA segment of the 15 guide RNA is 10-130 (e.g., 10-125, 10-100, 10-75, 10-50, or 10-25) nucleotides in length. In certain embodiments, the tracrRNA segment includes one or more hairpin or duplex structures in addition to any hairpin or duplex structure in the central segment.

In certain embodiments, the tracrRNA is truncated compared to a reference tracrRNA, such as a naturally existing mature tracrRNA. A range of lengths has been shown to function in both the separate type (FIG. 5A) and the chimeric sgRNA type (FIG. 5B). For example, in certain 25 embodiments, tracrRNA may be truncated from its 3' end by at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35 or 40 nts. In certain embodiments, the tracrRNA molecule may be truncated from its 5' end by at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75 or 80 30 nts. In certain embodiments, the tracrRNA molecule may be truncated from both the 5' and 3' end, e.g., by at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15 or 20 nts from the 5' end and at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35 or 40 nts from Mali et al. (2013) Science, 339:6121, 823-6; Cong et al. (2013) Science, 339:6121, 819-23; and Hwang et al. (2013) Nat. Biotechnol. 31:3, 227-9; Jinek et al. (2013) eLife, 2, e00471. In certain embodiments, the tracrRNA is untrun-

In certain embodiments, the disclosed modifications are in the crRNA segment or the tracrRNA segment or both. In certain embodiments, the disclosed modifications are in the guide sequence of the crRNA segment. In certain embodiments, the disclosed modifications are in the stem sequence 45 of the crRNA segment. In certain embodiments, the disclosed modifications are in the 5'-overhang sequence of the crRNA segment. In certain embodiments, the disclosed modifications are in the stem sequence of the tracrRNA segment. In certain embodiments, the disclosed modifica- 50 tions are in the loop sequence of the guide RNA. In certain embodiments, the disclosed modifications are in the 5' portion of the guide RNA. In certain embodiments, the disclosed modifications are in the 3' portion of the guide RNA. In certain embodiments, the disclosed modifications 55 are in the 5' portion of the guide RNA and the 3' portion of the guide RNA.

#### E. Synthesis of Guide RNA

In certain embodiments, guide RNAs, including single guide RNAs (sgRNAs; see FIGS. 1 and 5B) are produced by chemical synthesis using the art of synthetic organic chemistry. A guide RNA that comprises any nucleotide other than the four predominant ribonucleotides, namely A, C, G, and 65 U, whether unnatural or natural, such as a pseudouridine, inosine or a deoxynucleotide, possesses a chemical modifi-

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cation or substitution at the nucleotide which is chemically/structurally distinct from any of the four predominant nucleotides in RNAs.

The synthetic guide RNAs described herein can be chemically synthesized. For example, the synthetic guide RNAs can be synthesized using TC chemistry by the method described in Dellinger et al. (2011) J. Am. Chem. Soc., 133, 11540, U.S. Pat. No. 8,202,983, and US Patent Application 2010/0076183A1, the contents of which are incorporated by reference in their entireties. "TC chemistry" refers to the composition and methods of using RNA monomeric nucleotide precursors protected on the 2'-hydroxyl moiety by a thionocarbamate protecting group, to synthesize unmodified RNA or modified RNA comprising one or more modified nucleotides. The ability to chemically synthesize relatively long RNAs (as long as 200 mers or more) using TC-RNA chemistry allows one to produce guide RNAs with special features capable of outperforming those enabled by the four predominant ribonucleotides (A, C, G and U). Some syn-20 thetic guide RNAs described herein can also be made using methods known in the art that include in vitro transcription and cell-based expression. For example, 2'-fluoro NTPs can be incorporated into synthetic guide RNAs produced by cell-based expression.

Synthesis of guide RNAs can also be accomplished by chemical or enzymatic synthesis of RNA sequences that are subsequently ligated together by enzymes, or chemically ligated by chemical ligation, including but not limited to cyanogen bromide chemistry, "click" chemistry as published by R. Kumar et al. (2007) *J. Am. Chem. Soc.*, 129, 6859-64, or squarate conjugation chemistry as described by K. Hill in WO2013176844 titled "Compositions and methods for conjugating oligonucleotides."

1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35 or 40 nts from the 3' end. See, e.g., Jinek et al. (2012) *Science*, 337, 816-21; 35 Mali et al. (2013) *Science*, 339:6121, 823-6; Cong et al. (2013) *Science*, 339:6121, 819-23; and Hwang et al. (2013) *Nat. Biotechnol.* 31:3, 227-9; Jinek et al. (2013) *eLife*, 2, e00471. In certain embodiments, the tracrRNA is untruncated.

In certain embodiments, the disclosed modifications are in the crRNA segment or the tracrRNA segment or both. In certain embodiments, the disclosed modifications are in the crgna modified internucleotide linkages, can be used to perform various CRISPR-mediated functions (including but not limited to editing genes, regulating gene expression, cleaving target sequences, and binding to target sequences) in vitro or in vivo, such as in cell-free assays, in intact cells, or in whole organisms. For in vitro or in vivo applications, the RNA can be delivered into cells or whole organisms in any manner known in the art.

Libraries and Arrays

In one aspect, the present invention provides a set or library of multiple guide RNAs. In certain embodiments, the library contains two or more guide RNAs disclosed herein. The library can contain from about 10 to about 10<sup>7</sup> individual members, e.g., about 10 to about 10<sup>2</sup>, about 10<sup>2</sup> to about 10<sup>3</sup>, about 10<sup>3</sup> to about 10<sup>5</sup>, from about 10<sup>5</sup> to about 10<sup>7</sup> members. An individual member of the library differs from other members of the library at least in the guide sequence, i.e., the DNA targeting segment of the gRNA. On the other hand, in certain embodiments, each individual member of a library can contain the same or substantially the same nucleotide sequence for the tracrRNA segment as all the other members of the library. In this way, the library can comprise members that target different polynucleotides or different sequences in one or more polynucleotides.

In certain embodiments, the library comprises at least 10<sup>2</sup> unique guide sequences. In certain embodiments, the library comprises at least 10<sup>3</sup> unique guide sequences. In certain embodiments, the library comprises at least 10<sup>4</sup> unique guide sequences. In certain embodiments, the library comprises at least 10<sup>5</sup> unique guide sequences. In certain embodiments, the library comprises at least 10<sup>6</sup> unique guide sequences. In certain embodiments, the library comprises at least 10<sup>6</sup> unique guide sequences. In certain embodiments, the library comprises at least 10<sup>6</sup> unique guide sequences. In certain embodiments, the library comprises at least 10<sup>6</sup> unique guide sequences.

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prises at least 10<sup>7</sup> unique guide sequences. In certain embodiments, the library targets at least 10 different polynucleotides. In certain embodiments, the library targets at least 10<sup>2</sup> different polynucleotides. In certain embodiments, the library targets at least 10<sup>3</sup> different polynucleotides. In certain embodiments, the library targets at least 10<sup>4</sup> different polynucleotides. In certain embodiments, the library targets at least 10<sup>5</sup> different polynucleotides. In certain embodiments, the library targets at least 10<sup>6</sup> different polynucleotides. In certain embodiments, the library targets at least 10<sup>7</sup> different polynucleotides.

In certain embodiments, the library comprises a collection of guide RNAs having the same sequence and the same modifications in a progressively shifted window that moves across the sequence of the members in the library. In certain 15 embodiments, the windows collectively cover the entire length of the RNA.

In certain embodiments, the library allows one to conduct high-throughput, multi-target genomic manipulations and analyses. In certain embodiments, only the DNA-targeting 20 segments of the guide RNAs are varied, while the Cas protein-binding segment is the same. In certain embodiments, a first portion of the library comprises guide RNAs possessing a Cas-binding segment that recognizes, binds and directs a particular Cas protein and a second portion of the 25 library comprises a different Cas-binding segment that recognizes, binds and directs a different Cas protein (e.g., a Cas protein from a different species), thereby allowing the library to function with two or more orthogonal Cas proteins. In certain embodiments, induced expression of a first 30 orthogonal Cas protein utilizes the portion of the library which interacts with the first orthogonal Cas protein. In certain embodiments, induced expression of a first and second orthogonal Cas protein utilizes the portions of the library which interact with the first and second orthogonal 35 Cas proteins, respectively. In certain embodiments, induced expression of the first and second orthogonal Cas proteins occur at different times. Accordingly, one can carry out large-scale gene editing or gene regulation by specifically manipulating or modifying multiple targets as specified in 40 the library.

In certain embodiments, the library is an "arrayed" library, namely a collection of different features or pools of features in an addressable arrangement. For example, features of an array can be selectively cleaved and transferred 45 to a microtiter plate such that each well in the plate contains a known feature or a known pool of features. In some other embodiments, the library is synthesized in a 48-columns or in a 96-columns microtiter plate format or in a 384-columns plate.

In certain embodiments, synthesis of the guide RNA of this invention may be conducted on a solid support having a surface to which chemical entities may bind. In some embodiments, guide RNAs being synthesized are attached, directly or indirectly, to the same solid support and may 55 form part of an array. An "array" is a collection of separate molecules of known monomeric sequence each arranged in a spatially defined and a physically addressable manner, such that the location of each sequence is known. An "array," or "microarray" used interchangeably herein 60 includes any one-dimensional, two-dimensional or substantially two-dimensional (as well as a three-dimensional) arrangement of addressable regions bearing a particular chemical moiety or moieties (such as ligands, e.g., biopolymers such as polynucleotide or oligonucleotide sequences 65 (nucleic acids), polypeptides (e.g., proteins), carbohydrates, lipids, etc.) associated with that region. An array is "address50

able" when it has multiple regions of different moieties (e.g., different polynucleotide sequences) such that a region (i.e., a "feature" of the array) at a particular predetermined location (i.e., an "address") on the array will detect a particular target or class of targets (although a feature may incidentally detect non-targets of that feature). Array features are typically, but need not be, separated by intervening spaces. The number of features that can be contained on an array will largely be determined by the surface area of the substrate, the size of a feature and the spacing between features. Arrays can have densities of up to several hundred thousand or more features per cm², such as 2,500 to 200,000 features/cm². The features may or may not be covalently bonded to the substrate.

Suitable solid supports may have a variety of forms and compositions and derive from naturally occurring materials, naturally occurring materials that have been synthetically modified, or synthetic materials. Examples of suitable support materials include, but are not limited to, silicas, silicon and silicon oxides, teflons, glasses, polysaccharides such as agarose (e.g., Sepharose® from Pharmacia) and dextran (e.g., Sephadex® and Sephacyl®, also from Pharmacia), polyacrylamides, polystyrenes, polyvinyl alcohols, copolymers of hydroxyethyl methacrylate and methyl methacrylate, and the like. In some embodiments, the solid support is a plurality of beads.

The initial monomer of the guide RNAs to be synthesized on the substrate surface can be bound to a linker which in turn is bound to a surface hydrophilic group, e.g., a surface hydroxyl moiety present on a silica substrate. In some embodiments, a universal linker is used. In some other embodiments, the initial monomer is reacted directly with, e.g., a surface hydroxyl moiety. Alternatively, guide RNAs can be synthesized first according to the present invention, and attached to a solid substrate post-synthesis by any method known in the art. Thus, the present invention can be used to prepare arrays of guide RNAs wherein the oligonucleotides are either synthesized on the array, or attached to the array substrate post-synthesis. Subsequently, the guide RNAs or a pool or a plurality of pools of guide RNAs can optionally and selectively be cleaved from the array substrate and be used as a library or libraries.

#### IV. Cas Proteins

As mentioned above, a functional CRISPR-Cas system also requires a protein component (e.g., a Cas protein, which may be a Cas nuclease) that provides a desired activity, such as target binding or target nicking/cleaving. In certain embodiments, the desired activity is target binding. In certain embodiments, the desired activity is target nicking or target cleaving. In certain embodiments, the desired activity also includes a function provided by a polypeptide that is covalently fused to a Cas protein, as disclosed herein. In certain embodiments, the desired activity also includes a function provided by a polypeptide that is covalently fused to a nuclease-deficient Cas protein, as disclosed herein. Examples of such a desired activity include a transcription regulation activity (either activation or repression), an epigenetic modification activity, or a target visualization/identification activity, as described below. The Cas protein can be introduced into an in vitro or in vivo system as a purified or non-purified (i) Cas protein or (ii) mRNA encoded for expression of the Cas protein or (iii) linear or circular DNA encoded for expression of the protein. Any of these 3 methods of providing the Cas protein are well known in the art and are implied interchangeably when mention is made

herein of a Cas protein or use of a Cas protein. In certain embodiments, the Cas protein is constitutively expressed from mRNA or DNA. In certain embodiments, the expression of Cas protein from mRNA or DNA is inducible or

induced.

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In certain embodiments, the Cas protein is chemically synthesized (see e.g., Creighton, "Proteins: Structures and Molecular Principles," W.H. Freeman & Co., NY, 1983), or produced by recombinant DNA technology as described herein. For additional guidance, skilled artisans may consult 10 Frederick M. Ausubel et al., "Current Protocols in Molecular Biology," John Wiley & Sons, 2003; and Sambrook et al., "Molecular Cloning, A Laboratory Manual," Cold Spring Harbor Press, Cold Spring Harbor, N.Y., 2001).

In certain embodiments, the Cas protein is provided in 15 purified or isolated form. In certain embodiments, the Cas protein is provided at about 80%, about 90%, about 95%, or about 99% purity. In certain embodiments, the Cas protein is provided as part of a composition. In certain embodiments, the Cas protein is provided in aqueous compositions 20 suitable for use as, or inclusion in, a composition for an RNA-guided nuclease reaction. Those of skill in the art are well aware of the various substances that can be included in such nuclease reaction compositions.

In certain embodiments, a Cas protein is provided as a 25 recombinant polypeptide. In certain examples, the recombinant polypeptide is prepared as a fusion protein. For example, in certain embodiments, a nucleic acid encoding the Cas protein is linked to another nucleic acid encoding a fusion partner, e.g., glutathione-s-transferase (GST), 6x-His 30 epitope tag, or M13 Gene 3 protein. Suitable host cells can be used to expresses the fusion protein. In certain embodiments, the fusion protein is isolated by methods known in the art. In certain embodiments, the fusion protein can be further treated, e.g., by enzymatic digestion, to remove the 35 fusion partner and obtain the Cas protein. Alternatively, Cas protein:guide RNA complexes can be made with recombinant technology using a host cell system or an in vitro translation-transcription system known in the art. Details of such systems and technology can be found in e.g., 40 WO2014144761 WO2014144592, WO2013176772, US20140273226, and US20140273233, the contents of which are incorporated herein by reference in their entire-

#### Wild Type Cas Proteins

In certain embodiments, a Cas protein comprises a protein derived from a CRISPR-Cas type I, type II, or type III system, which has an RNA-guided polynucleotide binding and/or nuclease activity. Non-limiting examples of suitable Cas proteins include Cas3, Cas4, Cas5, Cas5e (or CasD), 50 Cash, Cas6e, Cas6f, Cas7, Cas8a1, Cas8a2, Cas8b, Cas8c, Cas9, Cas10, Cas10d, CasF, CasG, CasH, Csy1, Csy2, Csy3, Cse1 (or CasA), Cse2 (or CasB), Cse3 (or CasE), Cse4 (or CasC), Csc1, Csc2, Csa5, Csn2, Csm2, Csm3, Csm4, Csm5, Csm6, Cmr1, Cmr3, Cmr4, Cmr5, Cmr6, Csb1, Csb2, Csb3, 55 Csx17, Csx14, Csx10, Csx16, CsaX, Csx3, Csz1, Csx15, Csf1, Csf2, Csf3, Csf4, and Cu1966. See e.g., WO2014144592, WO2014144761 WO2013176772, US20140273226, and US20140273233, the contents of which are incorporated herein by reference in their entire- 60

In certain embodiments, the Cas protein is derived from a type II CRISPR-Cas system. In certain embodiments, the Cas protein is or is derived from a Cas9 protein. In certain embodiments, the Cas protein is or is derived from a 655 bacterial Cas9 protein, including those identified in WO2014144761. In certain embodiments, the Cas protein is

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or is derived from a *Streptococcus* sp. or *Staphylococcus* sp. Cas9 protein. In certain embodiments, the Cas protein is or is derived from the *Streptococcus thermophilus* Cas9 protein. In certain embodiments, the Cas protein is or is derived from a the *Streptococcus pyogenes* Cas9 protein. In certain embodiments, the Cas protein is or is derived from the *Staphylococcus aureus* Cas9 protein. In certain embodiments, the Cas protein is or is derived from the *Streptococcus thermophilus* Cas9 protein.

In certain embodiments, the wild type Cas protein is a Cas9 protein. In certain embodiments, the wild type Cas9 protein is the Cas9 protein from *S. pyogenes* (SEQ ID NO: 1). In certain embodiments, the protein or polypeptide can comprise, consist of, or consist essentially of a fragment of SEQ ID NO: 1.

In general, a Cas protein includes at least one RNA binding domain, which interacts with the guide RNA. In certain embodiments, the Cas protein is modified to increase nucleic acid binding affinity and/or specificity, alter an enzymatic activity, and/or change another property of the protein. For example, nuclease (i.e., DNase, RNase) domains of the Cas protein can be modified, mutated, deleted, or inactivated. Alternatively, the Cas protein can be truncated to remove domains that are not essential for the function of the protein. In certain embodiments, the Cas protein is truncated or modified to optimize the activity of the effector domain. In certain embodiments, the Cas protein includes a nuclear localization sequence (NLS) that effects importation of the NLS-tagged Cas protein into the nucleus of a living cell. In certain embodiments, the Cas protein includes two or more modifications.

#### Mutant Cas Proteins

In some embodiments, the Cas protein can be a mutant of a wild type Cas protein (such as Cas9) or a fragment thereof. In other embodiments, the Cas protein can be derived from a mutant Cas protein. For example, the amino acid sequence of the Cas9 protein can be modified to alter one or more properties (e.g., nuclease activity, binding affinity, stability, etc.) of the protein. Alternatively, domains of the Cas9 protein not involved in RNA-guided cleavage can be eliminated from the protein such that the modified Cas9 protein is smaller than the wild type Cas9 protein. For example, reducing the size of the Cas9 coding sequence can allow it to fit within a transfection vector that otherwise cannot accommodate the wild type sequence, such as the AAV vector among others. In some embodiments, the present system utilizes the Cas9 protein from S. pyogenes, either as encoded in bacteria or codon-optimized for expression in eukaryotic cells. Shown below is the amino acid sequence of wild type S. pyogenes Cas9 protein sequence (SEQ ID No. 1, Uniprot No. Q99ZW2.

MDKKYSIGLDIGTNSVGWAVITDEYKVPSKKFKVLGNTDRHSIKKNLIGA
LLFDSGETAEATRLKRTARRRYTRRKNRICYLQEIFSNEMAKVDDSFFHR
LEESFLVEEDKKHERHPIFGNIVDEVAYHEKYPTIYHLRKKLVDSTDKAD
LRLIYLALAHMIKFRGHFLIEGDLNPDNSDVDKLFIQLVQTYNQLFEENP
INASGVDAKAILSARLSKSRRLENLIAQLPGEKKNGLFGNLIALSLGLTP
NFKSNFDLAEDAKLQLSKDTYDDDLDNLLAQIGDQYADLFLAAKNLSDAI
LLSDILRVNTEITKAPLSASMIKRYDEHHQDLTLLKALVRQQLPEKYKEI
FFDOSKNGYAGYIDGGASOEEFYKFIKPILEKMDGTEELLVKLNREDLLR

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-continued KQRTFDNGSIPHQIHLGELHAILRRQEDFYPFLKDNREKIEKILTFRIPY YVGPLARGNSRFAWMTRKSEETITPWNFEEVVDKGASAQSFIERMTNFDK NLPNEKVLPKHSLLYEYFTVYNELTKVKYVTEGMRKPAFLSGEQKKAIVD LLFKTNRKVTVKQLKEDYFKKIECFDSVEISGVEDRFNASLGTYHDLLKI IKDKDFLDNEENEDILEDIVLTLTLFEDREMIEERLKTYAHLFDDKVMKQ LKRRRYTGWGRLSRKLINGIRDKQSGKTILDFLKSDGFANRNFMQLIHDD SLTFKEDIQKAQVSGQGDSLHEHIANLAGSPAIKKGILQTVKVVDELVKV  ${\tt MGRHKPENIVI} {\bf \underline{E}} {\tt MARENQTTQKGQKNSRERMKRIEEGIKELGSQILKEHP}$ VENTOLONEKLYLYYLONGRDMYVDOELDINRLSDYDVDHIVPOSFLKDD SIDNKVLTRSDK**N**RGKSDNVPSEEVVKKMKNYWROLLNAKLITORKFDNL TKAERGGLSELDKAGFIKROLVETROITKHVAOILDSRMNTKYDENDKLI REVKVITI,KSKI,VSDFRKDFOFYKVREINNYH**H**AH**D**AYI,NAVVGTAI,IKK YPKLESEFVYGDYKVYDVRKMIAKSEOEIGKATAKYFFYSNIMNFFKTEI TLANGEIRKRPLIETNGETGEIVWDKGRDFATVRKVLSMPOVNIVKKTEV QTGGFSKESILPKRNSDKLIARKKDWDPKKYGGFDSPTVAYSVLVVAKVE KGKSKKLKSVKELLGITIMERSSFEKNPIDFLEAKGYKEVKKDLIIKLPK  $\verb|YSLFELENGRKRMLASAGELQKGNELALPSKYVNFLYLASHYEKLKGSPE|$  ${\tt DNEQKQLFVEQHKHYLDEIIEQISEFSKRVILADANLDKVLSAYNKHRDK}$ PIREQAENIIHLFTLTNLGAPAAFKYFDTTIDRKRYTSTKEVLDATLIHQ SITGLYETRIDLSOLGGD

A Cas9 protein generally has at least two nuclease (e.g., DNase) domains. For example, a Cas9 protein can have a 35 RuvC-like nuclease domain and an HNH-like nuclease domain. The RuvC and HNH domains work together to cut both strands in a target site to make a double-stranded break in the target polynucleotide. (Jinek et al., Science, 337: 816-821). In certain embodiments, a mutant Cas9 protein is  $\,^{40}$ modified to contain only one functional nuclease domain (either a RuvC-like or an HNH-like nuclease domain). For example, in certain embodiments, the mutant Cas9 protein is modified such that one of the nuclease domains is deleted or mutated such that it is no longer functional (i.e., the nuclease activity is absent). In some embodiments where one of the nuclease domains is inactive, the mutant is able to introduce a nick into a double-stranded polynucleotide (such protein is termed a "nickase") but not able to cleave the double-50 stranded polynucleotide. For example, an aspartate to alanine (D10A) conversion in a RuvC-like domain converts the Cas9-derived protein into a nickase. Likewise, a histidine to alanine (H840A) conversion in a HNH domain converts the Cas9-derived protein into a nickase. Likewise, an arspara- 55 gine to alanine (N863A) conversion in a HNH domain converts the Cas9-derived protein into a nickase.

In certain embodiments, both the RuvC-like nuclease domain and the HNH-like nuclease domain are modified or eliminated such that the mutant Cas9 protein is unable to 60 nick or cleave the target polynucleotide. In certain embodiments, all nuclease domains of the Cas9-derived protein are modified or eliminated such that the Cas9-derived protein lacks all nuclease activity. In certain embodiments, a Cas9 protein that lacks some or all nuclease activity relative to a 65 wild-type counterpart, nevertheless, maintains target recognition activity to a greater or lesser extent.

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In any of the above-described embodiments, any or all of the nuclease domains can be inactivated by one or more deletion mutations, insertion mutations, and/or substitution mutations using well-known methods, such as site-directed mutagenesis, PCR-mediated mutagenesis, and total gene synthesis, as well as other methods known in the art.

In certain embodiments, the "Cas mutant" or "Cas variant" is at least 50% (e.g., any number between 50% and 100%, inclusive, e.g., 50%, 60%, 70%, 80%, 90%, 95%, 98%, and 99%) identical to SEQ ID NO: 1. In certain embodiments, the "Cas mutant" or "Cas variant" binds to an RNA molecule (e.g., a sgRNA). In certain embodiments, the "Cas mutant" or "Cas variant" is targeted to a specific polynucleotide sequence via the RNA molecule.

Fusion Proteins In certain embodiments, the Cas protein is fused to another protein or polypeptide heterologous to the Cas protein to create a fusion protein. In certain embodiments, the heterologous sequence includes one or more effector domains, such as a cleavage domain, a transcriptional activation domain, a transcriptional repressor domain, or an epigenetic modification domain. Additional examples of the effector domain include a nuclear localization signal, cellpenetrating or translocation domain, or a marker domain. In certain embodiments, the effector domain is located at the N-terminal, the C-terminal, or in an internal location of the fusion protein. In certain embodiments, the Cas protein of the fusion protein is or is derived from a Cas9 protein. In certain embodiments, the Cas protein of the fusion protein is or is derived from a modified or mutated Cas protein in which all the nuclease domains have been inactivated or deleted. In certain embodiments, the Cas protein of the fusion protein is or is derived from a modified or mutated Cas protein that lacks nuclease activity. In certain embodiments, the RuvC and/or HNH domains of the Cas protein are modified or mutated such that they no longer possess nuclease activity.

Cleavage Domains

In certain embodiments, the effector domain of the fusion protein is a cleavage domain. As used herein, a "cleavage domain" refers to a domain that cleaves DNA. The cleavage domain can be obtained from any endonuclease or exonuclease. Non-limiting examples of endonucleases from which a cleavage domain can be derived include restriction endonucleases and homing endonucleases. See, for example, New England Biolabs Catalog or Belfort et al. (1997) *Nucleic Acids Res.* 25, 3379-88. Additional enzymes that cleave DNA are known (e.g., 51 Nuclease; mung bean nuclease; pancreatic DNase I; micrococcal nuclease; yeast HO endonuclease). See also Linn et al. (eds.) "Nucleases," Cold Spring Harbor Laboratory Press, 1993. One or more of these enzymes (or functional fragments thereof) can be used as a source of cleavage domains.

In certain embodiments, the cleavage domain can be derived from a type II-S endonuclease. Type II-S endonucleases cleave DNA specifically at sites that are typically several base pairs away from the DNA recognition site of the endonuclease and, as such, have separable recognition and cleavage domains. These enzymes generally are monomers that transiently associate to form dimers to cleave each strand of DNA at staggered locations. Non-limiting examples of suitable type II-S endonucleases include Bfil, BpmI, BsaI, BsgI, BsmBI, BsmI, BspMI, FokI, MboII, and SapI. In certain embodiments, the cleavage domain of the fusion protein is a FokI cleavage domain or a fragment or derivative thereof. See Miller et al. (2007) *Nat. Biotechnol.* 

**55**25, 778-85; Szczpek et al. (2007) *Nat. Biotechnol.* 25, 786-93; Doyon et al. (2011) *Nat. Methods*, 8, 74-81.

Transcriptional Activation Domains

In certain embodiments, the effector domain of the fusion protein is a transcriptional activation domain. In general, a 5 transcriptional activation domain interacts with transcriptional control elements and/or transcriptional regulatory proteins (i.e., transcription factors, RNA polymerases, etc.) to increase and/or activate transcription of a gene. In certain embodiments, the transcriptional activation domain is a 10 herpes simplex virus VP16 activation domain, VP64 (which is a tetrameric derivative of VP16), a NFkB p65 activation domain, p53 activation domains 1 and 2, a CREB (cAMP response element binding protein) activation domain, an E2A activation domain, or an NFAT (nuclear factor of 15 activated T-cells) activation domain. In certain embodiments, the transcriptional activation domain is Gal4, Gcn4, MLL, Rtg3, Gln3, Oaf1, Pip2, Pdr1, Pdr3, Pho4, or Leu3. The transcriptional activation domain may be wild type, or it may be a modified or truncated version of the original 20 transcriptional activation domain.

Transcriptional Repressor Domains

In certain embodiments, the effector domain of the fusion protein is a transcriptional repressor domain. In general, a transcriptional repressor domain interacts with transcriptional control elements and/or transcriptional regulatory proteins (i.e., transcription factors, RNA polymerases, etc.) to decrease and/or prohibit transcription of a gene. In certain embodiments, the transcriptional repressor domains is inducible cAMP early repressor (ICER) domains, Kruppelassociated box A (KRAB-A) repressor domains, YY1 glycine rich repressor domains, Sp1-like repressors, E(spI) repressors, IkB repressor, or MeCP2.

Epigenetic Modification Domains

In certain embodiments, the effector domain of the fusion 35 protein is an epigenetic modification domain. In general, epigenetic modification domains alter gene expression by modifying the histone structure and/or chromosomal structure. In certain embodiments, the epigenetic modification domains is a histone acetyltransferase domain, a histone 40 deacetylase domain, a histone methyltransferase domain, a histone demethylase domain, a DNA methyltransferase domain, or a DNA demethylase domain.

Additional Domains

In certain embodiments, the fusion protein further comprises at least one additional domain. Non-limiting examples of suitable additional domains include nuclear localization signals (NLSs), cell-penetrating or translocation domains, and marker domains. An NLS generally comprises a stretch of basic amino acids. See, e.g., Lange et al. (2007) *J. Biol.* 50 *Chem.*, 282, 5101-5. For example, in certain embodiments, the NLS is a monopartite sequence, such as PKKKRKV (SEQ ID NO: 2) or PKKKRRV (SEQ ID NO: 3). In certain embodiments, the NLS is a bipartite sequence. In certain embodiments, the NLS is KRPAATKKAGQAKKKK (SEQ 55 ID NO: 4).

In certain embodiments, the fusion protein comprises at least one cell-penetrating domain. In certain embodiments, the cell-penetrating domain is a cell-penetrating peptide sequence derived from the HIV-1 TAT protein. As an 60 example, the TAT cell-penetrating sequence can be GRKKRRQRRRPPQPKKKRKV (SEQ ID NO: 5). In certain embodiments, the cell-penetrating domain is TLM (PLSSIFSRIGDPPKKKRKV; SEQ ID NO: 6), a cell-penetrating peptide sequence derived from the human hepatitis 65 B virus. In certain embodiments, the cell-penetrating domain is MPG (GALFLGWLGAAGSTMGAPKKKRKV; SEQ ID

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NO: 7 or GALFLGFLGAAGSTMGAWSQPKKKRKV; SEQ ID NO: 8). In certain embodiments, the cell-penetrating domain is Pep-1 (KETWWETWWTEWSQPKKKRKV; SEQ ID NO: 9), VP22, a cell penetrating peptide from Herpes simplex virus, or a polyarginine peptide sequence.

In certain embodiments, the fusion protein comprises at least one marker domain. Non-limiting examples of marker domains include fluorescent proteins, purification tags, and epitope tags. In certain embodiments, the marker domain is a fluorescent protein. Non limiting examples of suitable fluorescent proteins include green fluorescent proteins (e.g., GFP, GFP-2, tagGFP, turboGFP, EGFP, Emerald, Azami Green, Monomeric Azami Green, CopGFP, AceGFP, ZsGreen1), yellow fluorescent proteins (e.g. YFP, EYFP, Citrine, Venus, YPet, PhiYFP, ZsYellow1), blue fluorescent proteins (e.g. EBFP, EBFP2, Azurite, mKalama1, GFPuv, Sapphire, T-sapphire), cyan fluorescent proteins (e.g. ECFP, Cerulean, CyPet, AmCyan1, Midoriishi-Cyan), red fluorescent proteins (mKate, mKate2, mPlum, DsRed monomer, mCherry, mRFP1, DsRed-Express, DsRed2, DsRed-Monomer, HcRed-Tandem, HcRedl, AsRed2, eqFP611, mRasberry, mStrawberry, Jred), orange fluorescent proteins (mOrange, mKO, Kusabira-Orange, Monomeric Kusabira-Orange, mTangerine, tdTomato) and any other suitable fluorescent protein. In certain embodiments, the marker domain is a purification tag and/or an epitope tag. Exemplary tags include, but are not limited to, glutathione-Stransferase (GST), chitin binding protein (CBP), maltose binding protein, thioredoxin (TRX), poly(NANP), tandem affinity purification (TAP) tag, myc, AcV5, AU1, AU5, E, ECS, E2, FLAG, HA, nus, Softag 1, Softag 3, Strep, SBP, Glu-Glu, HSV, KT3, S, S1, T7, V5, VSV-G, 6×His, biotin carboxyl carrier protein (BCCP), and calmodulin.

#### V. Uses and Methods

In one aspect, the present invention provides a method for cleaving a target polynucleotide with a Cas protein. The method comprises contacting the target polynucleotide with (i) a guide RNA or a set of guide RNA molecules described herein, and (ii) a Cas protein. In certain embodiments, the method results in a double-strand break in the target polynucleotide. In certain embodiments, the Cas protein is a Cas protein having a single-strand nicking activity. In certain embodiments, the method results in a single-strand break in the target polynucleotide. In certain embodiments, a complex comprising a guide RNA and Cas protein having a single-strand nicking activity is used for sequence-targeted single-stranded DNA cleavage, i.e., nicking.

In one aspect, the present invention provides a method for cleaving two or more target polynucleotides with a Cas protein. The method comprises contacting the target polynucleotides with (i) a set of guide RNA molecules described herein, and (ii) a Cas protein. In certain embodiments, the method results in double-strand breaks in the target polynucleotides. In certain embodiments, the Cas protein is a Cas protein having a single-strand nicking activity. In certain embodiments, the method results in single-strand breaks in the target polynucleotides. In certain embodiments, a complex comprising a guide RNA and Cas protein having a single-strand nicking activity is used for sequence-targeted single-stranded DNA cleavage, i.e., nicking.

In one aspect, the present invention provides a method for binding a target polynucleotide with a Cas protein. The method comprises contacting the target polynucleotide with (i) a guide RNA or a set of guide RNA molecules described herein and (ii) a Cas protein, to result in binding of the target

polynucleotide with the Cas protein. In certain embodiments, the Cas protein is a Cas variant. In certain embodiments, the Cas variant lacks some or all nuclease activity relative to a counterpart wild-type Cas protein.

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In one aspect, the present invention provides a method for 5 binding two or more target polynucleotides with a Cas protein. The method comprises contacting the target polynucleotides with (i) a set of RNA molecules described herein and (ii) a Cas protein, to result in binding of the target polynucleotides with the Cas protein. In certain embodiments, the Cas protein is a Cas variant. In certain embodiments, the Cas variant lacks some or all nuclease activity relative to a counterpart wild-type Cas protein.

In one aspect, the present invention provides a method for targeting a Cas protein to a target polynucleotide. The 15 method comprises contacting the Cas protein with a guide RNA or a set of guide RNA molecules described herein. In certain embodiments, the method results in formation of a guide RNA:Cas protein complex. In certain embodiments, the Cas protein is a wild type Cas9 protein. In certain 20 embodiments, the Cas protein is a mutant or variant of a Cas9 protein. In certain embodiments, the Cas protein is a Cas protein is a Cas protein having a single-strand nicking activity. In certain embodiments, the Cas protein is a Cas protein lacking nuclease activity (e.g., a nuclease-deficient mutant of Cas 25 protein). In certain embodiments, the Cas protein is part of a fusion protein (e.g., a fusion protein comprising (i) the Cas protein and (ii) a heterologous polypeptide).

In one aspect, the present invention provides a method for targeting a Cas protein to two or more target polynucle-30 otides. The method comprises contacting the Cas protein with a set of guide RNA molecules described herein. In certain embodiments, the method results in formation of a guide RNA:Cas protein complex. In certain embodiments, the Cas protein is a wild type Cas9 protein. In certain 35 embodiments, the Cas protein is a mutant or variant of a Cas9 protein. In certain embodiments, the Cas protein is a Cas protein is a Cas protein having a single-strand nicking activity. In certain embodiments, the Cas protein is a Cas protein lacking nuclease activity (e.g., a nuclease-deficient mutant of Cas 40 protein). In certain embodiments, the Cas protein is part of a fusion protein (e.g., a fusion protein comprising (i) the Cas protein or and (ii) a heterologous polypeptide).

In certain embodiments, the guide RNA is introduced into a cell by transfection. Techniques for RNA transfection are 45 known in the art and include electroporation and lipofection. Effective techniques for RNA transfection depend mostly on cell type. See, e.g., Lujambio et al. (Spanish National Cancer Centre) Cancer Res. February 2007, which describes transfection of HTC-116 colon cancer cells and uses Oligo- 50 fectamine (Invitrogen) for transfection of commercially obtained, modified miRNA or precursor miRNA. See also, Cho et al. (Seoul National Univ.) Nat. Biotechnol. March 2013, which describes transfection of K562 cells and uses 4D Nucleofection<sup>TM</sup> (Lonza) electroporation for transfection 55 of transcribed sgRNAs (about 60 nts long). Techniques for transfection of RNA are also known in the art. For example, therapeutic RNA has been delivered in non-pathogenic E. coli coated with Invasin protein (to facilitate uptake into cells expressing β-1 integrin protein) and with the E. coli 60 encoded to express lysteriolysin O pore-forming protein to permit the shRNA to pass from the E. coli into the cytoplasm. See also Cho et al. (Seoul National Univ.) Nat. Biotechnol. March 2013.

In certain embodiments, the guide RNA is introduced or 65 delivered into cells. Technologies that can be used for delivery of guide RNA include those that utilize encapsu-

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lation by biodegradable polymers, liposomes, or nanoparticles. Such polymers, liposomes, and nanoparticles can be delivered intravenously. In certain embodiments, for in vivo delivery, guide RNA can be injected into a tissue site or administered systemically. In vivo delivery can also be effected by a beta-glucan delivery system, such as those described in U.S. Pat. Nos. 5,032,401 and 5,607,677, and U.S. Publication No. 2005/0281781, which are hereby incorporated by reference in their entirety. In certain embodiments, guide RNA or a delivery vehicle containing guide RNA is targeted to a particular tissue or body compartment. For example, in certain embodiments, to target exogenous RNA to other tissues, synthetic carriers are decorated with cell-specific ligands or aptamers for receptor uptake, e.g., RNA encased in cyclodextrin nanoparticles coated with PEG and functionalized with human transferrin protein for uptake via the transferrin receptor which is highly expressed in tumor cells. Further approaches are described herein below or known in the art.

The present invention has been tested in human cells as described in Hendel et al., *Nat. Biotechnol.* (2015) 33:9, 985-9 (which is incorporated in this application in its entirety). In the cited work, modified guide RNA was introduced into K562 cells, human primary T cells, and CD34+ hematopoietic stem and progenitor cells (HSPCs). The modified guide RNA significantly enhanced genome editing efficiencies in human cells, including human primary T cells and CD34+ HSPCs as compared to unmodified guide RNA.

FIGS. 11A and 11B illustrate experimental results showing that gene disruption in human cell lines can be achieved by high frequencies of indels or by cleavage-stimulated homologous recombination using synthesized and chemically modified sgRNAs disclosed herein. Gene disruption by mutagenic NHEJ was measured by deep sequencing of PCR amplicons (FIG. 12A) or gene addition by HR at the three loci IL2RG, HBB and CCR5 in K562 cells induced by Cas9 in combination with synthetic sgRNAs (FIG. 12B). The synthetic sgRNAs were delivered at 1 µg (light shade) or 20 μg (dark shade) per 1 million cells. Cas9 was expressed from a plasmid (2 μg) and for HR experiments 5 μg of GFPencoding donor plasmid was included. As a positive control, 2 μg of sgRNA plasmid encoding both the sgRNA and the Cas9 protein was used (gray bars). Bars represent average values+s.e.m., n=3.

FIGS. 12A, 12B, 12C and 12D illustrate experimental results showing that chemically modified sgRNAs as described herein can be used to achieve high frequencies of gene disruption or targeted genome editing in stimulated primary human T cells and CD34+ hematopoietic stem and progenitor cells (HSPCs).

FIG. 12A illustrates results from primary human T cells nucleofected with 10 µg of a synthetic CCR5 sgRNAs and either 15 µg Cas9 mRNA or 1 µg Cas9-encoding plasmid. 1 µg sgRNA plasmid encoding both the sgRNA and Cas9 protein was included for comparison. The bars represent average indel frequencies for three different donors+s.e.m., n=6, as measured by TIDE (tracking of indels by decomposition) analysis of PCR amplicons spanning the sgRNA target site, and using a mock-treated sample as control reference. Delivery of Cas9 mRNA with the unmodified or the M-modified sgRNA, and nucleofection of the plasmid encoding both the sgRNA and Cas9, did not give rise to allele modification frequencies above background. Co-transfection of the MSP-modified sgRNA with DNA expression plasmid for Cas9 generated 9.3% indel frequency. Cas9

59 mRNA with either the MS- or MSP-modified sgRNA generated 48.7% and 47.9% indel frequencies, respectively.

FIG. 12B illustrates results from stimulated T cells. The cells were nucleofected as above, but with 15 µg Cas9 protein complexed with a 2.5 molar excess of the indicated 5 synthetic CCR5 sgRNAs. Indel frequencies were measured by TIDE analysis. The bars represent average indel frequencies for three different donors+s.e.m., n=6. A 2.4-fold improvement in indel frequencies of the MS-modified sgRNA over the unmodified sgRNA (30.7% vs. 12.8%) was observed for chemically modified sgRNAs when delivered complexed with Cas9 protein. These results establish that chemically modified sgRNAs can be used for genome editing of stimulated T cells when delivered complexed with 15 Cas9 protein.

FIG. 12C illustrates results from human peripheral blood CD34+HSPCs. 500,000 mobilized cells were nucleofected with 10 μg of the indicated synthetic sgRNAs targeting IL2RG or HBB and either 15 μg Cas9 mRNA or 1 μg Cas9 20 plasmid. 1 µg of sgRNA plasmid encoding both the sgRNA and Cas9 protein was included for comparison. Bars represent average indel frequencies+s.e.m., n=3, as measured by T7 endonuclease cleavage assay. Indels were not detected at either locus using the unmodified or M-modified sgRNAs 25 when co-transfected with Cas9 mRNA. However, the IL2RG MS- and MSP-modified sgRNAs showed 17.5% and 17.7% indel frequencies, respectively, and 23.4% and 22.0%, respectively, for the HBB MS- and MSP-modified sgRNAs.

FIG. 12D illustrates results from stimulated T cells or mobilized human peripheral blood CD34+ HSPCs. One million cells were nucleofected with 15 µg Cas9 mRNA and 10  $\mu g$  of the indicated synthetic CCR5 sgRNAs. A recent  $_{35}$ study showed that the simultaneous use of two sgRNAs could improve gene disruption in human primary T cells and in CD34+ HSPCs. See, e.g., Mandal et al. (2014) Cell Stem Cell, 15, 643-52. MS- and MSP-modified CCR5 sgRNAs were chemically synthesized with the sequences reported in 40 Mandal study (termed 'D' and 'Q'), which cut 205 base pairs apart. When used in combination, the amount of each sgRNA was 5 µg. Indel frequencies for samples with single sgRNAs were measured by TIDE analysis as above and allele disruption frequencies for samples with two sgRNAs 45 were measured by sequencing of cloned PCR products. The bars represent average indel frequencies+s.e.m., n=3. In T cells, the 'D' sgRNA alone gave rise to 56.0% and 56.3% indels for the MS- and MSP-modified sgRNA, respectively, and the 'Q' sgRNA gave rise to 62.6% and 69.6% indels, 50 respectively. When used in combination, the frequencies of allele modification increased, as we observed 73.9% and 93.1% indels for the MS- and MSP-modified sgRNAs, respectively, of which the majority of the modification events were deletions between the two sgRNA target sites. 55 In CD34+HSPCs, observations were similar though the overall frequencies were lower. For the 'D' sgRNA, allele modification frequencies of 9.8% and 11.2% were observed for the MS- and MSP-modified sgRNA, respectively, and 17.8% and 19.2% for the 'Q' sgRNA. When used in com-  $_{60}$ bination the frequencies increased to 37.8% and 43.0% for the MS- and MSP-modified sgRNAs, respectively. This shows that the use of two chemically modified sgRNAs is a highly effective way to facilitate gene disruption in primary human T cells and CD34+ HSPCs.

Examples of other uses include genomic editing and gene expression regulation as described below.

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Genomic Editing

In one aspect, the present invention provides a method for genomic editing to modify a DNA sequence in vivo or in vitro ("in vitro" includes, without being limited to, a cellfree system, a cell lysate, an isolated component of a cell, and a cell outside of a living organism). The DNA sequence may comprise a chromosomal sequence, an episomal sequence, a plasmid, a mitochondrial DNA sequence, or a functional intergenic sequence, such as an enhancer sequence or a DNA sequence for a non-coding RNA. The method comprises contacting the DNA sequence with (i) a guide RNA or a set of guide RNA molecules described herein, and (ii) a Cas protein. In certain embodiments, the DNA sequence is contacted outside of a cell. In certain embodiments, the DNA sequence is located in the genome within a cell and is contacted in vitro or in vivo. In certain embodiments, the cell is within an organism or tissue. In certain embodiments, the cell is a human cell, a non-human mammalian cell, a stem cell, a non-mammalian vertebrate cell, an invertebrate cell, a plant cell, a single cell organism, or an embryo. In certain embodiments, the guide RNA aids in targeting the Cas protein to a targeted site in the DNA sequence. In certain embodiments, the Cas protein cleaves at least one strand of the DNA sequence at the targeted site. In certain embodiments, the Cas protein cleaves both strands of the DNA sequence at the targeted site.

In certain embodiments, the method further comprises introducing the Cas protein into a cell or another system. In certain embodiments, the Cas protein is introduced as a purified or non-purified protein. In certain embodiments, the Cas protein is introduced via an mRNA encoding the Cas protein. In certain embodiments, the Cas protein is introduced via a linear or circular DNA encoding the Cas protein. In certain embodiments, the cell or system comprises a Cas protein or a nucleic acid encoding a Cas protein.

In certain embodiments, a double-stranded break can be repaired via an error-prone, non-homologous end-joining ("NHEJ") repair process. In certain embodiments, a doublestranded break can be repaired by a homology-directed repair (HDR) process such that a donor sequence in a donor polynucleotide can be integrated into or exchanged with the targeted DNA sequence.

In certain embodiments, the method further comprises introducing at least one donor polynucleotide into the cell or system. In certain embodiments, the donor polynucleotide comprises at least one homologous sequence having substantial sequence identity with a sequence on either side of the targeted site in the DNA sequence. In certain embodiments, the donor polynucleotide comprises a donor sequence that can be integrated into or exchanged with the DNA sequence via homology-directed repair, such as homologous recombination.

In certain embodiments, the donor polynucleotide includes an upstream homologous sequence and a downstream homologous sequence, each of which have substantial sequence identity to sequences located upstream and downstream, respectively, of the targeted site in the DNA sequence. These sequence similarities permit, for example, homologous recombination between the donor polynucleotide and the targeted DNA sequence such that the donor sequence can be integrated into (or exchanged with) the DNA sequence targeted.

In certain embodiments, the target site(s) in the DNA sequence spans or is adjacent to a mutation, e.g., point mutation, a translocation or an inversion which may cause or be associated with a disorder. In certain embodiments, the method comprises correcting the mutation by introducing into the cell or system at least one donor polynucleotide

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comprising (i) a wild type counterpart of the mutation and (ii) at least one homologous sequence having substantial sequence identity with a sequence on one side of the targeted site in the DNA sequence. In certain embodiments, the donor polynucleotide comprises a homologous sequence having 5 substantial sequence identity with a sequence on both sides of the targeted site in the DNA sequence.

In certain embodiments, the donor polynucleotide comprises an exogenous sequence that can be integrated into or exchanged with the targeted DNA sequence via a homologydirected repair process, such as homologous recombination. In certain embodiments, the exogenous sequence comprises a protein coding gene, which, optionally, is operably linked to an exogenous promoter control sequence. Thus, in certain embodiments, upon integration of the exogenous sequence, 15 a cell can express a protein encoded by the integrated gene. In certain embodiments, the exogenous sequence is integrated into the targeted DNA sequence such that its expression in the recipient cell or system is regulated by the exogenous promoter control sequence. Integration of an 20 exogenous gene into the targeted DNA sequence is termed a "knock in." In other embodiments, the exogenous sequence can be a transcriptional control sequence, another expression control sequence, an RNA coding sequence, and

In certain embodiments, the donor polynucleotide comprises a sequence that is essentially identical to a portion of the DNA sequence at or near the targeted site, but comprises at least one nucleotide change. For example, in certain embodiments, the donor sequence comprises a modified or 30 mutated version of the DNA sequence at or near the targeted site such that, upon integration or exchange with the targeted site, the resulting sequence at the targeted site comprises at least one nucleotide change. In certain embodiments, the at least one nucleotide change is an insertion of one or more 35 nucleotides, a deletion of one or more nucleotides, a substitution of one or more nucleotides, or combinations thereof. As a consequence of the integration of the modified sequence, the cell may produce a modified gene product from the targeted DNA sequence.

In certain embodiments, the methods are for multiplex applications. In certain embodiments, the methods comprise introducing a library of guide RNAs into the cell or system. In certain embodiments, the library comprises at least 100 unique guide sequences. In certain embodiments, the library 45 comprises at least 1,000 unique guide sequences. In certain embodiments, the library comprises at least 10,000 unique guide sequences. In certain embodiments, the library comprises at least 100,000 unique guide sequences. In certain embodiments, the library comprises at least 1,000,000 50 unique guide sequences. In certain embodiments, the library targets at least 10 different polynucleotides or at least 10 different sequences within one or more polynucleotides. In certain embodiments, the library targets at least 100 different polynucleotides or at least 100 different sequences within 55 one or more polynucleotides. In certain embodiments, the library targets at least 1,000 different polynucleotides or at least 1,000 different sequences within one or more polynucleotides. In certain embodiments, the library targets at least 10,000 different polynucleotides or at least 10,000 60 different sequences within one or more polynucleotides. In certain embodiments, the library targets at least 100,000 different polynucleotides or at least 100,000 different sequences within one or more polynucleotides. In certain embodiments, the library targets at least 1,000,000 different 65 polynucleotides or at least 1,000,000 different sequences within one or more polynucleotides.

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Genomic Editing in Human and Mammalian Cells

Embodiments of the present invention are useful in methods for genomic editing to modify a target polynucleotide, for example a DNA sequence, in a mammalian cell.

In certain embodiments, the DNA sequence is a chromosomal sequence. In certain embodiments, the DNA sequence is a protein-coding sequence. In certain embodiments, the DNA sequence is a functional intergenic sequence, such as an enhancer sequence or a non-coding sequence. In certain embodiments, the DNA is part of a human gene. In some such embodiments, the human gene is the clathrin light chain (CLTA1) gene, the human interleukin 2 receptor gamma (IL2RG) gene, the human cytotoxic T-lymphocyteassociated protein 4 (CLTA4) gene, the human protocadherin alpha 4 (PCDHA4) gene, the human engrailed homeobox 1 (EN1) gene), the human hemoglobin beta (HBB) gene, which can harbor mutations responsible for sickle cell anemia and thalassemias, or the human chemokine (C-C motif) receptor 5 (CCR5) gene which encodes a co-receptor of HIV.

In certain embodiments, the mammalian cell is a human cell. In some such embodiments, the human cell is a primary human cell. In further embodiments, the primary human cell is a human primary T cell. The human primary T cell may be stimulated or unstimulated. In certain embodiments, the human cell is a stem/progenitor cell, such as a CD34+hematopoietic stem and progenitor cell (HSPC). In certain embodiments, the human cell is from a cultured cell line, for example such as can be obtained commercially. Exemplary cell lines include K562 cells, a human myelogenous leukemia line.

In certain embodiments, the cell is within a living organism. In certain other embodiments, the cell is outside of a living organism.

The method comprises contacting the DNA sequence with (i) a guide RNA or a set of guide RNA molecules described herein, and (ii) a Cas protein.

In certain embodiments, the method further comprises introducing or delivering the guide RNA into the cell. In some such embodiments, the guide RNA is introduced into a cell by transfection. Techniques for RNA transfection are known in the art and include electroporation and lipofection. In other embodiments, the guide RNA is introduced into a cell (and, more particularly, a cell nucleus) by nucleofection. Techniques for nucleofection are known in the art and may utilize nucleofection devices such as the Lonza Nucleofector 2b or the Lonza 4D-Nucleofector and associated reagents.

In certain embodiments, the method further comprises introducing or delivering the Cas protein into the cell. In some such embodiments, the Cas protein is introduced as a purified or non-purified protein. In other embodiments, the Cas protein is introduced via an mRNA encoding the Cas protein. In some such embodiments, the mRNA encoding the Cas protein is introduced into the cell by transfection. In other embodiments, the mRNA encoding the Cas protein is introduced into a cell (and, more particularly, a cell nucleus) by nucleofection.

In certain embodiments, the method employs ribonucleoprotein (RNP)-based delivery such that the Cas protein is introduced into the cell in a complex with the guide RNA. For example, a Cas9 protein may be complexed with a guide RNA in a Cas9:gRNA complex, which allows for codelivery of the gRNA and Cas protein. For example, the Cas:gRNA complex may be nucleofected into cells.

In certain embodiments, the method employs an all-RNA delivery platform. For example, in some such embodiments, the guide RNA and the mRNA encoding the Cas protein are

introduced into the cell simultaneously or substantially simultaneously (e.g., by co-transfection or co-nucleofection). In certain embodiments, co-delivery of Cas mRNA and modified gRNA results in higher editing frequencies as compared to co-delivery of Cas mRNA and unmodified 5 gRNA. In particular, gRNA having 2'-O-methyl-3'-phosphorothioate (MS), or 2'-O-methyl-3'-thioPACE (MSP) incorrothioate

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rothioate (MS), or 2'-O-methyl-3'-thioPACE (MSP) incorporated at three terminal nucleotides at both the 5' and 3' ends, provide higher editing frequencies as compared to unmodified gRNA.

In certain embodiments, the guide RNA and the mRNA encoding the Cas protein are introduced into the cell sequentially; that is, the guide RNA and the mRNA encoding the Cas protein are introduced into the cell at different times. The time period between the introduction of each agent may range from a few minutes (or less) to several hours or days. For example, in some such embodiments, gRNA is delivered first, followed by delivery of Cas mRNA 4, 8, 12 or 24 hours later. In other such embodiments, Cas mRNA is delivered first, followed by delivery of gRNA 4, 8, 12 or 24 hours later. In some particular embodiments, delivery of modified gRNA first, followed by delivery of Cas mRNA results in higher editing frequencies as compared to delivery of unmodified gRNA followed by delivery of Cas mRNA.

In certain embodiments, the gRNA is introduced into the 25 cell together with a DNA plasmid encoding the Cas protein. In some such embodiments, the gRNA and the DNA plasmid encoding the Cas protein are introduced into the cell by nucleofection. In some particular embodiments, an RNP-based delivery platform or an all-RNA delivery platform 30 provides lower cytotoxicity in primary cells than a DNA plasmid-based delivery system.

In certain embodiments, the method provides significantly enhanced genome editing efficiencies in human cells, including human primary T cells and CD34+HSPCs.

In certain embodiments, modified gRNA increases the frequency of insertions or deletions (indels), which may be indicative of mutagenic NHEJ and gene disruption, relative to unmodified gRNA. In particular, modified gRNA having 2'-O-methyl-3'-phosphorothioate (MS) or 2'-O-methyl-3'- 40 thioPACE (MSP) incorporated at three terminal nucleotides at both the 5' and 3' ends, increases the frequency of indels relative to unmodified gRNA.

In certain embodiments, co-delivery of modified gRNA and Cas mRNA to human primary T cells increases the 45 frequency of indels as compared to co-delivery of unmodified gRNA and Cas mRNA. In particular, modified gRNA having 2'-O-methyl-3'-phosphorothioate (MS) or 2'-O-methyl-3'-thioPACE (MSP) incorporated at three terminal nucleotides at both the 5' and 3' ends, increases the frequency of indels in human primary T cells relative to unmodified gRNA.

In certain embodiments, modified gRNA improves gRNA stability relative to unmodified gRNA. As one example, gRNA having 2'-O-methyl (M) incorporated at three terminal nucleotides at both the 5' and 3' ends, modestly improves stability against nucleases and also improves base pairing thermostability over unmodified gRNA. As another example, gRNA having 2'-O-methyl-3'-phosphorothioate (MS) or 2'-O-methyl-3'-thioPACE (MSP) incorporated at 60 three terminal nucleotides at both the 5' and 3' ends, dramatically improves stability against nucleases relative to unmodified gRNA. It is contemplated that gRNA end modifications enhance intracellular stability against exonucleases, thus enabling increased efficacy of genome editing 65 when Cas mRNA and gRNA are co-delivered or sequentially delivered into human cells.

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In certain embodiments, modified gRNA stimulates gene targeting, which, in turn, allows for gene editing by, for example, homologous recombination or NHEJ. In particular, gRNA having 2'-O-methyl-3'-phosphorothioate (MS), or 2'-O-methyl-3'-thioPACE (MSP) incorporated at three terminal nucleotides at both the 5' and 3' ends, stimulates higher levels of homologous recombination than unmodified gRNA.

In certain embodiments, modified gRNA retains high specificity. In certain embodiments, the ratio of on-target to off-target indel frequencies is improved with modified gRNA as compared to unmodified gRNA. In certain embodiments, modified gRNA delivered in an RNP complex with a Cas protein provides significantly better on-target: off-target ratios compared to a DNA plasmid-based delivery system.

Gene Expression Regulation

later. In other such embodiments, Cas mRNA is delivered first, followed by delivery of gRNA 4, 8, 12 or 24 hours later.

In some particular embodiments, delivery of modified gRNA first, followed by delivery of Cas mRNA results in higher editing frequencies as compared to delivery of unmodified gRNA followed by delivery of Cas mRNA.

In certain embodiments, the guide RNA described herein is used for regulating transcription or expression of a gene of interest. For example, in certain embodiments, a fusion protein comprising a Cas protein (e.g., a nuclease-deficient Cas9) and a transcription of a gene. Similarly, in certain embodiments, a fusion protein comprising a Cas protein (e.g., a nuclease-deficient Cas9) and a repressor polypeptide is used to knock-down gene expression by interfering with transcription of the gene.

In at least one aspect, the present invention provides a method for regulating the expression of a gene of interest in vivo or in vitro. The method comprises introducing into a cell or another system (i) a synthetic guide RNA described herein, and (ii) a fusion protein. In certain embodiments, the fusion protein comprises a Cas protein and an effector domain, such as a transcriptional activation domain, a transcriptional repressor domain, or an epigenetic modification domain. In certain embodiments, the fusion protein comprises a mutated Cas protein, such as a Cas9 protein that is a null nuclease. In certain embodiments, the Cas protein contains one or more mutations, such as D10A, H840A and/or N863A.

In certain embodiments, the fusion protein is introduced into the cell or system as a purified or non-purified protein. In certain embodiments, the fusion protein is introduced into the cell or system via an mRNA encoding the fusion protein. In certain embodiments, the fusion protein is introduced into the cell or system via a linear or circular DNA encoding the fusion protein.

In certain embodiments, the guide RNA aids in directing the fusion protein to a specific target polynucleotide comprising a chromosomal sequence, an episomal sequence, a plasmid, a mitochondrial DNA sequence, or a functional intergenic sequence, such as an enhancer or the DNA sequence for a non-coding RNA. In certain embodiments, the effector domain regulates expression of a sequence in the target polynucleotide. A guide RNA for modulating gene expression can be designed to target any desired endogenous gene or sequence encoding a functional RNA. A genomic target sequence can be selected in proximity of the transcription start site of the endogenous gene, or alternatively, in proximity of the translation initiation site of the endogenous gene. In certain embodiments, the target sequence is in a region of the DNA that is traditionally termed the "promoter proximal" region of a gene. In certain embodiments, the target sequence lies in a region from about 1,000 base pairs upstream of the transcription start site to about 1,000 base pairs downstream of the transcription start site. In

certain embodiments, the target sequence is remote from the start site for transcription of the gene (e.g., on another chromosome)

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In certain embodiments, the methods are for multiplex applications. In certain embodiments, the methods comprise 5 introducing a library of guide RNAs into the cell or system. In certain embodiments, the library comprises at least 100, at least 1,000, at least 10,000, at least 100,000, or at least 1,000,000 unique guide sequences. In certain embodiments, the library targets at least 10 different polynucleotides or at 10 least 10 different sequences within one or more polynucleotides. In certain embodiments, the library targets at least 100 different polynucleotides or at least 100 different sequences within one or more polynucleotides. In certain embodiments, the library targets at least 1,000 different 15 polynucleotides or at least 1,000 different sequences within one or more polynucleotides. In certain embodiments, the library targets at least 10,000 different polynucleotides or at least 10,000 different sequences within one or more polynucleotides. In certain embodiments, the library targets at 20 least 100,000 different polynucleotides or at least 100,000 different sequences within one or more polynucleotides. In certain embodiments, the library targets at least 1,000,000 different polynucleotides or at least 1,000,000 different sequences within one or more polynucleotides. Kits

In one aspect, the present invention provides kits containing reagents for performing the above-described methods, including producing gRNA:Cas protein complex and/or supporting its activity for binding, nicking or cleaving target 30 polynucleotide. In certain embodiments, one or more of the reaction components, e.g., one or more guide RNAs and Cas proteins, for the methods disclosed herein, can be supplied in the form of a kit for use. In certain embodiments, the kit comprises a Cas protein or a nucleic acid encoding the Cas 35 protein, and one or more guide RNAs described herein or a set or library of guide RNAs. In certain embodiments, the kit includes one or more other reaction components. In certain embodiments, an appropriate amount of one or more reaction components is provided in one or more containers or 40 held on a substrate.

Examples of additional components of the kits include, but are not limited to, one or more different polymerases, one or more host cells, one or more reagents for introducing foreign nucleic acid into host cells, one or more reagents 45 (e.g., probes or PCR primers) for detecting expression of the guide RNA and/or the Cas mRNA or protein or for verifying the status of the target nucleic acid, and buffers, transfection reagents or culture media for the reactions (in 1× or more concentrated forms). In certain embodiments, the kit 50 includes one or more of the following components: biochemical and physical supports; terminating, modifying and/or digesting reagents; osmolytes; and apparati for reaction, transfection and/or detection.

The reaction components used can be provided in a 55 variety of forms. For example, the components (e.g., enzymes, RNAs, probes and/or primers) can be suspended in an aqueous solution or bound to a bead or as a freeze-dried or lyophilized powder or pellet. In the latter case, the components, when reconstituted, form a complete mixture 60 of components for use in an assay. The kits of the invention can be provided at any suitable temperature. For example, for storage of kits containing protein components or complexes thereof in a liquid, it is preferred that they are provided and maintained below 0° C., preferably at about 65 –20° C., possibly in a freeze-resistant solution containing glycerol or other suitable antifreeze.

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A kit or system may contain, in an amount sufficient for at least one assay, any combination of the components described herein. In some applications, one or more reaction components may be provided in pre-measured single use amounts in individual, typically disposable, tubes or equivalent containers. With such an arrangement, a RNA-guided nuclease reaction can be performed by adding a target nucleic acid, or a sample or cell containing the target nucleic acid, to the individual tubes directly. The amount of a component supplied in the kit can be any appropriate amount and may depend on the market to which the product is directed. The container(s) in which the components are supplied can be any conventional container that is capable of holding the supplied form, for instance, microfuge tubes, microtiter plates, ampoules, bottles, or integral testing devices, such as fluidic devices, cartridges, lateral flow, or other similar devices.

The kits can also include packaging materials for holding the container or combination of containers. Typical packaging materials for such kits and systems include solid matrices (e.g., glass, plastic, paper, foil, micro-particles and the like) that hold the reaction components or detection probes in any of a variety of configurations (e.g., in a vial, microtiter plate well, microarray, and the like). The kits may further include instructions recorded in a tangible form for use of the components.

#### **EXAMPLES**

#### Example 1

To evaluate the ability of the chemically synthesized guide RNAs to target and cleave a DNA target sequence, an in vitro cleavage assay was developed. Briefly, as shown in FIG. 3, ~4-kb PAM-addressable DNA targets were prepared by preparative PCR amplification of plasmid-borne human sequences (here, a sequence from the human clathrin light chain CLTA gene). In a 20-uL reaction volume, 50 fmoles of linearized DNA target in the presence of 50 nM sgRNA, 39 nM recombinant purified Cas9 protein (S. pyogenes; Agilent) and 10 mM MgCl<sub>2</sub> at pH 7.6 was incubated at 37° C. for 30 min. Upon completion, 0.5 uL of RNace It (Agilent) was added, and incubation was continued at 37° C. for 5 min and then at 70° C. for 15 min. Subsequently 0.5 μL of Proteinase K (Mol. Bio. grade, NEB) was added and incubated at  $37^{\circ}$  C. for 15 min. Aliquots were loaded into a DNA 7500 LabChip and were analyzed on a Bioanalyzer 2200. The workup steps served to release Cas9 from binding to target DNA, which were assayed for cleavage.

A series of guide RNAs as listed in FIG. 4 were chemically synthesized. Briefly, individual RNA strands were synthesized and HPLC purified. All oligonucleotides were quality control approved on the basis of chemical purity by HPLC analysis and full-length strand purity by mass spectrometry analysis. Each of these guide RNAs was designed to target the human CLTA gene.

The results are shown in FIG. 4. As shown in the Table 1 of FIG. 4, all but one of the chemically synthesized guide RNAs targeted and cleaved the CLTA-encoded DNA target sequence with significant cleavage rates. The one exception was "CLTA\_37\_Deoxy" guide RNA, which had a contiguous sequence of 37 deoxyribonucleotides at its 5' end.

As disclosed herein, a variety of chemical modifications were tested at specific positions in the sequence of a guide RNA. Surprisingly, the tested positions in the guide sequence of the guide RNA (a.k.a. the spacer sequence in the guide RNA) tolerated most of the modifications tested,

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including combinations of multiple modifications within single nucleotides in the guide RNA, even when modifications were instantiated in the target-binding sequences.

The results revealed that guide RNAs containing modifications at specific positions were tolerated by active Cas protein and gRNA:Cas protein complexes, as the modifications did not prevent target-specific cleavage of the target polynucleotide. In all the guide RNA sequences listed in the Table 1 of FIG. 4, the first 20 nucleotides at the 5' end are complementary to the target sequence in target DNA. The modifications that were tested and found to be tolerated at specific positions include 2'-O-methylribonucleotide (=2'OMe), 2'-deoxyribonucleotide, racemic phosphorothioate internucleotide linkage(s) (=P(S)), 3'-phosphonoacetate (=PACE), 3'-thiophosphonoacetates (=thioPACE), Z nucleotides, and combinations of these.

It is contemplated that the chemical modifications disclosed and tested herein, particularly at the tested positions (as listed in the Table 1 of FIG. 4), will be tolerated at equivalent positions in a variety of guide RNAs. In certain embodiments, the chemical modifications disclosed and tested herein are tolerated in any position in a guide RNA.

As disclosed herein, chemically modified nucleotides were incorporated into guide RNAs in an effort to improve certain properties. Such properties include improved nuclease resistance of the guide RNA, reduced off-target effects of a gRNA:Cas protein complex (also known as improved specificity), improved efficacy of gRNA:Cas protein complex when cleaving, nicking or binding a target polynucleotide, improved transfection efficiency, and/or improved organelle localization such as nuclear localization.

While the use of modified RNA is known (e.g., to block nucleotlytic degradation in certain applications), it is widely known that one cannot simply incorporate modifications at any or all positions in an RNA sequence and expect it to 68

function, particularly when the RNA sequence needs to complex with a protein or an enzyme to exert certain functions. Thus, it was not predictable whether these guide RNAs could tolerate chemical modifications at a variety of nucleotide positions while performing sufficient or improved function in a CRISPR-Cas system. In fact, it was unexpected that the guide RNA can tolerate specific modifications to the extent instantiated and tested, especially at several of the positions tested.

#### Example 2

To evaluate the ability of the chemically synthesized guide RNAs to target and cleave a DNA target sequence, an in vitro cleavage assay similar to that described in Example 1 was used. Target DNA constructs were for human DNA targets (sequences from the human clathrin light chain (CLTA1) gene, the human Interleukin 2 Receptor Gamma (IL2RG) gene, the human cytotoxic T-lymphocyte-associated protein 4 (CLTA4) gene, the human protocadherin alpha 4 (PCDHA4) gene, and the human engrailed homeobox 1 (EN1) gene), along with off-target DNA constructs differing from the target DNA by one or more nucleotides.

Table 3 sets forth the guide RNA constructs and their sequences, along with DNA constructs used for assessing the ability of those guide RNA constructs to target and cleave. In all the guide RNA sequences listed in the Table 3, the first 20 nucleotides at the 5' end are complementary to the target sequence in target DNA. ON target constructs comprise the 20 nt target sequence. OFF target constructs comprise most of the same 20 nucleotides as the target DNA, with 1, 2 or 3 nucleotide differences. Accordingly, the guide RNA is mostly, but not completely, complementary to the sequence of the OFF target constructs. The OFF target constructs are based on gene sequences known to occur in the human genome.

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Entry # Guide RNA Construct  Construct  2-piece  Unmodified  1 CLTA1 CRNA + tracrRNA CLTA1 ON1- target  2 CLTA1 CRNA + tracrRNA CLTA1 ON1- target  4 CLTA1 CRNA + tracrRNA CLTA1 OFF1- target  5 CLTA1 CRNA + tracrRNA CLTA1 OFF2- target		RNA length
CLTA1 CIRNA + tracirna CLTA1 ON1- target  CLTA1 CIRNA + tracirna CLTA1 ON1- target  CLTA1 CIRNA + tracirna CLTA1 OFF1- target  CLTA1 CIRNA + tracirna CLTA1 OFF1- target  CLTA1 CIRNA + tracirna CLTA1 OFF1- target	RWA sequence (5'→3')	
CLTA1 CTRNA + tracrRNA CLTA1 CTRNA + tracrRNA CLTA1 CTRNA + tracrRNA CLTA1 CTRNA + tracrRNA	2-piece dual-guide scaffold Unmodified dual-guide RNA (dgRNA)	
CLTA1 CRNA + tracrRNA CLTA1 CRNA + tracrRNA CLTA1 CRNA + tracrRNA	AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGUUUUGAAUGGUCC CAAAAC (SEQ ID NO: 25) + GGAACCAUUCAAAACAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA	98 + 95
CLTA1 CTRNA + tracrRNA CLTA1 CTRNA + tracrRNA CLTA1 CTRNA + tracrRNA	ACUUGUAAAAGUGGCACCGAGUCGGUGCUUUUUU (SEQ ID NO: 26) AGUCCUCAUCUCCUCAAGCGUUUAAGAGCUAUGCUGUUUUGAAUGGUCC CAAAAC (SEQ ID NO: 25) + GGAACCAUUCAAAACAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA	9 + 20 + 21 +
CLTA1 CrRNA + tracrRNA CLTA1 target CLTA1 CrRNA + tracrRNA CLTA1 target	ACUNGUAAAGUGCCACCCAGUCGUUGCUUUUUU (SEQ ID NO: 26) AGUCCUCAUCCCUCAAGCGUUUAAGAGCUAUGCUGUUUGAAUGGUCC CAAAAC (SEQ ID NO: 25) + GGAACCAUUCAAAAGCAUAAGAGUUUAAAUAAGGCUAGUCCGUUAUCA	98 + 99
CLTA1 crRNA + tracrRNA CLTA1 target	AGUCCUCARAGOGGCACCGAGGGGCUOOOOOOOOOOOOOOOOOOOOOOO	98 + 96
	AGUCCUCARCCUCAACCUUUAAGAGCUAUUUAAAUACUUUUAAAUGGUCC CAAAAC (SEQ ID No: 25) + GUUUAAAAGAGCUAGUUAAAUAAGGCUAGUCCUUAUCA AGAACCAUUCAAAAGAGCAUAAGAGAGUUAAAUAAGAGCUAGUCCUUAUCA	98 + 99
6 CLTA1 crRNA + tracrRNA CLTA1 OFF2- target	AGUCCUCAPATAGOGGCACCGGGGGCUOOOOOOOOOOOOOOOOOOOOOOOO	98 + 95
7 CLTA1 crRNA + tracrRNA CLTA1 OFF3- target	ACUGGOALARGOGGCACCGAGGGGGGGGGGGGGGGGGGGGGGGGGGGG	-56 + 86
8 CLTA1 crRNA + tracrRNA CLTA1 OFF3- target	AGUCGUGAAAAGGGGGACCGGGGGGCUOOOOOOOOOOOOO	98 + 95
9 CLTA1 crRNA + tracrRNA CLTA1 target	AGUCCUCAAGGGGGCGGGGGGGGGGGGGGGGGGGGGGGG	98 + 96
10 IL2RG_CTRNA + IL2RGrrig ON- tracrRNA target		96 + 56 +

TABLE 3

			TABLE 3-continued	
Entry	Entry # Guide RNA Construct	Target DNA Construct	RNA sequence (5'→3')	RNA length
		P1.	Fluorophore-coupled dgRNA	
11	CLTA1 CTRNA + tracrRNA aminoallyl- US7 + Cy5	CLTA1 ON1- target	AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGUUUUGAAUGGUCC CAAAAC (SEQ ID NO: 29) + GGAACCAUUCAAAACAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA ACUUG(aminoallylu + Cy5) AAAAGUGGCACCGAGUCGGUGCUUUUU (SEQ ID NO: 30)	9 + 9 9
		. 2	2'OMethyl-modified dgRNA	
12	<pre>IL2RG_crRNA_5', 3'- 3x(2'OMe) + tracrRNA_5', 3'- 3x(2'OMe)</pre>	IL2RGmg ON- target	UGGUAAUGAUGGCUUCAACAGUUUUAGAGCUAUGCUGUUUUGAAUGGUC CCAAAAC (SEQ ID NO: 31) + GGAACCAUUCAAAACAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA ACUUGUAAAAGUGGCACCGAGGGCGUGCUUUUUU (SEQ ID NO: 32)	98 + 20 20 4
		2'OMethyl,	3'Phosphorothioate-modified dgRNA	
13	<pre>IL2RG crRNA 5', 3'- 3x(2'OMe, 3'P(S)) + tracrRNA 5', 3'- 3x(2'OMe, 3'P(S))</pre>	IL2RGmg ON- target	<pre>UsGsgsUaAUGGCUUCAACAGUUUUAGAGCUAUGCUGUUUUGAAUGG UCCCAASASASC (SEQ ID NO: 33) + GsGsAaCGUUCAAAACAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAU CAACUUGUAAAAGUGGCACCGAGUCGGUGCUUUsususu (SEQ ID NO: 34)</pre>	98 + 95
		2'OMethyl, 3	3'PhosphorothioPACE-modified dgRNA	
14	<pre>IL2RG_crRNA_5', 3'- 3x(2'OMe, 3'thioPACE) + tracrRNA_5', 3'- 3x(2'OMe, 3'thioPACE)</pre>	IL2RGmg ON- target	<u>U*sG*sG*sUrangauggcuucaacaguuuuraargcuaugcuguuugaau</u> ggucccaa*sa*sa*sc (seq id no: 35) + <b>g*sg*sa</b> *saccauucaaacagcauagcaaguuuaaauaaggcuaguccgu uaucaacuuguaaaaguggcaccaagucggugcuuu <u>u</u> *s <u>u</u> *su (seq	98 + 29
15	<pre>IL2RG_crRNA_5', 3'- 1x(2'OMe, 3'thioPACE) + tracrRNA_5', 3'- 1x(2'OMe, 3'thioPACE)</pre>	IL2RGmg ON- target	ID NO: 36)  UD NO: 36)  UVAGAGARGAUGGCUUCAACAGUUUUAGAGCUAUGCUGUUUUGAAUGGU  CCCAAAAASC (SEQ ID NO: 37) +  G*sGAACCAUUCAAAACAGCAUAGCAAGUUUAAAUAAGGCUAGUCGUAU  CAACUUGUAAAAGGGCACCGAGUCGGUGCUUUU <u>U</u> *sU (SEQ ID NO: 38)	98 + 92
		2	2-thioU-modified dgRNA	
16	CLTAL_2thioU + 3 crRNA + tracrRNA	CLTA1 ON1- target	AG (2sU) CCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGUUUUGAAUGGU CCCAAAAC (SEQ ID NO: 39) + GGAACCAUUCAAAACAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA	98 + 89
17	CLTA1_2thioU + 3 crRNA + tracrRNA	CLTA1 ON1- target	ACUUGUAAAAGUGGCACCGAGUCGGUGCUUUUUUU (SEQ ID NO: 26) AG(2sU) CCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGUUUGAAUGGU CCCAAAAC (SEQ ID NO: 39) + GGAACCAUUCAAAACAGUUAAAUAAGGCUAGUCCGUUAUCA ACATHCAAAAACAGUCAAAACAGUUAAAUAAGGCUAGUCCGUUAUCA	98 + 20

			73		US	10,90	U,U34 I	32		74		
RNA length	56 + 86	56 + 86 56 + 86	56 + 86	56 + 86	56 + 86 5 + 86	56 + 86 54 + 86	56 + 86	26 + 86	26 + 86	98 + 99	98 + 89	56 + 86 56 + 86
RNA sequence (5'→3')	AG (2sU) CCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGUUUUGAAUGGU CCCAAAAC (SEQ ID NO: 39) + GGAACCAUUCAAAAACGCAAGGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA ACHIICIIAAAAACHGCCAACHCGGICCHIIIIIIIII (SEO ID NO: 26)	AG(2st) CCUCACUCCOCCACAGCGUUDAAGAGCUAUGCUGUUUGAAUGGU CCCAAAAC (SEQ ID NO: 39) + CCCAAACCGUUUAAGAGCCUAUGCAAGCGUUUCAAGACCAACACAACAACAACAACAACAACAACAACAACA	ACOSGUARAGOGGCACCGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	ACCOACACCACCAGAGCGUUAAGAGCGUUUGAAGAGCUUUGAAUGGU CCCAAAAC (SEQ ID NO: 39) + GGAACCAUUCAAAACAGCAGUUAAAAGAGCUUGUUGAAUGGU	AG (2st) CCUCACUCCCCCAAACGUUUAAGAGCUAUGCUAUUUGAAUGGU CCCAAAAC (SEQ ID NO: 39) + CCCAAACCGUUUAAGAGAGCUAUGCAAUGGU CGCAAAACAGCAGCAAGAACAGAGAGUUAAAUAAAGAAGAGAGAG	ACCOMMENSOR OF THE STATE OF THE	ACCOCOMMENTATION OF THE STATE O	AGUCCUCA (280) CUCCOCCAGAGGUUDAAGAGCUAUGCUGUUUGAAUGGU CCCAAAAC (280) ID NO: 40) + GGAACCAUUCAAAACAGCAUAGCAAGUUDAAAUAAGGCUAGUCCGUUAUCA ACHIIGITAAAAAACAGCAAGCAAGUUDAAAUAAGGCUAGUCCGUUAUCA	AGUCCUCA (280) CUCCCUCAGGGUUDAAGAGCUAUGCUGUUUGAAUGGU CCCAAAAC (280 ID No: 40) + GGAACCAUUCAAAAAGGGAAGUUDAAAUAAGGCUAGUCCGUUAUCA ACUUGUAAAAGGGCACCAAGCCAAG	AGUCCUCA (28U) CUCCCUCAAGCGUUUAAGAGCUAUGCUGUUUUGAAUGGU CCCAAAAC (SEQ ID NO: 40) + GGAACCAUUCAAAACAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA ACUUGUAAAAGUGGCACCGAGUCGGUGCUUUUUUU (SEO ID NO: 26)	AGUCCUCA (25U) CUCCCUCAAGCGUUUAAGAGCUAUGCUGUUUGAAUGGU CCCAAAAC (5EQ ID No: 40) + GGAACCAUUCAAAACAGCUAGCAAGUUAAAUAAGGCUAGUCCGUUAUCA ACUUGHAAAAGHGGCACCGAGUCCGAGUCCGAGUCGUUUUU (5RO ID NO: 26)	AGUCCUCA (28U) CUCCCUCAAGCGUUUAAGAGCUAUGCÜGUUUUGAAUGGU CCCAAAAC (SEQ ID NO: 40) + GGAACCAUUCAAAACAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA ACUUGUAAAAGUGGCACCGAGUCGGUGCUUUUUU (SEQ ID NO: 26)
Target DNA Construct	CLTA1 OFF1- target	CLTA1 OFF1- target	CLTA1 OFF2- target	CLTA1 OFF2- target	CLTA1 OFF3- target	CLTA1 OFF3- target	CLTA1 ON1- target	CLTA1 ON1- target	CLTA1 OFF1- target	CLTA1 OFF1- target	CLTA1 OFF2- target	CLTA1 OFF2- target
# Guide RNA Construct	CLTA1_2thioU + 3 crRNA + tracrRNA	CLTA1_2thioU + 3 crRNA + tracrRNA	CLTA1_2thioU + 3 crRNA + tracrRNA	CLTA1_2thioU + 3 crRNA + tracrRNA	CLTA1_2thioU + 3 crRNA + tracrRNA	CLTA1_2thioU + 3 crRNA + tracrRNA	CLTA1_2thioU + 9 crRNA + tracrRNA	CLTA1_2thioU + 9 crRNA + tracrRNA	CLTA1_2thioU + 9 crRNA + tracrRNA	CLTA1_2thioU + 9 crRNA + tracrRNA	CLTA1_2thioU + 9 crRNA + tracrRNA	CLTA1 2thioU + 9 crRNA + tracrRNA

TABLE 3-continued

Entry	# Guide RNA Construct	Target DNA Construct	RNA sequence $(5' \rightarrow 3')$	RNA length
30	CLTA1_2thioU + 9 crRNA + tracrRNA	CLTA1 OFF3- target	AGUCCUCA (2sU) CUCCCUCAAGCGUUUAAGAGCUAUGCUGUUUUGAAUGGU CCCAAAAC (SEQ ID NO: 40) + GGAACCAUUCAAAACAGCAUAAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA	98 + 99
31	CLTA1_2thioU + 9 crRNA + tracrRNA	CLTA1 OFF3- target	ACUGUAAAAGUGGCACCGAGUGCUUUUUUU (SEQ ID NO: 26) AGUCCUCA(2sU) CUCCCUCAAGCGUUUAAGAAGCUAUGCUGUUUUGAAUGGU CCCAAAAC (SEQ ID NO: 40) + GGAACCAUUCAAACAGCAUGAAGUAAAUAAGGCUAGUCCGUUAUCA	98 + 95
32	CLTAl_2thioU + 11 crRNA + tracrRNA	CLTA1 ON1- target	ACUGUAAAAGUGGCACCGAGUCGGUGCUUUUUU (SEQ ID NO: 26) AGUCCUCAU(2sU) CCCUCAAGCGUUUAAAGAGCUAUGCGGUUUGAAUGGU CCCAAAAC (SEQ ID NO: 41) + GGAAACCUUCAAAACAGCAUAGCAAGUUUAAAUAAAGCGUUCGUU	98 + 99
33	CLTA1_2thioU + 11 crRNA + tracrRNA	CLTA1 ON1- target	ACUDGAAAAQUGACACCGAGUCGUUUUUU (SEQ ID NO: 26) AGUCCUCAUC (2sU) CCCUCAAGCGU UUAAGAGCUAUGCUGUUUGAAUGGU CCCAAAAC (2sQ ID No: 41) + GGAACCAUUCAAAACAGCAAGGCAAGUUUAAAUAAGGCUAGUCCGUUAUCA	98 + 99 2
34	CLTA1_2thioU + 11 crRNA + tracrRNA	CLTA1 OFF1- target	ACUCCUCANC (2st) CCCUCAAGCGUUDAGAGCUAUGCUGUUUGAAUGGU CCCAAAAC (2st) IN NO: 41) + GGAACCAUGAAACAGCAAUGAGAGUUDAAAGAGCUAGGUCGUUAUCA ACUTGUA AAAACAGCAAUGCAAGUUDAAAUAAGGCUAGUUCGUUAUCA	98 + 95
35	CLTA1_2thioU + 11 crRNA + tracrRNA	CLTA1 OFF1- target	AGUCCUCATO CCCCCAAGGU UUAAGAGCUUUGAAUGU CCCCAAAAC (2sV) CCCCCCAAGGU UUAAGAGCUUUGAAUGGU CCCAAAAC (2sV) NO: 41) + GGAACAUUCAAAAAAGGAGUUAAAUAAGGUUAAAUAGCUUAAAAACAGUUAAAAACAGUUAAAAAGGUUAAAAACAGUUAAAAAAGUUAAAACAGUUAAAAAAGUUAAAAAAGUUAAAAAAAA	98 + 99
36	CLTA1_2thioU + 11 crRNA + tracrRNA	CLTA1 OFF2- target	AGUCCUCAUC (2st) CCCUCAAGCGUUDAAGAGCUAUGCUUUUGAAUGGU CCCAAAAC (SEQ ID NO: 41) + GGAACCAUUCAAAACAGCAAUGCAAGGGUUAAAGAGCCGUUAACA ACHIGIPAAAACAGCCAUGCAAAGCAGCAAACAGCAAGACAGCAAAACAGCAAAACAGCAAAACAGCAAAACAGCAAAACAGCAAAACAGCAAAACAGCAAAACAGCAAAACAGCAAAACAGCAAAAACAGCAAAAACAGCAAAAACAGCAAAAACAGCAAAAACAGCAAAAAA	98 + 99
37	CLTA1_2thioU + 11 crRNA + tracrRNA	CLTA1 OFF2- target	AGOCCUCANC (250) CCCUCAAGCGUUDAAGAGCUAUGCUUUUGAAUGGU CCCAAAAC (550) ID NO: 41) + GGAACCAUUCAAAACAGCAAUGCAAGGGUUDAAAUAAGGCAAGUUUAAAAUAGCAAUUCAAAACAGCCAUGCAAAGCAGUUAAAUAAA	98 + 99
80	CLTA1_2thioU + 11 crRNA + tracrRNA	CLTA1 OFF3- target	AGUCCUCAUC (280) CCCUCAAGCGUUDAAGAGCUAUGCUGUUUGAAUGGU CCCAAAAC (520 ID No: 41) + GGAACCAUUCAAAACAGCAAUGCAAGGUUDAAAUAAGGCAAGUUUAAAAUAGGUUAAGAUAGGUUAAAACAGUUAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAACACAAACAGUUAAAACAGUUAAACAGUUAAAACAGUUAAAACAGUUAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAGUUAAAACAAGUUAAAACAGUUAAAACAGUUAAAAACAGUUAAAAACAGUUAAAAACAGUUAAAAACAGUUAAAAACAAGUUAAAAACAGUUAAAAACAGUUAAAAACAGUUAAAAACAGUUAAAAACAGUUAAAAACAGUUAAAAACAGUUAAAAACAGUUAAAAAAAA	98 + 95
<u>ა</u>	CLTA1_2thioU + 11 crENA + tracrENA	CLTA1 OFF3- target	CCCCAAGCGUUUAAGAGCUAUGCUGUUUGAA NO: 41) + SCAUAGCAAGUUUAAUAAGGCUAGGCCGUU SCAUAGCAAGUUUAAUAAGGCUAGUCCGUU CCCAGGUGGUGCUUUUUU (SEQ ID NO:	98 + 9
40	CLTA1 sgRNA (Batch #1)	CLTA1 ON1- target	Unmodified single-guide RNA (sgRNA) AGUCCUCAUCUCCUCAAGGGUUUAAGAGGUAUGGUGGUAACAGGAUAGGA AGUUUAAAUAAGGCUAGUCCGUUAUCAAAAAAGUGGCACCGAGUC	113
41	CLTA1 sgRNA (Batch #1)	CLTA1 ON1- target	GGUGCUUUUUU (SEQ ID NO: 42) AGUCUCAUUUCACUCAAGCGUUUAAGAAGCUUUAAACAGCUGAAGCGUUUAAACUUGACAACUUGAAACUUGAAAAAGUGCAACCGAGUC	113

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Entry	# Guide RWA Construct	Target DNA Construct	RNA sequence (5'→3')	RNA length
42	CLTA1 sgRNA (Batch #2)	CLTA1 ON1- target	AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAAGUGGCACCGAGUC	113
43	CLTA1 sgRNA (Batch #2)	CLTA1 ON1- target	GGUGCUUUUUU (SEQ ID NO: 42) AGUUCAGAUCCCUCAAGGGUUUAAGAGCUAUGCUGGUAAACAGCAUAGCA AGUUUAAUAAGGGUUAUCAACUUAUCAACAGGAAGGGGCACCGAGUC GGUGCUUUUUUUU (SCO ID NO: 42)	113
44	CLTA1 sgRNA (Batch #3)	CLTA1 ON1- target	AGUCCUCAUCUCCCUCAAGGGUUDAAGAGCUAUGCUGGUAACAGGAUAGCA AGUUUAAUUAAQAGGCUGGUACCACGUUGAAAAAAGUGGCACCGAGUC	113
45	CLTA1 sgRNA (Batch #3)	CLTA1 ON1- target	GGUGCUGUOUU (SEQ ID NO: 420) AGUUUAAUUAAGGCUAGCGUUUUAAGAGCGUAUGCAGAAAAGGCACGAGCA AGUUUAAAUAAGGCUAGUCAACUUGAAAAAAGUGGCACCGAGUC	113
46	CLTA1 sgRNA (Batch #3)	CLTA1 ON1- target	GGUGCUUUUUU (SEQ ID NO: 42/) AGUUUAAUAAGGCUAAGGCUUUAAGAGCUAUGCUGGUAAAAAGGCACGAGCA AGUUUAAAUAAGGCUAGUCAACUUAUCAACUUGAAAAAGUGGCACCGAGUC	113
47	CLTA1 sgRWA (Batch #3)	CLTAlmg ON1- target	GGGGCOCCOCC (SEX ID NO: 420) AGUUDAAQAAGGCUGGUAACAAGUGGGGAAAAAGGGAAAAAGGGAAGGGAAGGCAAGGGUAACAACUGAAAAAAGGGCACCGAGCCAACCGAGCCAACCGAACAACUGAAAAAAGGGCACCGAGCCAACCGAGCCAACCGAACAACA	113
48	CLTA1 sgRNA (Batch #3)	CLTAlmg ON1- target	GGUGCUUUUUU (SEY ID NO: 42) AGUUUAAUAAGGGUUAUGAAGAGGUAUGCUGGUAAGGAGCAUAGCA AGUUUAAAUAAGGGUUAUCAACUUGAAAAAGUGGCACCGAGUC	113
49	CLTA1 sgRNA (Batch #3)	CLTAlmg ON1- target	AGUCCOCOCOCCCCCAAGCGUUDAAGACUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGCGUUAUCAACUUGAAAAAGUGGCACCGAGCC AGUUUAAAUAAGCCUAGUCAACUUGAAAAAGUGGCACCGAGUC	113
20	CLTA1 sgRNA (Batch #3)	CLTAlmg OFF1- target	GGUGCUUUUUU (SEQ ID NO: 427) AGUUCCUCAUCCCUCAAGCGUUUDAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAAGCGUUAUCAACUUGAAAAAGUGGCACCGAGUC	113
51	CLTA1 sgRNA (Batch #3)	CLTAlmg OFF3- target	GGUGCUUUUUU (SEQ ID NO: 420 AGUUCUCAUCCUCCAUCAGGUUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCAACUUGAAAAAAGUGGCACCGAGUC	113
52	CLTA1 sgRMA (crude)	CLTA1 ON1- target	GGUGCUUUUUU (SEQ 1D NO: 42) AGUUCAGAUCCUCCAUCAGAGGUUUAAGAGCUAUGCUGGUAAAUAAGGCUAAGCAUAACAGCUUAUCAACUGAAAAAGUGGCACCGAGUC	113
53	CLTA1_Bos sgRNA	CLTAlmg ON1- target	GGGGCOGGGGG (SEE ID NO: 42) AGUCCUCAUCUCCCUCAAGCGUUUNAGAGCAAGUUAAAAUA AGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUGGUUCUUU (ADO ID NO 42)	100
54	CLTA1_Bos sgRNA	CLTAlmg ON1- target	(SEQ ID NO: 43) AGUCCUCAUCUCCCUCAAGCGUUUUAGAGCUAGUAAUAGCAAGUUAAAAUA AGGCUAGUCGUUAUCAACUUGAAAAGUGGCACCGAGUGGUUGCUUUU (CEC ID NO: 42)	100
55	CLTA1_Bos sgRNA	CLTAlmg ON1- target	(SEQ ID NO: 43) AGUCCUCAUCUCCCUCAAGCGUUUUAGAGCUAGUAAUAGCAAGUUAAAAUA AGGCUAGUCGUUAUCAACUUGAAAAGUGGCACCGAGUGGUUGCUUUU (CFC ID NO: 42)	100
26	CLTA1_Bos sgRNA	CLTAlmg OFF1- target	(SEG ID NO. 45) AGUCCUCCUCCAGCGUUUUAGAGCUAGUAAGAAGUUAAAAUA AGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUU	100
57	CLTA1_Bos sgRNA	CLTA1mg OFF3- target	(SEQ ID NO: 43) AGUCCUCAUCUCCCUCAAGCGUUUVAGAGCUAGUAAUAGCAAGUUAAAAUA AGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUU //CEO ID NO: 42)	100

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Entry	# Guide RNA Construct	Target DNA Construct	RNA sequence (5'→3')	RNA length
58	CLTA4 sgRNA	CLTA4 ON- target	GCAGAUGUAGUGUUUCCACAGUUUAAGAGCUAUGCUGGAAACAGCAUAGC	113
50	CLTA4 sgRNA	CLTA4 ON- target	CGGUG-CUUUUUU (SEQ 1D NO: 44) GCAGAUGHAGUUUAAGUUUCCACAGUUUAAGGGAAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGGCACCGAGU CGGIGCHIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	113
09	CLTA4 sgRNA	CLTA4 ON- target	CGGGGACOUGOOO (SEG ID NO: 44) GCAGAUGAGGUGUUCCACAGUUUAAGAGGCUAUGGAAAACAGCAUAGC AAGUUUAAAUAAGGCUGGUGAAAAAGGGCACCGAGU CGGGCAGUUIAAHUAAGGCUGUUAAAAAAAAGGCUAAGU	113
61	CLTA4 sgRNA	CLTA4mg ON- target	CGGGGGCUOUGUO (25E, ID NO: 44) GCAGAUGAGUGUUCCACAGUUUAAGAGGCUAUGGAAAACAGCAUAGC AAGUUDAAAUAAGGUCGGUGUUGAAAAAGGCCACCGAGU CGGIGCHIIIIIIIIII (SPO ID NO: 44)	113
62	CLTA4 sgRNA	CLTA4mg ON- target	GGGGGAUGUGGO (SEE IL NO. 11) GCAGANGUGAGUGUCACAGUUNAAGAGCUAUGGAAACAGCAUAGC AAGUUDAAAAAGGCUGGUAUCAACUUGAAAAAGGCACCGAGU CGGIGGIIIIIIIIIII (SEO ID NO. 44)	113
63	CLTA4 sgRNA	CLTA4mg OFF5- target	GCGGAGGGGGGGGGGGGGGGGGGGAACAGCGGGAACAGCGGGGGG	113
64	CLTAl_Truncated_18 mer	CLTAlmg ON1- target	UCCUCAUCUCCCUCAAGCGUUDAAGACUAUGCUGGUAACAGCAUAGCAAG UUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGG UGCUUUUUUU (SEO ID NO: 45)	111
65	CLTAl_Truncated_18 mer	CLTAlmg ON1- target	UCCUCAUCUCCCUCAAGCGUUDAAGAGCUAUGCUGGUAACAGCAUGGAAG UUUAAAUAAGGCUAGCUCGUUDAUCAACUUGAAAAAGUGGCACCGAGUCGG HGCHHHHHHHH (SEO 1D NO. 45)	111
99	CLTAl_Truncated_18 mer	CLTAlmg OFF1- target	UCCUCAUCUCCCUCAGGGUUNAAGAGCUAUGCUGGUAACAGCAUAGGAAG UUUAAAUAAGGGCUAGUCGGUAACUUGAAAAAGUGGCACCGAGUCGG IIGCUIIIIIIIIII (SC) D NO: 45)	111
67	CLTA1_Truncated_18 mer	CLTAlmg OFF3- target	UCCUCAUCUCC (122 12 10) 12) UCCUCAUCUCC (122 12 10) 12) UUJAAAUAAGGCUAAGGGUUJAAGAGCUAAGUAGAAAAGUGGCACCGAGUCGG UGCUUJUUU (SC) ID NO: 45)	111
8 9	CLTA1_Truncated_17 mer	CLTAlmg ON1- target	CCCCAUCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	110
69	CLTAl_Truncated_17 mer	CLTA1mg ON1- target	CCUCAUCUCCCUCAAGGGUUUAAGAGCUAUGCUGGUAACAGCAUAGCAAGU UUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGU GCUUUIIIIIII (SRO ID NO. 46)	110
70	CLTAl_Truncated_17 mer	CLTAlmg OFF1- target	CCCCAUCCCCUCAAGGGUUDAAGAGCUAUGGUAACAGCAUAGCAAGU UUAAAUAAGGGUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGU GCUUUUUUU (SRO ID NO: 46)	110
71	CLTAl_Truncated_17 mer	CLTAlmg OFF3- target	CCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCAAGU UUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGU GCUUUUUUU (SRO ID NO. 46)	110
72	CLTA1_1xExtraG	CLTAlmg ON1- target	GAGUCCUCAUCUCCCUCAAGCGUUAAGAGCUAUGCUGGUAACAGCAUAGC AAGUUUAAUAAGGCUAGUCCGUUAACAACUUGAAAAAGUGCCACCGAGU CGGUGCUUUUUUU (SEO ID NO: 47)	114
73	CLTAl_lxExtraG	CLTAlmg ON1- target	GAGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGIGCIIIIIIIIIII (SRO ID NO 47)	114

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Entry	# Guide RNA Construct	Target DNA Construct	RNA sequence (5'→3')	RNA length
74	CLTA1_1xExtraG	CLTAlmg OFF1- target	GAGUCCUCAUCUCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGC AAGUUUAAAUAAGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGU	114
75	CLTA1_1xExtraG	CLTAlmg OFF3- target	CGGUGCUUUUUU (SEQ ID NO: 47) GAGUCCUCACCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAGU CACGIGCHIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	114
76	CLTA1_2xExtraG	CLTA1mg ON1- target	CAGGGCCOCOUN (SEG 1D NO: 41) GCAGUCCUCAUCCCCCCAAGCGUUGAAGAGCUGGDAACAGCGAUAG CAAGUUUAAAGAGCGUUGAAAGUUGAAAAGUGGCACCGAG TCCCTTCCTTTCTTTTTTTTTTTTTTTTTT	115
77	CLTA1_2xExtraG	CLTAlmg ON1- target	GCGGUGCUUUUU (SEQ ID NO: 48) GCAGUUUAAUCCCCCCACAGCGUUUAAAGAGCUGAAGUUGAAAAGCGAGG CAAGUUUAAAUAAGCCGUUACCGUUAACACUUGAAAAGUGGCACCGAG	115
78	CLTA1_2xExtraG	CLTAlmg OFF1- target	UCGGUGCUUUUUU (SEQ 1D NO: 48) GGAGUCUCAUCUCCCUCAAGCGUUUAAAGAGCUAUGAAACAGCAAG CAAGUUUAAAUAAGGCUACCGUUAUCAACUUGAAAAGUGGCACCGAG HCGGIGCHIITHIITHII (FOO 1D NO: A9)	115
79	CLTA1_2xExtraG	CLTA1mg OFF3- target	GCGGGCGCGCGCGCGCGCGCGCGGGGGGGGGGGGGGGG	115
80	CLTA1_63U,64U	CLTA1mg ON1- target	AGUCCUCAUCCUCAAGCGUUDAAGAGUAUGCUGGUAACAGCAUAGCA AGUUDAAUAAUUUAAUUCUAGCCGUUDAACAACUUGAAAAAGUGGCACCGAGUC GGUGCUUUUUUU (SRO 1D NO: 49)	113
81	CLTA163A,64A	CLTA1mg ON1- target	AGUCCUCAUCCCUCAGGGUUUAAGAGCUAUGGUAACAGCAUAGCA AGUUUAAAAAAACUAGCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUGCTIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	113
82	CLTA1_63A,64A,70U,71U	CLTAlmg ON1- target	AGUCCUCAUCCCCUCAGGGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAAACUAGUUUGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUGCUUIIIIIIIII (SRO 1D NO 51)	113
83	CLTAl_cis-block(1- 5)_polyU_sgRNA	CLTAlmg ON1- target	GAGCUUUUUUGUCCUCAUCCCUCAAGCGUUUUAGAGCUAGAAAUAG GAAGUUAAAUAAGCUAGUCCGUUAAGAGCUUUUAGAAAAAGUAGCACCGAG ICGGIGCIIIIIII (SC)	111
84	CLTAl_cis-block(1- 5)_polyU_sgRNA	CLTAlmg ON1- target	GAGCUUUUUUUU (SEG II) NO. 52) GAAGUUAAAUAAGUCCUCCUCAAGCGUUUUAGAACUAGAAAUAG CAAGUUAAAAUAAGGCGUCGUCGUCAACUGAAAAAGUGGCACCGAG UCGGUGCUUUU (SEG ID NO. 52)	111
82	CLTAl_cis-block(1- 5)_polyU_sgRNA	CLTAlmg OFF1- target	GGACUUUUUUUGUCCUCAUCUCCUCAGGGUUUUAGAGCUAGAAAUAG CAAGUUAAAUAAGGCUAGUCCGUUACAACUUGAAAAGUGGCACCGAG ICCGIGCIIIIIII (SEG ID NO. E2)	111
98	CLTA1_cis-block(1- 5)_polyU_sgRNA	CLTA1mg OFF3- target	GGGCGCCCCC (SEZ ID NO: 52) GGACCUUDADAGCCCCCCCCCCCCCCCCCCUCAAGGGCCGAGCCGA	111
87	CLTA1_cis-block(1-10)_ polyU_sgRNA	CLTA1mg ON1- target	GAUGAGGACUU (VIEG. ID. NO. 127) GAUGAGGACUUUUUUUAAUCCUCAUCUCCCUCAAGCGUUUUAAAAUGAGCAAGCUAGUCCGUUAUCCAACUGAAAAGUGGCA AAUAGCAAGUUAAAAUAAGCUAGUCCGUUAUCAACUGAAAAAGUGGCA CCCAAGIICAGGCTIITIIII (SPO. ID. NO. R.3.)	116
88	CLTA1_cis-block(1-10)_ polyU_sgRNA	CLTA1mg ON1- target	GOGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	116
8	CLTAl_cis-block(1-10)_ polyU_sgRNA	CLTA1mg OFF1- target	GAUGAGGACUUUUUUAGUCCUCAUCCCCCCAAGCGUUUUAGAGCUAGA AAUAGCAAGUUAAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCA	116

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	Entry # Guide RNA Construct	Target DNA Construct	RNA sequence $(5' \rightarrow 3')$	RNA length
90 CLTA poly	CLTAl_cis-block(1-10)_ polyU_sgRNA	CLTAlmg OFF3- target	GAUGAGGACUUUUUUUAGUCCUCAUCUCCCUCAAGCGUUUUAGAGCUAGA AAUAGCAAGUUAAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCA	116
91 CLTA poly	CLTAl_cis-block(16-20)_ polyU_sgRNA	CLTA1mg ON1- target	CCAGUGADUAGUCCOCCAGCGUUUAGAGCUAGAAAAG GCUUGUUAAAGUCCUCGUCCAGCGUUUAGAAGUGGCACCGAG CAGUUAAAAUAAGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAG	111
92 CLTA Poly	CLTA1_cis-block(16-20)_ polyU_sgRNA	CLTA1mg ON1- target	occessorovo (SEC ID NO: 94) GCUUGUUUUUAGUCCUCAUCUCCCUCAAGCGUUUAGAGCCUAGAAAUAG CAAGUUAAAAUAAGCUAGUCCGUAUCAACUUGAAAAAGUGGCACCGAG	111
93 CLTA Poly	CLTA1_cis-block(16-20)_ polyU_sgRNA	CLTAlmg OFF1- target	UCGGUGCUUUU (SEQ 1D NO: 54) GCUUGUUUUUUAGUCCCUCAAGCGUUUUAGAGCUAGAAAUAG CAAGUUAAAAUAAGGCUAGUCCCUCAAGCGUUUAAAAGUGGCACCGAG CAAGUUAAAAUAAGCUAGUCCCUUAUCAACUUGAAAAAGUGGCACCGAG	111
94 CLTA poly	CLTA1_cis-block(16-20)_ polyU_sgRNA	CLTAimg OFF3- target	GCUGGUUUUUUAGUCCUCAACCUCAAGCGUUUUAGAGCUAGAAAUAG CAAGUUAAAAAAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAG UCGGUGCUUUU (SEQ ID NO: 54)	111
		MQ	DMT-modified sgRNA	
95 CLTA	CLTA1_DMT-ON sgRNA	CLTA1 ON1- target	(dmt) AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAU AGCAAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCG	113
96 CLTA1_ sgRNA_	CLTA1_DMT-ON/OFF sgRNA	CLTA1 ON1- target	AGUCCUCAGUCCOCCAAGGGUUAAGAGGUAAGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUGCUUUUUU (SEQ ID NO: 56)	113
		Fluoro	Fluorophore-modified sgRNA	
97 CLTA	CLTA1_IntF1_sgLoop	CLTA1 ON1- target	AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGG (FI) AACAGCAUAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGU	113
98 CLTA	CLTA1_IntF1_sgLoop	CLTA1mg ON1- target	CGGGGCOCOUOUU (SEQ ID NO: 37) AGUUDAAUGAGGCUCAUGAGGGCUAUGCUGG (F1) AACAGCAUAGC AGGUUAAAUGAGGCGGUUAAAGAGGGCACCGAGU CGGGCACUTITITITITITITITITITITITITITITITITITITI	113
99 CLTA	CLTA1_IntFl_sgLoop	CLTA1mg ON1- target	CGGUGCUOUUUU (SEQ ID NO: 37) AGUUDAAUDAGGUGAGGUGAGGUGG (FI) AACAGCAUAGC AAGUUDAAUAAGGCUGUUAAGAGGUAUGCAGCAGCAGU CGGUGCUUIIIIIIIII (SEO ID NO: 57)	113
100 CLTA	CLTA1_IntFl_sgLoop	CLTAlmg OFF1- target	CGGGGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	113
101 CLTA	CLTA1_IntF1_sgLoop	CLTA1mg OFF3- target	AGCCOCACOCC (25g 12 no. 0) AAGUUAAAUAAGGCUGCGCUUAAGAGCAAAAGUGGCACCGAGU CCCICCIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	113
102 CLTA 3x(2	CLTA1_IntF1_sgLoop_5', 3'-3x(2'OMe)	CLTA1mg ON1- target	CGGGCGCCCCCCCCCCCGGGGGGGGGGGGGGGGGGGGG	113
103 CLTA 3x(2	CLTAl_IntFl_sgLoop_5', 3'- 3x(2'0Me)	CLTAlmg ON1- target	AGGCCUCAUCUCCCUCAGOUDAAGGCUUDAGCUGG (FI) AACAGCAUAGC AAGUUDAAQAAGAGCUUDAAGCUUDAAAAGGUGGAAAAGGCACGAGU CGGUGCUUUUUU (SEQ ID No: 58)	113

	RNA length	113	113	113	113	113	113	113	113	113	113	102	102	100	100	113
TABLE 3-continued	RNA sequence $(5^{\circ}\rightarrow 3^{\circ})$	AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGG (FI) AACAGCAUAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGUIGCUUUIIIIIII (SEO ID NO 58)	AGUCCCAUCACCUCAAGCGUUDAAGAGCUDUGCUGG (FL) AACAGCAUAGC AAGUUDAAUAAGGCUUGUUAACAACUUGAAAAAGUGCCACCGAGU CGGUGCUUUUUUU (SRO ID NO: 58)	ASGSUSCOUCAUCOCCCUCAAGCGUUUAAAGCUAUGCUGG (F1) AACAGCAUA GCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGA GICGGIGGIIIIIIIIAIISIIGII (SRO ID NO 59)	ASGSUSCOCOCOCCOCCOCAGGGUADAGGGUADGCUGG (F1) AACAGCAUA GCAGUUUDAAUAAGGCUAGUCGUUDUCAAGCUAAGUGAAAAGUGGCACCGA GUCGGUUUUAAUAGUGU (SEO ID NO: 59)	Asgsuscoucaucucococagegunuaagacuaugeuge (R1) aacagcaua Gcaaguuuaauaaggcuaguccguuaucaacuugaaaaaguggcaccga Gucggugcuuuusususu (SRO ID No: 59)	AAGCGUUUAAGAGCGUCAAAGAGAAAAAAAAAAAAAAAA	A*sG*sU*sccucaucuccucaagcguuuaagagcuaugcugg(F1)aacagc auagcaaguuuaaauaaggcuaguccguuaucaacuugaaaaaguggcac cgagucggugcuuuu*su*su*su (Seo ID NO: 60)	A*sg*su*scucaucoucaucoucagasasasasasasasasasasasasasasasasasasa	A*sG*su*scucalcuccucadedumaaaaguuugeugg(R1)aacagc Auagcaaguuuaaauaaggcuaguccguuaucaacuugaaaaaguggcac Cgagucggugcuuuu*su*su*su (Seq id No: 60)	A*sG*sU*sccucaucucccucaagcguuuaagagcuaugcugg(R1)aacagc Auagcaaguuuaaauaaggcuaguccguuaucaacuugaaaaaguggcac cgagucggugcuuuu*su*su*su (Seo ID NO: 60)	<pre>Uo*s(Flo) GCAGAŪGUĀGUĞUUUCCACĀGUUUAAGAGCUAGUAAUAGCAAGU UuAAAUAAGGCUAGUCGGUUA(Fl) CAACUUGAAAAAGUGGCACCGAGUCGG UGCU(Fl) U*sU (SEQ ID NO: 61)</pre>	<u>uo*s(Flo</u> ) GCAGAUGUAGUGUUCCACAGUUUAAGAGCUAGUAAUAGCAAGU UuAAAUAAGGCUAGUCCGUUA(Fl) CAACUUGAAAAAGUGGCACCGAGUCGG UGCU(Fl) U*sU (SEQ ID NO: 61)	<pre>G*scagauGuaguguuuccacaguuuaagagcuag (F1) Aavagcaaguuaa AuAagcuaguccguuaucaacuug (F1) AaAaguggcaccgag (F1) cggugc uuv*su (SEO 1D NO: 62)</pre>	<pre>g*sCagaUGUAGUGUUUCCACAGUUUAAAAGCUAG(F1) AAUAGCAAGUUUAA AUAAGGCUAGUCCGUUAUCAACUUG(F1) AAAAGUGGCACCGAG(F1) CGGUGC UUU*sU (SEQ ID NO: 62) 3'Phosphorothioate-modified sgRNA</pre>	AsGSUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGAAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGUGCUUUUUUU (SEQ ID NO: 63)
	Target DNA Construct	3'- CLTA1mg OFF1- target	3'- CLTAlmg OFF3- target	3'- CLTAlmg ON1- target	3'- CLTA1mg ON1- target	3'- CLTAlmg OFF1- target	3'- CLTAlmg OFF3- target	3'- CLTAlmg ON1- target	3'- CLTAlmg ON1- target	3'- CLTAImg OFF1- target	3'- CLTAlmg OFF3- target	CLTA4mg ON- target	CLTA4mg OFF5- target	CLTA4mg ON- target	CLTA4mg OFF5- target 3'Ph	CLTA1 ON1- target
	Guide RNA Construct	CLTAl_IntFl_sgLoop_5', 3 3x(2'0Me)	CLTAl_IntFl_sgLoop_5', 3	CLTAl_IntFl_sgLoop_5', 3 3x(2'OMe, 3'P(S))	CLTA1_IntF1_sgLoop_5', 3 3x(2'OMe, 3'P(S))	CLTA1_IntF1_sgLoop_5', 3 3x(2'OMe, 3'P(S))	CLTA1_IntF1_sgLoop_5', 3 3x(2'OMe, 3'P(S))	CLTAl_IntFl_sgLoop_5', 3 3x(2'OMe, 3'thioPACE)	CLTA1_IntFl_sgLoop_5', 3 3x(2'OMe, 3'thioPACE)	CLTAl_IntFl_sgLoop_5', 3 3x(2'OMe, 3'thioPACE)	CLTAl_IntFl_sgLoop_5', 3 3x(2'OMe, 3'thioPACE)	CLTA4_3xFl- Int_3x(2'OMe, 3'thioPACE)	CLTA4_3xFl- Int_3x(2'0Me, 3'thiopACE)	CLTA4_3xF1- Loops_3x(2'OMe, 3'thiopACE)	CLTA4 3xF1- Loops 3x(2'OMe, 3'thioPACE)	CLTA1_5'-2xP(S) sgRNA
	Entry #	104	105	106	107	108	109	110	111	112	113	114	115	116	117	118

		TABLE 3-continued	
	Target DNA Construct	RNA sequence (5'→3')	RNA length
sgRNA	CLTA1 ON1- target	AsGSUSCCUCAUCUCCUCAAGCGUUUAAGAGCUAUGGAAAACAGCAUAG CAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAG	113
sgRNA	CLTA1 ON1- target	UCGGUGCUUUUUU (SEQ 1D NO: 04) ASGSUSCSCUCAUCUCCUCAAGCUUDAAGAGCUAUGCAAACAGCAUA GCAAGUUUAAAUAAGCCCUCAAGCUUUAACAGUGAAAAGUGGCACCGA GUACCCICCHIIIIIIIIII (SEO 1) NO: 66)	113
	CLTA1 ON1- target	GUCCUCANCUCCUCACUCUCAAGCGUUAAGAGCUUAAGAACAGCAUAGCA AGUCCUCANCUCCCUCAAGCGUUAACAAGUUGAAAAAGUGGCACCGAGUC GGUGCUUUSUSUSUSU (SEQ ID NO: 66)	113
	2'OM	2'OMethyl-modified sgRNA	
	CLTA1 ON1- target	AGUCCUCAUCUCCCUCAAGGGUUUAAGAGCUAUGCUGGUAACAGCAUGGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC	113
sgRNA	CLTA1 ON1- target	AGUCCUCAUCUC (222 pp. 2010) AGUUDAAUAGCUCAGUGUAAGGCUAUGCUGUAACAGCAUGGCA AGUUDAAUAAGGCUCCGUUDAACAGUGAAAAAGUGGCACCGAGUC GGUGCUTHITIITITI (5R0 1D N0 68)	113
	CLTAlmg ON1- target	AGUCCUCAUCUCCCUCAAGCGUUDAGAGCUAUGCUGGUAACAGCAUAGCA AGUUDAAUAAGGCUUAGUCCGUUDAACAGUGGCAACGGAGUC GGGCGUUUUUUU (SEO ID NO: 68)	113
sgRNA	CLTAImg ON1- target	AGUCCUCAUCUCCCUCÂAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGIGCIIIIIIIIIIII (SCO ID NO. 68)	113
sgRNA	CLTAlmg OFF1- target	AGUCCUCAUCUCCUCAAGCGUUUAAGAGCUAUGCUGUAACAGCAUAGCA AGUUUAAUAAGGCUCGUUAGUCAACUUGAAAAAGUGGCACCGAGUC GGUGCTUTTITITITITI (SRO ID NO. 68)	113
	CLTAlmg OFF3- target	AGUCCUCAUCOC (25g T. 70. 73) AGUCCUCAUCAAGCGUUDAAGCGUAUGCUGGUAACAGCAUAGCA AGUUDAAUAAGGUUGACGGUUDAAAAGUGGCACCGAGUC GGUGCHIHHHHHH (5R0 ID NO. 68)	113
sgRNA	CLTA1 ON1- target	AGUCCUCAUCUCCUCAAGGGUUUAAGAGGUAUGCUGUAACAGCAUAGCA AGUUUAAAUAAGGUUGUCGUUAACAACUGAAAAAGUGGCACCGAGUC GGUGCUUUUUUU (SRO ID No. 69)	113
	CLTAlmg ON1- target	AGUCCUCAUCUCCUCAAGGGUUUAAGAGGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGGUUAGUCAACUUGAAAAAGUGGCACCGAGUC GGUGCUUUUUUU  100. 100. 100. 100. 100. 100. 100. 100	113
	CLTAImg ON1- target	AGUCCUCAUCUCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUGAAAAGUGGCACCGAGUC GGUGCUUUUUUU	113
sgRNA	CLTAImg OFF1- target	AGUCCUCAUCUCCUCAAGGGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGGUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUGCUUUUUUU (SEO ID NO: 69)	113
	CLTAlmg OFF3- target		113
	CLTA1 ON1- target	AGUCCUCAUCUCCCCC <mark>A</mark> AGCGUUUDAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUGCUUUUUU (SEQ ID NO: 70)	113

89 90

Entry #	# Guide RNA Construct	Target DNA Construct	RNA sequence (5'→3')	RNA length
134	CLTA1_2'OMe + 17 sgRNA	CLTA1mg ON1- target	AGUCCUCAUCUCCCUC <u>A</u> AGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAGUC	113
135	CLTA1_2'OMe + 17 sgRNA	CLTA1mg ON1- target	GGGGCCOOUGUG (SEG ID NO: 70) AGUCUUGAAQUACCUCAAGGGUUAAGAGCUAUGCUGGUAACAGGGAUAGCA AGUUUAAAUAAGGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGGGCTIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	113
136	CLTAL_2'OMe + 17 sgRNA	CLTA1mg OFF1- target	AGUCCAUCOCO (ABG ID NO. 70) AGUCUAAAUAAUAAGAGCUAUGAAAAAGUGGCAACGAGGCA AGUUUAAAUAAGGCUAGUCAUCAACUUGAAAAAGUGGCACCGAGUC	113
137	CLTA1_2'OMe + 17 sgRWA	CLTA1mg OFF3- target	(SEQ ID NO: CUCAAGCGUUU? CUAGUCCGUUAI	113
138	CLTA1_2'OMe + 17,18 sgRNA	CLTA1 ON1- target	AGUCCOCOCOCO (NEG ID NO. 70) AGUCUCARAUAAGAGCUAUGAAAAGAGCUAUGAAAAGAGCAUAGCA AGUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGGGCACCGAGUC GGIGCIIIIIIIIIIII (SR0 I) NO. 71)	113
139	CLTA1_2'OMe + 17,18 sgRNA	CLTA1mg ON1- target	AGUCCUCAUCOCOCOME A COMPANA AGUCCUGADA CAGA CAGA AGUUDA AGUUDA ADA UDA AGUUDA CAGA COCOUNTO CAGA COMBA A AGUUGA AGUUGA AGUUGA AGUUGA CAGA COCOUNTITUTUTUTUTUTUTUTUTUTUTUTUTUTUTUTUTUTU	113
140	CLTA1_2'OMe + 17,18 sgRNA	CLTA1mg ON1- target	AGUCCUCAUCUCCUC <u>AA</u> GCGUUDAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUDAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAGUC GGUCCUUUUUUU (SRO 11) NO: 71)	113
141	CLTA1_2'OMe + 17,18 sgRNA	CLTA1mg OFF1- target	CUCAAGCGUUA CUAGUCCGUUAL	113
142	CLTA1_2'OMe + 17,18 sgRNA	CLTAimg OFF3- target	AGUCCUOUCUC (25g ID NO. 71) AGUCUCAUUCACAUAGCA AGUUDAAAUAAGGCUAGUAACACUUGAAAAAGUGGCACCGAGUC AGUUTAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC	113
143	CLTA1_5', 3'-3x(2'OMe) sgRNA	CLTA1 ON1- target	AGUCCUCUCUCU (SEG ID NO: 71) AGUCUCUCUCUCAAGCGUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUC CGGCTUTUTUTTTT (SEO ID NO: 72)	113
144	CLTA4_5', 3'-3x(2'OMe) sgRNA	CLTA4mg ON- target	GCAGGOOOGOOO (ABG) ID NO: 7.2)  GCAGAGUGAGGOUCCCACAGUUDAAGAGCUAUGCUGGAAACAGCAUAGC AAGUUDAAAUAAGGUCGGUGGUCAACUUGAAAAAGUGGCACCGAGU CGGUGCUUUUUU (SRO ID NO: 73)	113
145	CLTA4_5', 3'-3x(2'OMe) sgRNA	CLTA4mg OFF5- target	GCAGAUGUAGUGUUCCACAGUUAAGAGCUAUGCUGGAAACAGCAUAGC AAGUUAAAUAAGGCUAGUCCGUUAAGAGCUAGCAACUGGAAAAGUGGCACCGAGU CGCIICCTIIIIIIIII (SFO ID NO. 73)	113
146	CLTA1_5'-20x(2'OMe) sgRNA	CLTA1 ON1- target	AGUCCUCANCUCCUCAAAGGUUDAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGTGCTTITITITITITI (SR) 17 NO: 74)	113
147	CLTA1_5'-20x(2'OMe) sgRNA	CLTAimg ON1- target	AGUCCUCAUCUCCOCOCAAGCUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC AGUUTAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC	113
148	CLTA1_5'-20x(2'OMe) sgRNA	CLTA1mg ON1- target	AGUCCUCANUCUCCUCAAAGGUUDAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUUGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGTGCTIIIIIIIIIII (SPO 11 NO: 74)	113
149	CLTA1_5'-20x(2'0Me) sgRNA	CLTAimg OFF1 target	CUCAAGCGUUDA CUAGUCCGUUAU (SEO ID NO:	113
			17	

RWA length	CAUAGCA 113 CGAGUC	CAUAGC 113 CCGAGU	CAUAGC 113 CCGAGU	CAUAGCA 113 CGAGUC	CAURGCA 113 CGAGUC	CAURGCA 113 CGAGUC	CAUAGCA CGAGUC	CAUAGCA 113 CGAGUC	CAUAGCA CGAGUC	C <u>AUAGCA</u> C <u>GAG</u> UC	C <u>AUAGCA</u> 113	C <u>AUAGCA</u> 113 C <u>GAG</u> UC	C <u>AUAGCA</u> 113	C <u>AUAGCA</u> 113	<u>uaaaaua</u> cuuu	UAAAAUA 100
RNA sequence (5'→3')	AGUCCUCAUCUCAGGUUUAAGAGCUUUAAGAGCUAGGUAACAGCAUAGGA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC	AGUCCUCOUOU (SEE, 1D NO. 71) AGUCCUCAUCUCCCUCAGGGUUUAAGAGGCUAUGAAGAGGUAACAGGAUGG AAGUUUAAAUAAGGCUAGUCGGUAUCAACUUGAAAAAGUGGCACCGAGU AAGUUUAAAUAAGGCUAGUCGGUAUAUCAACUUGAAAAAGUGGCACCGAGU	AGUCCUCAUCUCCOCAGGGUUNAAGAGCUAUGGUACAGGAUAGC AGUUUAAAUGAGCGUUUAAGAGGUUAGCAAGUAGGAGG AGUUUAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGU	AGGCGCOCOUCO (SEQ. 1D. NO.3. 76.) AGGUCGCCUCAUCCUCCAUCCUCAAGCGUUAACAAGCGUAACCAAGCAUAGCAACCAUGAAAAAGUGAACAGCGAGUCAGCACCGAAGCUAGAAAAAAAGUGGGACACCGAAGUCAGCAACUUGAAAAAAAA	AGUCCUCAUCUCCUCCAGAGGGUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUDAAAUAAGGCUGGUUAUCCACUUGAAAAAGUGGCACCGAGUC			AGUCCUCAUCUCCCOCÁAGCGUUUDAGAGGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAQAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUCCUUUUUUU (SRO ID NO: 79)	AGUCCUCAUUUCCCUCAAGCGUUUDAAGAGCUAUGCUGAAGAGCAUAGCA AGUUUAAAQAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUGCUUUUUUUU (SEO ID NO: 79)	AGUCCUCAUUUCCCCCAAGGGUUUAAGAGGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUGCUUUUUUU (SEO ID NO: 80)	AGUCCUCAUCUCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGA)AAAAGUGGCACCGAGUC GGUGCUUUUUUU (SEQ ID NO: 80)	AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUGCUUUUUUU (SBO ID NO: 80)	AGUCCUCAUCUCCCCCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUC GGUGCUUUUUUU (SEO ID NO: 80)	AGUCCUCAUCUCCCUCAAGCGUUUAAGAGGCUAUGCUGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAGUC GGUCCUUUUUUU (SEO ID NO: 80)	AGUCCUCAUCUCCCUCAAGCGUUUUAGAGCUAGUAAUAGCAAGUGAAUAAAAA AGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUU (SEO ID NO: 81)	AGUCCUCAUCUCCCUCAAGCGUUUUAGAGCUAGUAAUAGCAAGUUAAAAUA
Target DNA Construct	CLTAlmg OFF3- target	CLTA1 ON1- target	CLTA1 ON1- target	CLTAlmg ON1- target	CLTA1 ON1- target	CLTAlmg ON1- target	CLTAlmg ON1- target	CLTAlmg OFF1- target	CLTAlmg OFF3- target	CLTA1 ON1- target	CLTAlmg ON1- target	CLTAlmg ON1- target	CLTAlmg OFF1- target	CLTAlmg OFF3- target	CLTA1 ON1- target	CLTA1mg ON1-
Guide RNA Construct	CLTA1_5'-20x(2'OMe) sgRNA	CLTA1_5'-26x(2'OMe) sgRNA	CLTA1_5'-37x(2'OMe) sgRNA	CLTA1_41x(2'OMeC/U)_ QB3	CLTA1_47x(2'OMeC/U)_ QB3	CLTA1_47x(2'OMeC/U)_ QB3	CLTA1_47x(2'OMeC/U)_ QB3	CLTA1_47x(2'OMeC/U)_ QB3	CLTA1_47x(2'OMeC/U)_ QB3	CLTA1_47x(2'OMeG/A)_ QB3	CLTA1_47x(2'OMeG/A)_ QB3	CLTA1_47x(2'OMeG/A)_ QB3	CLTA1_47x(2'OMeG/A)_ QB3	CLTA1_47x(2'OMeG/A)_ QB3	CLTA1_43x(2'OMeG/A)_ Bos	$CLTAl_43x(2'OMeG/A)_$
Entry #	150	151	152	153	154	155	156	157	158	159	160	161	162	163	164	165

	RNA length	100	100	100	113	113	113	113	113	100	100	100	100	100	100	113
TABLE 3-continued	RNA sequence (5'→3')	<u>AGU</u> CCUC <u>A</u> UCUCCCUCAAGCGUUUUAGAGCUAGUAAUAGAAAUAA AGGCUAGUCCGUUAUCAACUGAAAAAGUGGCACCGAGUCGGUGCUUUU (SEO TD NO. 81)	AGGCUÇAUCUCCUCAAGCGUUUU <u>AGAGCUAGUAAUAGCAAGUUAAAAUA</u> AGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUUU (SEO TD NO: 81)	AGUCCUCAUCCCCCCCAAGCGUUUUAGAGCUAGUAAUAGCAAGUUAAAAUA AGGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAGUCGGUGCUUUU (SEO ID NO: 81)	GCAGAUGUAGUGUUUCCACAGUUUAAGAGCUAUGCAAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAGU CGGUGCUUUUUUU (SEO ID NO: 82)	GCAGANGUAGUGUUCCACAGUUAAGAGCUAUGCAAACAGCAAGCAUAGC AAGUUUAAAUAAGGCUGUCCGUUAUCAACUUGAAAAGUGGCACCGAGU CGGIGCUIIIUUUU (SRO ID NO: 82)	GCAGAUGUAGUGUUUCCACAGUUUAAGAACUAUGCUGGUAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGUGCUUUUUUU (SEQ ID NO: 83)	GCAGAUGUAGUGUUUCCACAGUUUAAGAGCUAUGCUAGUAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGUGCUUUUUUU (SEO ID NO: 83)	GCAGAUGUAGUGUUUCCACAGUUUAAGAACUAQUGGUAGUAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGUGCUUUUUU (SEO ID NO: 83)	GCAGAUGUAGUGUUUCCACAGUUUUAGAGCUAGUAAUAGCAAGUUAAAAU AAGGCUAGUCCGUUAUC <u>AA</u> CUU <u>GAAAAAGUGGCACCGAG</u> UC <u>GGUGGUUUU</u> (SEQ ID NO: 84)	GCAGAUGUAGUGUUUCCACAGUUUUAGAGCUAGUAAUAGCAAGUUAAAAU AAGGCUAGUCCGUUAUCAACUUGAAAAGUGGC <u>A</u> CC <u>GAG</u> UC <u>GGUGCUUUU</u> (SEQ ID NO: 84)	GCAGAUGUAGUGUUUCCACAGUUUUAGAGCUAGUAAUAGCAAGUUAAAAU AAGGCUAGUCCGUUAUCAACUUGAAAAGUGGC <u>A</u> CC <u>GAG</u> UCGGUGCUUUU (SEQ ID NO: 84)	GCAGAUGUAGUGUUUCCACAGUUUUAGAGCUAGUAA <u>UAGCAAGUUAAAAU</u> AAGG <u>CUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUGCUUU</u> (SEQ ID NO: 85)	GCAGAUGUAGUGUUUCCACAGUUU <u>UAGAGCUAGUAAUAGCAAGUUAAAAU</u> AAGG <u>CUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGGUCG</u>	GCAGAUGUAGUGUUUCCACAGUUUUAGAG <u>CUAGUAAUAGAAAUAGCAAGUUAAAAU</u> AAGG <u>CUAGUCGUUAUCAACUU</u> GAAAAAG <u>UGGACCGAGUCGGUGGUUUU</u> U (SEQ_ID_NO: 85) 2'Deoxy-modified sgRNA	AGTCCTCATCTCCTCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCAA GUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCG GUGCUUUUUU (SEQ ID NO: 86)
	Target DNA Construct	CLTAlmg ON1- target	CLTAlmg OFF1- target	CLTAlmg OFF3- target	CLTA4 ON- target	CLTA4 ON- target	CLTA4 ON- target	CLTA4 ON- target	CLTA4 ON- target	CLTA4 ON- target	CLTA4 ON- target	CLTA4 ON- target	CLTA4 ON- target	CLTA4 ON- target	CLTA4 ON- target	CLTAl ON1- target
	# Guide RNA Construct	CLTA1_43x(2'OMeG/A)_ Bos	CLTA1_43x(2'OMeG/A)_ Bos	CLTA1_43x(2'OMeG/A)_ Bos	CLTA4 sgRNA_5', 3'- 3x(2'OMe)	CLTA4 sgRNA_5', 3'- 3x(2'0Me)	CLTA4_47x(2'OMeC/U)_ QB3	CLTA4_47x(2'OMeC/U)_ QB3	CLTA4_47x(2'OMeC/U)_ QB3	CLTA4_49x(2'OMeG/A)_ Bos	CLTA4_49x(2'OMeG/A)_ Bos	CLTA4_49x(2'OMeG/A)_ Bos	CLTA4_39x(2'OMeC/U)_ Bos	CLTA4_39x(2'OMeC/U)_ Bos	CLTA4_39x(2'OMeC/U)_ Bos	CLTA1_5'-20x(21deoxy)
	Entry #	166	167	168	169	170	171	172	173	174	175	176	177	178	179	180

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		TABLE	NLE 3-continued	
try #	Entry # Guide RNA Construct	Target DNA Construct	RNA sequence $(5^{1}\rightarrow 3^{1})$	RNA length
181	CLTA1_5'-26x(2'deoxy)	CLTA1 ON1- target	AGTCCTCATCTCCCTCAAGCGTTTAAGGGCUAUGCUGGUAACAGCAUAGCAAG UUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAGUCGG	113
182	CLTA1_5'-37x(2'deoxy)	CLTA1 ON1- target	UGCUUUUUU (SEQ ID NO: 87)  AGTCCTCATCTCCCTCAAGGGTTAAGAGCTATGCTGUAACAGCAUAGCAAG UUDAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAGUCGG UGCUUUUUU (SEQ ID NO: 88)	113
		2'Deoxy,	3'PACE-modified sgRNA	
183	CLTA4_2'deoxy3'PACE + 15	CLTA4mg ON- target	GCAGAUGUAGUGUUU*CCACAGUUUAAGAGCUAUGGUAACAGCAUAG CAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAG	113
184	CLTA4_2'deoxy3'PACE + 15	CLTA4mg OFF5- target	GCAGANGUAGUGUUW *CCACAGUUAAGAGCUAUGCUGGUAACAGCAUAG CAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAG UCGGUGCUUUUUU (SEQ ID NO: 89)	113
		2'OMethyl	2'OWethyl, 3'PACE-modified sgRNA	
185	5'-	CLTA1mg ON1-	A*GUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGC	113
	1x(2'0Me, 3'PACE)_CLTA1 sqRNA	target	AAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGUGCUUUUUU (SEQ ID NO: 90)	
186	$5$ $1$ x(2'OMe, 3'PACE)_CLTA1 sqRNA	CLTAImg ON1- target	A*GUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGUGGCACCGAGU CGGUGCUUUUUUU (SEO ID NO: 90)	113
187	5'- 2x(2'0Me, 3'PACE)_CLTA1	CLTA1 ON1- target	A×G×UCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGGGGGAACAGCAUAG CAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAG	113
188-	sgRNA 5'-	CLTA1 ON1-	UCGGUGCUUUUUU (SEQ ID NO: 91) A*G*UCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAG	113
	2x(2'OMe, 3'PACE)_CLTA1	target	CAAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAGUGGCACCGAG UCGGUGCUUUUUU (SEO ID NO: 91)	
189	2x(2'OMe, 3'PACE)_CLTA1	CLTAlmg ON1- target	A*G*UCCUCAUCUCCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAG CAGUUUJAAUAAGUAGUCCUAGUCCAGUUAAAAAAGUGGCACGAG	113
190	sgkna 5'- 2x(2'OMe, 3'PACE) CLTA1	CLTA1mg ON1- target	OGGOCCOUDUDO (SEQ ID NO: 91)  A* <u>G</u> *UCCOUDUDO (SEQ ID NO: 91)  CAAGUUDAANDAGGOODGUCCOUDAGGOODAGGOGOCOCOGOGO  CAAGUUDAANDAGGOODGUCCOUDUDAAAAGUGGAAAGGOGOCOCOGAG	113
191		CLTA1mg ON1-	UCGGUGCUUUUUU (SEQ ID NO: 91) G*G*A*GUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA	115
	3x(2'OMe, 3'PACE)_CLTA1	target	UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACC GAGUCGGUGCUUUUUU (SEO ID NO: 92)	
192	5'- 3x(2'OMe, 3'PACE)_CLTA1 eqPNa	CLTAlmg ON1- target	G*G*A*AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCA UAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACC GACHICCCHGCHIIIIIIIIII (SPO TD NO. 92)	115
193	5. 4x(2'OMe, 3'PACE)_CLTA1	CLTA1 ON1- target	A*GTU*C*CUCAUCUCCCCUCAAGGGGUUUAGAGGGCUAUGCUGGUAACAGCAU AGCAAGUUUAAAUAAGGCUAGUCGGUUAUCAAACUGAAAAAGUGGCACCG AGIICGGGGGGTIIIIIIIIIIIIIIIIIIIIIIIIIIII	113
194	4x(2'OMe, 3'PACE)_CLTA1	CLTAimg ON1- target	A*GTU*C*CUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAU AGCAAGUUUAAAUAAGGGUGUCGUUAUCAACUUGAAAAAGUGGCACCG AGUCGUGCUUUUUU (SEG ID NO: 93)	113

97 98

Entry	# Guide RNA Construct	Target DNA Construct	RNA sequence (5'→3')	RNA length
195	5'- 4x(2'OMe, 3'PACE) CLTA1	CLTA1mg ON1- target	<u>A</u> * <u>G</u> * <u>U</u> * <u>C</u> *CUCAUCUCCCUCAAGGGUUUAAAGGCUAUGCUGGUAACAGCAU AGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCG	113
196	3'PACE)	CLTAlmg ON1- target	AGUCGGUGCUUUUUUU (SEQ ID NO: 93) $\underline{G*G*\underline{A}*G*U}*CCUCAUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCAAGUUUAAAUAAA$	115
197	sgRNA 5'- 5x(2'OMe, 3'PACE)_CLTA1	CLTAlmg ON1- target	CCGAGUCGGUGCUUUUUU (SEQ ID NO: 94) G*G* <u>A</u> *G*U*CCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAG CAVAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCA	115
198	sgRNA CLTA1_3'- 4x(2'OMe, 3'PACE) sgRNA	CLTA1 ON1- target	CCGAGUCGGUGCUUUUUUU (SEQ ID NO: 94) AGUCCUCAUCUCAAGCGUUUAAGAGCUAUGCGGGUAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCGGUAACCUGAAAAAGUGGCACCAGUC	113
199	C LTA 1 3'- 4x(2'OMe, 3'PACE) sgRNA	CLTA1 ON1- target	GGGGCUUU*U*U*U*U*U*U (SEQ ID NO: 95) AGUCCUCAUCUCCCUCAAGCGUUUAAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAUAAGGCUCGUUAUCAACUUGAAAAAGUGGCACCGAGUC	113
200	CLTA1_3'- 4x(2'OMe, 3'PACE) sgRNA	CLTA1mg ON1- target	GGUGCUUNTY,V,U,U,U,U,U,U,U,U,U,U,U,U,U,U,U,U,U,U,	113
201	CLTA1_3'- 4x(2'OMe, 3'PACE) sgRNA	CLTA1mg ON1- target	GGUGCUUUVAVAVAU (SEQ ID NO: 95) AGUUCACQUCCCCUCAAGCGUUAAGAGCUAGUGCUGGUAACAGCAUGGA AGUUAAAUAAGGCUAGUCCGUUAUCAAAAAGUGGCACCAAGUC	113
202	CLTA1 3'- 5x(2'OMe, 3'PACE) sgRNA	CLTA1 ON1- target	GGOCCOGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	113
203	CLTA1 3'- 5x(2'OMe, 3'PACE) sgRNA	CLTA1 ON1- target	GGUGCUU4U*U*U*U*U* (SEQ ID NO: 96) AGUCCUCAGCGCAGCGUUAAGAGCUUGUAAAGAGCAUAGCA AGUUUAAAUAAGGCCGUUAUCAAGCGUUAAAAAGUGGCACCGAGUC	113
204	CLTA1_3'- 5x(2'OMe, 3'PACE) sgRNA	CLTAlmg ON1- target	GGUGCUUAUXUXUXUXU (SEQ 1D NO: 96) AGUCUCAUCUCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAUAAGGCUAGUCGUUAUCAACUUGAAAAGUGGCACCGAGUC	113
205	CLTA1_3'- 5x(2'OMe, 3'PACE) sgRNA	CLTA1mg ON1- target	GGGGCUU4U*U*U*U*U*() (SEQ ID NO: 96) AGUCCUCAGCGCCAGCGUUAAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUAAAUAAGGCCGUCGUUAUCAAGCGCUGGAAAAGUGGCACCGAGUC	113
206	5'- 3x(2'0Me, 3'PACE)_plus1 overhg_CLTA1	CLTA1 ON1- target	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	114
207	5'- 3x(2'OMe, 3'PACE)_plus1 overhq CLTA1	CLTA1mg ON1- target	C_*A*C*UCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAU AGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCG AGUCGGUGCUUUUUU (SEQ ID No: 97)	114
208	3x(2'OMe, 3'PACE)_plus1	CLTA1mg ON1- target	C.*A.G.*UCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGUAACAGCAU AGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCG AATICCATICCHIIIIIIIIIII (CPO 11) NO. 07)	114
209	5	CLTA1 ON1- target	G.*A.*CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	114
210	5 3x(2'0Me, 3'PACE)_plus1 NC overhq CLTA1	CLTAlmg ON1- target	G.*A.G.*UCCUCAUCUCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAU AGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCG AGUCGGUGCUUUUUU (SEO ID NO: (SEO ID NO: 98)	114

TABLE 3-continued

99 100

# Guide RNA Construct	Target DNA Construct	RNA sequence (5'->3')	RNA length
5'- 3x(2'OMe, 3'PACE)_plus1	CLTA1mg ON1- target	$\underline{\mathbf{G}}_{s} \star \underline{\mathbf{A}} \star \underline{\mathbf{G}} \star \mathbf{U}$ CCUCAUCCCUCAAGCGUUUAAGACUAUGCUGGUAACAGCAU AGCAUGAAAAGUGGCACCG	114
NC_overhg_CLTA1 5'- 5x(2'0Me, 3'PACE)_plus2 overhg_ClTA1	CLTA1 ON1- target	AGUCGGUGCUUUUUU (SEQ ID NO: 98)  U.*C.XAGAU*CCUCAUCCCUCAAGCGUUUAAGAGCUAUGCUGAAGCUACGCGCGCG	115
overing_critter 5 5x(2'OMe, 3'PACE)_plus2 overthy Ct.Thl	CLTA1mg ON1- target	U.*C.*A.*G.*COCCOCCOCCOCCAAGGGGGGGGGGGGGGGAACA G.*C.*A.*G.*U.*CCUCAUCCCCCCAAGGGGGGGGGGGGGGGGGAACA GCAAAGCAAGUUAAAAAAAGGGCAAGGGGGGAACUUGAAAAAGGGGC ACCGAAGGGGGGGGGGGGGGGGGGGGG	115
5'- 5x(2'OMe, 3'PACE)_plus2	CLTA1mg ON1- target	$u_s + c_s + a * d * u * c c u c a c u c a c u c a c u c a c u c u$	115
overhg_CLTAl 5'- 5x(2'OMe, 3'PACE)_plus2	CLTA1 ON1- target	ACCARACCGGGCUUUUUU (SEQ ID NO: 99)  A, G, *A,*C*U*CCCCCCCCCCCCCCUUAAGAGCUAUGCUGGUAACA GCAUAAGAAGAGGCUAAGGCCUAGCCUUUACAAAAAGGCUAGGC ACCAAAAAAAGGCCACCAGCCUAGAAAAAGGCCAACACAAAAAAGGCCAACAAAAAAGGCCAACAA	115
5.'-Overing_ciral 5x(2'OMe, 3'PACE)_plus2 MC overho ciral	CLTA1mg ON1- target	ACCAROCCIOCOCOCO (SEE 12 NO. 100)  A, E, A, A, A, COUCAUCOCCOCCARGCGUUDAGAGCUAUGCAGACUAACACUCCCOCARGCGUUDACACUUGAAAAGUGGC GCAUAGCAAGUUVAAAUAAGGCUAGCCUAGUAUCAACUUGAAAAAGUGGC ACCAACICCAICCIIIIUIIIIIIIIIIIIIIIIIIII	115
5.'- 5x(2'OMe, 3'PACE) plus2 NC overha CTR1	CLTA1mg ON1- target	ACCAROCCIOCOCOCO (SEE 12 NO. 100)  A, Ye A & YE CUCCUCCUCCACCOUUDAGAGCUAUGCUGUAACA GCAUAGCAAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGC ACCGAAUCGAGCUAGUUUMA (SEO ID NO. 100)	115
5 7x(2'0Me, 3'PACE) plus3 overhg_CLTA1_3'- 4x(2'0Me, 3'PACE)	CLTA1 ON1- target	$\mathbb{C}_s \times \mathbb{U}_s \times \mathbb{U}_s = \mathbb{U}_s \times \mathbb$	116
5'- 7x(2'0Me, 3'PACE)_plus3 overhg_CLTAl_3'- 4x(2'0Me, 3'PACE)	CLTAlmg ON1- target	$C_s*U_s*C_s*\underline{A}*\underline{A}*\underline{G}*\underline{U}*\underline{C}*$ CUCANCCCCCCCAAGCGUUNAAGAGCUAUGCUGUA ACAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGU GCACCGAGUCGGUGCUU $\underline{U}*\underline{U}*\underline{U}*\underline{U}*$ (SEQ ID NO: 101)	116
5 7x(2'0We, 3'PACE)_plus3 overbg_CLTAl_3'- 4x(2'0We, 3'PACE)	CLTA1mg ON1- target	$C_s * U_s * C_s * \underline{A} * \underline{G} * \underline{U}_s \times \underline{C}_s * \underline{U}_s \times C_s * \underline{A} * \underline{G} * \underline{U}_s \times \underline{C}_s * \underline{U}_s \times \underline{C}_s * \underline{U}_s \times \underline{C}_s \times \underline{G} \times $	116
5:- 7x(2'0Me, 3'PACE) plus3 NC_overhg_CLTA1_3'- 4x(2'0Me, 3'PACE)	CLTA1 ON1- target	$G_a * B_a * G_a * A * G * U * C * C * U C * U C C U C C U C C U C C C C$	116
5'- 7x(2'OMe, 3'PACE) plus3 NC_overhq_CLTA1_3'- 4x(2'OMe, 3'PACE)	CLTAlmg ON1- target	G,*B,*G,*A*G*U*C*CCUCAAGCGUUUAAGAGCUAUGCUGUA ACAGCAUAGCAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAGU GGCACCGAGUCGGUGCUU⊻*U*U*U*U*U (SEQ ID NO: 101)	116
5:- 7x(2:0Me, 3:PACE) plus3 NC_overhg_CLTA1_3'- 4x(2:0Me, 3:PACE)	CLTAlmg ON1- target -	$G_a*B_a*G_a*A*G^*U*C^*CCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUUAAGAAGGCUAGUCGGUAUCAACUUCAACUUCAAAGUUGAAGUCGGUGCUUUAUGAAAGUUGGGUGCUUUUU*U*U*U*U*U*U*U*U*U*U*U*U*U*U*U*U*$	116
CLTA1_2'OMe, 3'PACE + 20	CLTA1 ON1- target	AGUCCUCAUCUCCCUCAAGC*GUUUAAGAGCUAUGCUGGUAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGUGCUUUUUUU (SEQ ID NO: 102	113

TABLE 3-continued

101 102

Entry	# Guide RNA Construct	Target DNA Construct	RNA sequence (5'→3')	RNA length
225	CLTA1_2'OMe, 3'PACE + 20 sgRNA	CLTA1mg ON1- target	AGUCCUCAUCUCCCUCAAGC*GUUUAAGAGCUAUGCUGGUAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGU	113
226	CLTA1_2'OMe, 3'PACE + 20 sgRNA_	CLTA1mg ON1- target	CGGGCCUUUUUU (SEQ ID NO: 102) AGUUUAAAUCCCUCCAAGC*GUUDAAGAGCUAUGCUGGUAACAGCAGGAGCAAGCAAGUAAAAAAGUGGCACGAGU AGUUUAAAUAAAGGCGAGCUAGCAACUUGAAAAAGUGGCACCGAGU AGGUUTIIIIIIIII (SEO ID NO: 102)	113
227	CLTA1_2'OMePACE + 19 sgRNA_	CLTA1 ON1- target	AGUCCOCCOUCO (ELS TO NO. 102) AGUCCUCAUCUCCCUCAGA*CGUUDAAGACUAUGCAGAGACAGCAGCGAGC AAGUUUAAAUAAGGCUAGCGCAGCAACUUGAAAAAGUGGCACCGAGU	113
228	CLTA1_2'OMePACE + 19 sgRNA_	CLTA1mg ON1- target	CGGGCCOUOUOU (SEQ_TD_NOS) AGUCCUCAUCUCCUCAAGA*CGUUDAAGAGCUAUGCUAGCAACAGCAAGCAAGCAAGA*CGUUDAAGAGCUGAAAAGGCGAGCACCGAGU CGGUGCUUTIIIIIIIIII (SEO_TD_NO-103)	113
229	CLTA1_2'OMePACE + 19 sgRNA	CLTA1mg ON1- target	CGGOCCOCOCO (SEQ. ID NO. 103) AGUCCUCAUCACCAGACAGCACCAGCAGCAGCAAGC AAGUUJAAAUAAGCUGAGCGCACCACCACACACACAGGCACCGAGU CGGGGCHIIIIIIIIIII (SEO ID NO. 103)	113
230	CLTA1_2'OMePACE + 19 sgRNA	CLTAlmg OFF1- target	AGUCCUCAUCUC (222 - 2010) AGUCCUCAUCUC (222 - 2010) AAGUUUAAAUAAGGCUCAGUCCUUUAAAAAAAAGUGCACCGAGU CGUIGCIIIIIIIIIII (SEO ID NO. 103)	113
231	CLTA1_2'OMePACE + 19 sgRNA	CLTAlmg OFF3- target	AGUCCUCAUCUCCUCAGA EQUIDA AGAS AAGUUUAAAUAAGGCUCAGGCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGGGCUUUUUUU (SRO ID NO: 103)	113
232	CLTA1_2'OMePACE + 18 sgRNA	CLTA1 ON1- target	AGUCCUCAUCUCCCUCAA*GCGUUUAAGAGCUAUGCUGGUAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCGUUUCAACUUGAAAAAGUGGCACCGAGU CCCUCCUITIIIIIIIII (SEO ID NO. 104)	113
233	CLTA1_2'OMePACE + 18 sgRNA	CLTAlmg ON1- target	AGUCCUCAUCUCCCUCAA, TD NO. 101) AAGUUVAAAUAAGGCUGGUAGCCACCGAGU AAGUUVAAAUAAGGCUAGUCCGUUVCAACUGAAAAAGUGGCACCGAGU CGGIIGCIIIIIIIIIIII (SFO ID NO. 104)	113
234	CLTA1_2'OMePACE + 18 sgRNA	CLTAlmg ON1- target	CGCOCCOCOCO (25% ID NO: 104) AGUCCUCAUCOCCUCAA*GCGUUDAAGAGCUAUGCUGGUAACAGCAUAGC AAGUUJAAAUAAGCUAGUCGGUUJAAACACUUGAAAAAGUGGCACCGAGU CGCUGCUIIIIIIIIII (SFO ID NO: 104)	113
235	CLTA1_2'OMePACE + 17 sgRNA	CLTA1 ON1- target	AGUCCUCAUCUC (25g 12 no. 101) AGUCCUCAUCUC (25g 12 no. 101) AAGUUUAAAUAAGCUAGUCGUUUAAAAAAAGUGCACCGAGU CGUIGCUUUUUUU (SRO ID No. 105)	113
236	CLTA1_2'OMePACE + 17 sgRNA	CLTAlmg ON1- target	AGUCCUCAUCUCCCUCA*AGCGUUANGAGAGCUAUGCUGGUAACAGCAUAGC AAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGIIGCIIIIIIIIIIII (SRO ID NO. 105)	113
237	CLTA1_2'OMePACE + 17 sgRNA	CLTA1mg ON1- target	AGUCCOCCOCC (25% 1D NO. 160) AGUCCUCAUTOCCCCCA*AGCGUUDAAGAGCAACAGCAACAGCAAAAAAAAAAAAAAA	113
238	CLTA1_2'OMePACE + 17,1 8 sgRNa	CLTA1 ON1- target	AGUCCUCAUCUC (2xxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxx	113
239	CLTA1_2'OMePACE + 17,1 8 sgRNA	CLTAlmg ON1- target	AGUCCUCAUCOCOCO (A 2 TO	113
240	CLTA1 2'OMEPACE + 17,1 8 sgRNA	CLTAimg ON1- target		113

TABLE 3-continued

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	RNA length		113	113	113	113	113	113	113	113	113	113	113	113	113	113	113
TABLE 3-continued	RNA sequence (5'→3')	3'Phosphorothioate-modified sgRNA	Assuscence de la contra del contra de la contra del la contra del la contra del la contra de la contra de la contra del la contra de la contra del la contra de la contra del la contra de la contra del	Ascausecucoussusus (seg id no: 10/) Ascausecucaucoucousageduudadagecuaugudadagecauag Caaguudaaahahagecuaguccguuaucaacuugaaahagugecaccaag	ACGGUGGUOUUBSUBUBU (SEQ 1D NO: 10/)  ASGBUSCCUCAUCACCCUCAAGCGUUUAAGAGCUAUGGUGGUAACAGCAUAG  CAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAG  TOCCUCAUTUTUTUTUTUTUT (SE)	ASGSUSCOUCUESTES (SEX ID NO: 10/) ASGSUSCOUCATUCOCCUCCAGGGUUAAGAGGGAACAGCAUAG CAAGUUAAAUAAGGGUAGUCCGUUAUCAAAAAGUGGCACCGAG	ACGGUECUUBSUSBEN (SEQ 1D NO: 10/) ASGSUSCCUCAUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAG CAAGUUUAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGAG	GCGGGGCOOU <u>DSGJGS</u> O (SEX, 1D AO) 10/7) GCGAGGUGGAGAGCUAUUGCCACAGUUUAAGAGCUAUGGAAAAAGUGGCACCGA GCAAGUUUAAAUAAGGUGGCUAGUCCGUUAAAAAAGUGGCACCGA	GUCGGGCCUU <u>USSISUS</u> U (SEQ ID NO: 107)  GSCSAGAUGUAGUGCACAGUUUACAAGUUUAAAAAAGUGGCAACGA  GCAAGUUUAAAUAAGCUAGUCCGUUAAAAAAGUGGCACCGA  GTAGUUAAAUAAAAAAAAAAAAAAAAAAAAAAAAAA	GUCGGGCCUU <u>BSISI</u> SU (SEQ ID NO: 107)  GECSAGAUGUAGUUUCCACAGUUUAAGAGCUAUGGAAACAGCAUA  GCAAGUUUAAAUAAGGCUAGUCCGUUAAAAAAGUGGCACCGA  CHACCUITUMAAUAAGCUAGUCCGUAAUAAAAAAAAAAAAAAAAAAAA	GUCGGGCCUU <u>USSISU</u> SU (SEQ ID NO: 107)  GECSAGGUGUAGUUCCACAGUUUAAGAGCUAUGGAAAAGUGGCACCGA  GCAAGUUUAAAUAAGCUAGUCCGUUAAAAAAGUGGCACCGA  GTAGGUITHIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	GCCGGGCCUU <u>DSSISU</u> SU (SEQ ID NO: 10/)  GSCSAGAUGUAGUGUCCACAGUUUAAGAGCUGGAAACAGCAUA GCAAGUUUAAAUAAGGUUCCACAGUUUAAAAAAGUGGCACCGA GTAGUUTAAUAAGTUAGCCOTO NO: 10/7)	GSCSGGSCCUUGSSGGGGGGGGGGGGGGGGGGGGGGGGG	GSCSGSCSCOORSECTOR (SEE TO NOT 127)  GSCSASGAUGUAGAUGCACAGUUUAGAGACAUGGAAACAGCAUA GCAAGUUUAAAUAAGAGCUAGUCAACAUUGAAAAAGUGGCACCGA GCAAGUUAAAUAAGAGUAGCUAGUCAACAUUGAAAAAGUGGCACCGA	GCCGGGCCC <u>GSGSGSGS</u> CGCCCCCCCCCCCCCCCCCCC	GSCSAGAGAGAGAGAGAGAGAGAGAGAGAGAAAAAAAAAA	GUCGGGCC <u>USUSUSUSU</u> GECSASCSASUGUAGUGUUUCCACAGUUUAAGAGCUGGAAACAGCAU AGCAAGUUAAAUAAGGGUAGCCGUUAUCAACUUGAAAAAGGGCACCG AGCGGGGGCU <u>SUSUSUSUSU</u> SU (SEQ ID NO: 109)
H	Target DNA Construct Construct	2'OMethyl, 3	3'-3x(2'OMe, CLTAl ON1- target	3'-3x(2'OMe, CLTAlmg ON1- target	3'-3x(2'OMe, CLTAlmg ON1- target	3'-3x(2'OMe, CLTAlmg OFF1- target	3'-3x(2'OMe, CLTAlmg OFF3- target	3'-3x(2'0Me, CLTA4 target	3'-3x(2'0Me, CLTA4 target	3'-3x(2'OMe, CLTA4mg ON- target	3'-3x(2'0Me, CLTA4mg ON- target	3'-3x(2'OMe, CLTA4mg ON- target	CLTA4_5', 3'-3x(2'OMe, CLTA4mg OFF5- 3'P(S)) target	CLTA4_5'-3x(2'OMe, CLTA4mg ON- 3'P(S)), 3'-5x(2'OMe, target	CLTA4mg ON- CLTA4 5'-3x(2'OMe, CLTA4mg ON- (S)), 3'-5x(2'OMe, target	CLTA4_5'-3x(2'OMe, CLTA4mg ON- 3'P(S)), 3'-5x(2'OMe, target	) 5', 3'-5x(2'OMe, CLTA4mg OM- ) target
	Entry # Guide RNA		241 CLTA1_5', 3'P(S))	242 CLTA1 5', 3'P(S))	243 CLTA1_5', 3'P(S))	244 CLTA1_5', 3'P(S))	245 CLTA1_5', 3'P(S))	246 CLTA4_5', 3'P(S))	247 CLTA4_5', 3 3'P(S))	248 CLTA4_5', 3'P(S))	249 CLTA4_5', 3'P(S))	250 CLTA4_5', 3'P(S))	251 CLTA4_5', 3'P(S))	252 CLTA4_5'-3 3'P(S)), 3	253 CLTA4_5'-3 3'P(S)), 3	254 CLTA4_5'-3 3'P(S)), 3	3.F(S)) 255 CLTA4 5', 3'P(S))

Entry :	# Guide RNA Construct	Target DNA Construct	RNA sequence (5' $\rightarrow$ 3')	RNA length
256	CLTA4_5', 3'-5x(2'OMe, 3'P(S))	CLTA4mg ON- target	<u>GsCsasgsaguduuccacaguuuaagagcuaggegaaacagcau</u> Agcaaguuuaaauaaggcuaguccguuaucaacuugaaaaaguggcaccg Agiicciigciigciiiaiisiisiisiisii (SPO ID NO. 100)	113
257	CLTA4_5', 3'-5x(2'OMe, 3'P(S))	CLTA4mg OFF5- target	GSCSASGSAS GOURGE OF THE TABLE OF THE STATE	113
		2'OMethyl, 3'	3'PhosphorothioPACE-modified sgRNA	
258	CLTA1_5', 3'-3x(2'OMe, 3'thioPACE)	CLTA1 ON1- target	A*sG*su*sccucaucuccucaageguuuaagagagagagagagagagagagagagagaga	113
259	CLTA1_5', 3'-3x(2'OMe, 3'thioPACE)	CLTA1mg ON1- target	GAGUCGGUGCUQUESE, SU (SEQ ID NO: 110)  A*sG*su*sccuchucuccagagauunaagagcudugagagagagagagagagagagagagagagagagagag	113
260	CLTA1_5', 3'-3x(2'OMe, 3'thioPACE)	CLTAimg ON1- target	A*sG*sU*sccucaucuccuccaucaucuunaaaacuudaaacuudaaacuudaaaaaadugccu Uagcaaguuuaaauaaggcuaguccuuuaucaacuudaaaaagugccacc Gagucgguccuuuu*su*su*su*su*su*su*su*su*su*su*su*su	113
261	CLTA1_5', 3'-3x(2'OMe, 3'thioPACE)	CLTAlmg OFF1- target	A*sG*su*sccucaucuccuccacguunaagacunugaagacuaugcacaca Uagcaaguuunaauaagacuaguccguuaucaacuugaaaaaguggcacc gagucggucguuut*su*su*su*su (SEO ID NO: 110)	113
262	CLTA1_5', 3'-3x(2'OMe, 3'thiopacE)	CLTAlmg OFF3- target	A*sG*su*scucaucaucaccucaaccuuuaaaacuaucuacuacaca Uagcaaguuuaauaaggcuaguccguuaucaacuugaaaaggcacc Gagucgguccuuuu*su*su*su*su*su*su*su*su*su*su*su*su	113
263	CLTA1_5', 3'-1x(2'0Me, 3'thioPACE)	CLTAimg ON1- target	A*sGUCCUCAUCCCCCCAAGCGUUDAAGAGCUGGUAACAGCAUAG CAAGUUUAAUAAGCUAGUCCGUUAAAAGUUGAAAAGUGGCACCGAG UCGGUGCUUUUUUU*SU CRO TD NO: 111)	113
264	CLTA1_5', 3'-1x(2'OMe, 3'thioPACE)	CLTAIMG ON1- target	A*succucaucuccuchagcguuhaaqagcuhuggcuqguhacagcaung Caaguuuhaanaaggcuhguucaacuugaaaaaguggcaccgag Ucggugcuuuuut*su (SEO ID NO: 111)	113
265	CLTAL_5', 3'-1x(2'OMe, 3'thiopace)	CLTAlmg OFF1- target	A*SGUCCUCAUCÜCCCUCAAĞCGUUDAAQAĞCUAUGGUAACAĞCAUAĞ CAAĞUUDAADAAQAĞCUAĞUCĞUUAUCAACUUĞAAAAĞUĞĞCACCĞAĞ UCĞĞUĞCUUUUU*SU (SEQ ID NO: 111)	113
266	CLTAL_5', 3'-1x(2'OMe, 3'thiopace)	CLTAlmg OFF3- target	A*SGUCCUCAUCUCCCUCAAGCGUUDAAGAGCUAUGCUGGUAACAGCAUAG CAAGUUDAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAG UCGGUGCUUUUUV*SU (SEO 1D NO: 111)	113
267	CLTA1_5', 3'-3x(2'OMe, 3'thiopace) 75 mer	CLTA1 ON1- target	A*sG*su*sccucaucruccucaagegunuaagacuaugeugguacagea	75
268	CLTA1_5', 3'-1x(2'0Me, 3'+hiopack) 74 mer	CLTAIMG ON1-	A*sGUCCUCAUCUCCCUCAAGCGUUDAAGAGCUAUGCUGGUAACAGCAUAG	74
269	CLTA1_5', 3'-1x(2'OMe, 3'thioPACE) 75 mer	CLTAIMG ON1- target	A**SGUCCUCAAGCGUUAAAGGCUAGCCGGAAGCCAAGCGAAGGCAAGGAAGCAAGGAAGCAAGGAAGAA	75
270	CLTA1_5', 3'-1x(2'OMe, 3'thioPACE) _77 mer	CLTAimg ON1- target	A*sGUCCUCAUCUCCCUCAAGCGUŪUAAGAGCUAUGCUGGUAACAGCAUAG CAAGUUUAAAUAAGGCUAGUCGUUAU*sC (SEQ ID NO: 113)	77
271	CLTA1_5', 3'-1x(2'OMe, $3$ 'thioPACE) 77 mer + G	CLTA1mg ON1- target	G*sAGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUA GCAAGUUUAAAUAAGGCUAGUCCGUUAU*sC (SEO ID NO: 114)	78
272	121	CLTA4 ON- target	G*sC*sA*scauguaguguuuccacaguuuaagagcuaugcaaacagca Uagcaaguuuaauaaggcuaguccguuaucaacuugaaaaaguggcacc	113

RNA →3') length	GGCUACCCACAGUUDAGAGCUAUGCAAAAGCAGCA GGCUAGUCCGUUAUCAACUGAAAAAGUGGCACC T*** CFO ID NO. 11E)	UGUUCCCACAGUUDAGAGCUAUGGAAAAGCA GGCUAGUCCGUUDAAAAAAGUGGCACC GGCOGUUDACACUUGAAAAAGUGGCACC ###### (SFO ID NO: 115)	UGUUCCACACAUVAAGAGCUUGAAAAAGGGAACAGCA GGCUUGUCCGUUAUCAACUUGAAAAAGUGGCACC U*8If*sU (SRO ID No: 115)	UCCACAGUUDAGAGCUACACACAGCAUAG UCCACAGUUDAGAGCUAGACACACAGAG UAGUCCGUUDAUCACUUGAAAAGUGGCACCGAG (SEO ID NO: 116)	UCCACAGUUDAGAGACUAUGGAAACAGCAUAG UAGUCCGUUAUCAACUUGAAAAGUGGCACCGAG (SRO ID NO: 116)	UCCACAGUUDAAGAGCUAUGGAAACAGCAUAG UAGUCCGUUDAAGAUGAAAAGUGGCACCGAG (SEQ ID NO: 116)	UCCACAGUUUAAGAGCUAUGCUGGAAACAGCAUAG UAGUCCGUUAUCAACUUGAAAAGUGGCACCGAG (SEQ ID NO: 116)	COLLIGE COLLEGES!		AAGUGUUVAAGAGCUAUGCUGGVAACAGCAUAGC AGUCCGUVAUCAACUUGAAAAGUGGCACCGAGU EQ ID No: 117)	2aA) AAGUGUUUAAGAGCUAUGCUGGUAACAGCAU GCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCG (SEO ID NO: 118)	2aA) AAGUGUUUAAGAGCUAUGCUGGUAACAGCAU GCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCG (SEQ ID NO: 118)	UGAAGUUUAAGAGCUAUGCUAACAGCAUAGC AGUCCGUUAUCAACUUGAAAAGUGGCACCGAGU EO ID NO: 119)	UGAAGUUUAAGAGCUAUGCUAACAGCAUAGC AGUCCGUUAUCAACUUGAAAAGUGGCACCGAGU EO ID NO: 119)	aA) UUGAAGUUUAAGAGCUAUGCUGGUAACAGCAU GCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCG (SEQ ID NO: 120)	aA) UUGAAGUUUAAGAGCUAUGGUAACAGCAU GCUAGUCGUUAUCAACUUGAAAAAGUGGCACCG (SEQ ID NO: 120)
TABLE 3-continued  Target DNA  Entry # Guide RNA Construct  Construct  RNA sequence (5'->3')	273 CLTA4_5', 3'-3x(2'OMe, CLTA4 ON- G*sC*sA*sGAUGUAGUGUUCCACCAGUUDAAGAGCUAUGCUGGAAACACACA 3'thioPACE) target target target cannot cann	274 CLTA4_5', 3'-3x(2'OMe, CLTA4 ON- G*SC*COUGUS-SU*SU*SU*SU*SU*SU*SU*SU*SU*SU*SU*SU*SU*S	275 CLTA4_5', 3'-3x(2'OMe, CLTA4mg OFF5- G*s2*sA*sGAUGUUCCACAGUUUAAAAGAGCUAUCCACAGUUAAAAAGAGCUAGAAAAAGGCACC UAGCAAAAAGGCACC UAACAACUUAAAAAAGGCAACC GAGUCAGUUAAAAAGGCAACUUAAAAAAAAAA	276 CLTA4_5', 3'-1x(2'OMe, CLTA4mg ON- G*sCAGAUGUGAGGGUGUUCCAGGUUUAAGAGGGAAACAGCAAGGAAACAGAAACAAACAAAA	277 CLTA4_5', 3'-1x(2'OMe, CLTA4mg ON- G*sCAGAUGUAGUGUUCCACAGUUUAAGAGGGGGGGGGAACAGGAACAGGAAGAGGAACAGGAAGAGGAACAGGAAGAGGAAGAGGAAGAGGAAGAGGAAGAGGAAGAGGAAGAG	278 CLTA4_5', 3'-1x(2'OMe, CLTA4mg ON- G*sCAGAUGUAGUGUUUCCACAGUUAAGAGGGAACAGCAGAACAGAACAGCAAGAACAGCAAGAACAGCAAGAACAGCAAGAACAGCAAGAACAGCAAGAACAGCAAGAACAGCAAGAAAAAA	279 CLTA4_S', 3'-1x(2'OMe, CLTA4mg OFF5- G*SCAGAUGUAGAGUGUCCACAGUUUAAGAGCUAUGCUGGAAACAGCAUAG 3'thioPACE) target CAAGUUUAAAUAAGGCUAGUCCGUUAUCAACAGUUAAAAAAAGUGGCACCGAG UCGGUGCUUUUU¥sU (SEQ ID NO: 116)	Z-aminoa-modified	target	281 ENI ENI ENIMING OFF- GAUGUUGUCGAUGAAAAAGUGUUAAGCUAUUAAAAAGAGUUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGGCACCGAGU AAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAAGUGCCACCGAGU CGGUGCUUUUUUU (SEQ ID NO: 117)	282 EN1_2aminoA + 16 EN1mg ON- GAUGUUGUCGAUGAA(2aA)AAGUGUUUAAGAGCUAGUCGUAACAGCAU AGCAAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAGUGGCACCG AGUCGGUGCUUUUUUU (SEO ID NO: 118)	283 ENI_2aminoA + 16 ENImg OFF- GAUGUUGUCGAUGACGAUGAA(2aA)AAGUGUUUAAAGAGCUAUUGAAAAGUGGUAACAGCAU AGCAAGUUDAAUAAGGCUAGUCGUUAUCAACUUGAAAAGUGGCACCG AGUCGGUGCUUUUUU (SEQ ID NO: 118)	284 PCDHA4 PCDHA4mg ON-GAUUUAGACGAUUGAAGGUUUAAGACGUUUAAGACGUUUAAAGACUUUAAAGACUUUAAAAAGUGGCACCGAGU AAGUUUAAAAAAGUCGUUAUCAAAAAGUGGCACCGAGU CGGUGCUUUUUUU (SEO ID NO: 119)	285 PCDHA4 PCDHA4mg OFF- GAUUUAGACGAAGGAUUGAAGGUUUAAGCGUUAAGCGUUAAGCGUUAAGCGUUAUCAACUUGAAAAGUGGCACCGAGU AAGUUUAAAUAAGGCUAGUCGUUAUCAACUUGAAAAAGUGGCACCGAGU CGGUGCUUUUUUU (SEO ID NO: 119)	PCDHA4mg ON- target	287 PCDHA4_2aminoA + 15 PCDHA4mg OFF- GAUUUAGACGAAGG(2aA)UUGAAGUUUAAGAGCUUAAGAGCUGAACAGCAGAAGAGAGAAGAGAAGAGAAGAAAAAAGAGAAAAAA

109

110

	RNA length		113	113		113	113		100	113
TABLE 3-continued	RNA sequence $(5' \rightarrow 3')$	5-methylU-modified sgRNA	GCAGA (5mU) G (5mU) G (5mU) G (5mU) (5mU) (5mU) CCACAGUUUAAAGAGC (5mU) A (5mU) GC (5mU) GG (5mU) AACAGCA (5mU) AGCAGCA (5mU) AGCAGCA (5mU) GGCACCGUUUAUCAAC (5mU) (5mU) GAAAAAG (5mU) GGCACCGAGUCGG	(5mU) GC (5mU) (5mU) (5mU) (5mU) (5mU) (5mU) U (SEQ ID NO: 121)  GCAGA (5mU) G (5mU) AG (5mU) G (5mU) (5mU) (5mU) CCACAGUUUAAGAGC (5mU) A (5mU) GC (5mU) GG (5mU) AACAGCA (5mU) AGCAAGUUUAAAUAAGG CUAGUCCGUUAUCAAC (5mU) (5mU) GAAAAAG (5mU) GGCACCGAGUCGG (5mU) GC (5mU) (5mU) (5mU) (5mU) (5mU) U (5mU) U (5mU) U (5mU) U (5mU) U (5mU) U	Z base-modified sgRNA	AGUCCUCAUCUCCCUCAAGCGUUUAAGAGCUAUGCUGGUAACAGCAUAGCA AGUUUAAUAAUAAGGCUAGUZZGUUAUCAACUUGAAAAAGUGGCACCGAGUC	AGUCCUCAUCUCCOCCAGAGUDAAGAGUDUGCUGGUDAACAGCAUAGCA AGUUUAAAUAAGGCUAGUCCGUUAACAACUUGAAAAAGUGGCA <b>ZZ</b> GAGUC GGUGCUUUUUUU (SEQ ID NO: 122)	sgRNA modified to disfavor misfolding	A*sGUCCUCAUCUCCCUCAAGGGUUDAAGAGCUAGUAAUAGCAAGUUUAAA UAAGGUUAAUCCGUUAUCAACAAGAAAUUGUGGCACCGAGUCGGUGCUU <u>U</u>	*8U (SEQ ID NO: 123)  **ASGUCCUCAUCUCCCCAAGCGUUDAAGAGCUAUGCUAACAACAUAG CAAGUUDAAAUAAGG <u>U</u> UAAUCCGUUAUCAACAAGAAAUUGUGGCACCGAG UCGGUGCUUUU <u>U</u> **SÜ (SEQ ID NO: 124)
	Target DNA Construct		CLTA4mg ON- target	CLTA4mg OFF5- target		CLTA1 ON1- target	CLTA1 ON1- target	bs.	CLTAlmg ON1- target	CLTAlmg ON1- target
	Entry # Guide RNA Construct		CLTA4_21x(5-MeU)	CLTA4_21x(5-MeU)		290 CLTA1_22_70,71	CLTA1 22_95,96		CLTAl opti_short_5', 3'- 1x(2'OMe,	3.thloPACE, 2.OME 54,5/ CLTAL opti short 5', 3'- 1x(2'OME, 2'OME 64,67
	Entry #		7 8 8 8	289		290	291		292	293

N = 2'00We
Ns = 3'P(3)
N\* = 3'-D(3)
N\* = 3'-D(3)
N\* = 3'-D(3)
N\* = 2'0Me, 3'-DACE
(2sU) = 2'-DACE
(2sU) = 2'-DACE
(5sU) = 5'-DACE
(5sU)

 $Z\,=\,Z$  base IntFl = Fluorophore incorporated at an internal position in the RNA sequence

111 112

The DNA target constructs in Table 3 had the following sequences:

CLTA1 ON1target: AGAATTTAACTGTGGTCACATTTGCTTTATCGACTGCTTCATCTCACAGCTCATC
TTACGCAAGTTCGATCAGTTACCAGTCACTTTCAATTTGGTTGAATGTTCCGTTG
ACATGCGAGTTCTCTTCGACCATGTGCCCGCGGATTGAATTCCTCAAGCAGGTGGTGATA
GATGCTACGGTGGTGATGCGCTCAGTCCTCATCTCCTCAAGCAGGCCCC
GCTGGTGGGTCGGAGTCCCTAGTGAAGCCACCAATATAGTGGTCGTGTCAAGCAAC
TGTCCACGCTCCACCCTCGAGGTCGTAACATAAACGTACTAAGCACCAGTAAACA
AGATCGATAGCAAGAACATGGTATAGACTGACGAGAGCTCGCCATTAGTCTGA
(SEO ID NO: 10)

CLTA1 OFF1target: CLTA1 OFF2target: AGAATTTAACTGTGGTCACATTTGCTTTATCGACTGGCTTCATCTACCAGCTCATC
TTACGCAAGTTCGATGAGTATGCCAGTCACTTTCAATTTGATTTCATTTCATTTGATGTTCCCGTG
ACATGCGAGTTCTGTCGACCATGTGCCGCGGATTGAATTCCTCAAGGGGTGGTGATA
GATGCTACGGTGGTGATGCAATAAATTTCAGCCCTCATTTCCCTCAAGCAGGGGTT
ACTTTAGGGTCGGAGTCCCTAGTGAAGCCACCAATATAGTGGTCGTGTCAAGCAAC
TGTCCACGCTCCACCCTCGAGGTCGTAACATAAACGTACTAAGGCACCAGTAAACA
AGATCGATAGCAAGAACATGGTATAGACTGACGGAGAGCTCGCCATTAGTCTGA
(SEQ ID NO: 12)

CLTA1 OFF3target: AGAATTTAACTGTGGTCACATTTGCTTTATCGACTGGCTTCATCTCACAGCTCATC
TTACGCAAGTTCGATGAGTATGCCAGTCACTTTCAATTTGGTTGAATGTTCCGTTG
ACATGCGAGTTCTGTCGACCATGTGCCAGGTTGAATTCCTCAAGCGGTGGTGATA
GATGCTACGGTGGTGATGCCTCCACCCCTCATCCCCCTCAAGCCGGTCCC
AGGCTGGGGTCGGAGTCCCTAGTGAAGCCACCAATATAGTGGTCGTGTCAAGCAAC
TGTCCACGCTCCACCCTCGAGGTCGTAACATAAACGTACTAAGCACCAGTAAACA
AGATCGATAGCAAGAACAATGGTATAGACTGACGAGGAGCTCGCCATTAGTCTGA
(SEQ ID NO: 13)

CLTA1mg ON1target:  $\tt GCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGG$ GCGACACGGAAATGTTGAATACTCATACTCTTCCTTTTTCAATATTATTGAAGCAT TTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATA  ${\tt AACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTAAATTGTAAGCG}$  ${\tt TTAATATTTTGTTAAAATTCGCGTTAAATTTTTGTTAAATCAGCTCATTTTTTAAC}$ CAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAAGAATAGACCGAGATAGG  $\tt GTTGAGTGTTCCAGTTTGGAACAAGAGTCCACTATTAAAGAACGTGGACTCCA$ ACGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCA  $\tt CCCTAATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTAAATCGGAACCCTAA$ AGGGAGCCCCCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTGGCGAGAAAGG  ${\tt AAGGGAAGAAAGCGAAAGGAGCGGCGCTAGGGCGCTGGCAAGTGTAGCGGTCACG}$  $\tt CTGCGCGTAACCACCACCACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCA$ TTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGC  ${\tt TATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACG}$ CCAGGGTTTTCCCAGTCACGACGTTGTAAAACGACGCCAGTGAGCGCGCGTAATA CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCGCAGGCCAAAGATG  $\texttt{TCTCCCGCATGCGCTCAGTCCTCATCTCCCTCAAGC} \underline{\texttt{AGG}} \underline{\texttt{CCCTGCTGGTGCACTGA}}$ AGAGCCACCCTGTGCGCGTGATATGCAGCTCCAGCTTTTGTTCCCTTTAGTGAGGG TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTA TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAA GGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCC CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC  ${\tt GACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGC}$  $\tt TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG$  $\tt CTGGGCTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA$ CTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCA  $\tt CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG$  $\tt TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT$  $\tt CCGCTGGTAGCGGTGGTTTTTTTTTTTTGCAAGCAGCAGATTACGCGCAGAAAAAAA$  $\tt GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA$  ${\tt AAACTCACGTTAAGGGATTTTGGTCATGAGATTNTCAAAAAGGATCTTCACCTAGA}$ TCCTTTTAAATTAAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACT TGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCT ATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGG  $\tt AGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCG$ 

114

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-continued

GCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGG
TCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAAGCTAGAG
TAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCTACAGGCATC
GTGGTGTCCACGCTCGTTGTTTGGTATGGCTTCATTCAGCTCCGGTTCCCTACCGATC
AAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAAGCGGTTAGCTCCTTCCGTC
CTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCA
GCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGG
TGAGTACTCAAACCAAGTCATTCTGAGAAATAGTGTATGCGGCGACCGAGTTGCTCTT
GCCCGGCGTCAATACGGGATAATACCGCGCCACATAGC
(SEO ID NO: 14)

CLTA1mg OFF1- G

target:

GCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAAATGCCGCAAAAAAAGGGAATAAGG  $\tt GCGACACGGAAATGTTGAATACTCATACTCTTTCCTTTTTCAATATTATTGAAGCAT$ TTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATA  $\verb|AACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTAAATTGTAAGCG|$  ${\tt TTAATATTTTGTTAAAATTCGCGTTAAATTTTTGTTAAATCAGCTCATTTTTTAAC}$ CAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAAGAATAGACCGAGATAGG  $\tt GTTGAGTGTTGCCAGTTTGGAACAAGAGTCCACTATTAAAGAACGTGGACTCCA$ ACGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCA  $\tt CCCTAATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTAAATCGGAACCCTAA$  $\tt AGGGAGCCCCCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTGGCGAGAAAGG$  $\tt CTGCGCGTAACCACCACACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCA$  $\tt TTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGC$ TATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACG  $\tt CCAGGGTTTTCCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCGTAATA$ CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCGCAGGGCAAAGAGG TCTCCTGTATGCACTCAGTCCTCAACTCCCTCAAGCAGGCGACCCTTGGTGCACTG ACAAACCGCTCCTGCGCGTGATATGCAGCTCCAGCTTTTGTTCCCTTTAGTGAGGG TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTTCCTGTGTGAAATTGTTA TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAA GGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCC CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC GACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGC TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG CTGGGCTGTGCACGAACCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA CTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCA  $\tt CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG$  $\tt TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT$  $\tt CCGCTGGTAGCGGTGGTTTTTTTTTTTGCAAGCAGCAGATTACGCGCAGAAAAAAA$  $\tt GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA$  $\verb|AAACTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGA||$ TCCTTTTAAATTAAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACT  $\tt TGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCT$  ${\tt ATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGG}$ AGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCG GCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGG  ${\tt TCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAG}$ TAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCATC  $\tt GTGGTGTCACGCTCGTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATC$  ${\tt AAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTC}$  $\tt CTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCA$  $\tt GCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGG$  $\tt TGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTT$ GCCCGGCGTCAATACGGGATAATACCGCGCCACATAGC (SEO ID NO: 15)

CLTA1mg\_OFF3 target:

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CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCAGGAGAGGGAGCCA AAAGCTTGTCTTTGCGCGTGATATGCAGCTCCAGCTTTTGTTCCCTTTAGTGAGGG  $\tt TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTA$  ${\tt TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG}$  $\tt GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC$  $\tt CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG$  ${\tt AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT}$  $\tt CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA$ TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAAGGCCAGCAAAA CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA  $\tt CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC$ TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG  $\tt CTGGGCTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA$  $\tt CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG$  $\tt TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT$  $\tt CCGCTGGTAGCGGTGGTTTTTTTTTTTTTGCAAGCAGCAGATTACGCGCAGAAAAAAA$  $\tt GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA$  ${\tt AAACTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGA}$ TCCTTTTAAATTAAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACT TGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCT ATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGG AGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCG GCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGG TCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAG TAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCATC GTGGTGTCACGCTCGTCTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATC AAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTC CTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCA GCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGG TGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTT GCCCGGCGTCAATACGGGATAATACCGCGCCACATAGC (SEO ID NO: 16)

CLTA4 ONtarget: GCCGGGCAAGAGCAACTCGGTCGCCGCATACACTATTCTCAGAATGACTTGGTTGA GTACTCACCAGTCACAGAAAAGCATCTTACGGATGGCATGACAGTAAGAGAATTAT ATCGGAGGACCGAAGGAGCTAACCGCTTTTTTGCACAACATGGGGGATCATGTAAC TCGCCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATACCAAACGACGAGCGTG  $\tt CTGGAGCCGGTGAGCGTGGGTCTCGCGGTATCATTGCAGCACTGGGGCCAGATGGT$  ${\tt AAGCCCTCCCGTATCGTAGTTATCTACACGACGGGGAGTCAGGCAACTATGGATGA}$  ${\tt ACGAAATAGACAGATCGCTGAGATAGGTGCCTCACTGATTAAGCATTGGTAACTGT}$  ${\tt CAGACCAAGTTTACTCATATATACTTTAGATTGATTTAAAACTTCATTTTTAATTT}$ AAAAGGATCTAGGTGAAGATCCTTTTTGATAATCTCATGACCAAAATCCCTTAACG  ${\tt TGAGTTTTCGTTCCACtGaGCGTCAGACCCCGTAGAAAGATCAAAGGATCTTCTT}$  $\tt CCAGCGGTGGTTTGTTTGCCGGATCAAGAGCTACCAACTCTTTTTCCGAAGGTAAC$ GCCACCACTTCAAGAACTCTGTAGCACCGCCACATACCTCGCTCTGCTAATCCTGT TACCAGTGGCTGCTTGCCAGTGGCGATAAGTCGTGTCTTACCGGGTTGGACTCAAG AGCCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCTACAGCGTGAGCTA  $\tt TGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGG$  $\tt TTTATAGTCCTGTCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTTGTGATGC$ TCGTCAGGGGGGGGGGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTTACGGTT CCTGGCCTTTTGCTGGCCTTTTGCTCACATGTTCTTTCCTGCGTTATCCCCTGATT CTGTGGATAACCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGCAGCCGA ACGACCGAGCGCAGCGAGTCAGTGAGCGAGGAAGCGGCAAGAGCGCCCAATACGCAA ACCGCCTCTCCCCGCGCGTTGGCCGATTCATTAATGCAGCTGGCACGACAGGTTTC  ${\tt TAGGCACCCCAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGGAATTGT}$ GAGCGGATAACAATTTCACACAGGAAACAGCTATGACCATGATTACGCCAAGCGCG CAATTAACCCTCACTAAAGGGAACAAAAGCTGGGTACCGGGCCCCCCTCGACACC  ${\tt AGTTGCATTCGATTCCTGTTTGTAATTGTCCAATTCCTGCAGCCCGGGGGATCGGC}$  ${\tt AGATGTAGTGTTTCCACA} \underline{\tt GGG} \underline{\tt GATCCACTAGTTCTAGAGCGGCCGCCACCGCGGTG}$  ${\tt GAGCTCCAATTCGCCCTATAGTGAGTCGTATTACGCGCGCTCACTGGCCGTCGTTT}$ TACAACGTCGTGACTGGGAAAACCCTGGCGTTACCCAACTTAATCGCCTTGCAGCA CATCCCCTTTCGCCAGCTGGCGTAATAGCGAAGAGGCCCGCACCGATCGCCCTTC  $\tt CCAACAGTTGCGCAGCCTGAATGGCGAATGGAAATTGTAAGCGTTAATATTTTGTT$ AAAATTCGCGTTAAATTTTTGTTAAATCAGCTCATTTTTTAACCAATAGGCCGAAA TCGGCAAAATCCCTTATAAATCAAAAGAATAGACCGAGATAGGGTTGAGTGTTGTT

117

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CLTA4mg ONtarget:  $\tt GCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGG$ GCGACACGGAAATGTTGAATACTCATACTCTTCCTTTTTCAATATTATTGAAGCAT  $\tt TTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATA$  $\verb|AACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTAAATTGTAAGCG|$  ${\tt TTAATATTTTGTTAAAATTCGCGTTAAATTTTTGTTAAATCAGCTCATTTTTTAAC}$ CAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAAGAATAGACCGAGATAGG  $\tt GTTGAGTGTTCCAGTTTGGAACAAGAGTCCACTATTAAAGAACGTGGACTCCA$ ACGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCA  $\tt CCCTAATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTAAATCGGAACCCTAA$ AGGGAGCCCCCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTGGCGAGAAAGG  ${\tt AAGGGAAGAAAGCGAAAGGAGCGGCGCTAGGGCGCTGGCAAGTGTAGCGGTCACG}$ CTGCGCGTAACCACCACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCA TTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGC TATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACG CCAGGGTTTTCCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCGTAATA CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCAAGAGCTTCACTGA GTAGGATTAAGATATTGCAGATGTAGTGTTTCCACAGGGTGGCTCTTCAGTGCACC AGCGGAACCTGCTGCGCGTGATATGCAGCTCCAGCTTTTGTTCCCTTTAGTGAGGG TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTA TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAA GGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCCC CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA  $\tt CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC$  ${\tt GACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGC}$  $\tt TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG$  $\tt CTGGGCTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA$  $\tt CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG$  $\tt TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT$  $\tt CCGCTGGTAGCGGTGGTTTTTTTTTTTTGCAAGCAGCAGATTACGCGCAGAAAAAAA$ GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA AAACTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGA TCCTTTTAAATTAAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACT  $\tt TGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCT$ ATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGG  $\tt AGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCG$  $\tt GCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGG$  ${\tt TAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCATC}$  $\tt GTGGTGTCACGCTCGTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATC$  ${\tt AAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTC}$  $\tt CTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCA$  $\tt GCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGG$ TGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTT GCCCGGCGTCAATACGGGATAATACCGCGCCACATAGC (SEO ID NO: 18)

CLTA4mg OFF5target:

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CTGCGCGTAACCACCACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCA TTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGC TATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACG  $\tt CCAGGGTTTTCCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCGTAATA$  $\tt CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCTCTGAATAGAGTTG$  $\tt GGAAGAGATGCATACAACATATGTAGTATTTCCACA\underline{GGG}AATACAATGGACAAATG$  ${\tt ACCTCAAGAGCAGGCGCGTGATATGCAGCTCCAGCTTTTGTTCCCTTTAGTGAGGG}$  ${\tt TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTTCCTGTGTGAAATTGTTA}$  ${\tt TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG}$  $\tt GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC$  ${\tt CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG}$ AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT  $\tt CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA$ TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAA GGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCCC  $\tt CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA$  $\tt CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC$  $\tt TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG$  $\tt CTGGGCTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA$  $\tt CTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCA$  $\tt CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG$  $\tt TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT$ CCGCTGGTAGCGGTGGTTTTTTTTTTTTCCAAGCAGCAGATTACGCGCAGAAAAAAA GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA AAACTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGA TCCTTTTAAATTAAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACT TGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCT ATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGG AGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCG GCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGG TCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAG TAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCATC GTGGTGTCACGCTCGTCTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATC AAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTC CTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCA GCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGG TGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTT GCCCGGCGTCAATACGGGATAATACCGCGCCACATAGC (SEQ ID NO: 19)

IL2RGmg\_ON target:  $\tt GCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAAATGCCGCAAAAAAAGGGAATAAGG$  $\tt GCGACACGGAAATGTTGAATACTCATACTCTTTCCTTTTTCAATATTATTGAAGCAT$  $\tt TTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATA$  $\verb|AACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTAAATTGTAAGCG|$  $\tt TTAATATTTTGTTAAAATTCGCGTTAAATTTTTGTTAAATCAGCTCATTTTTTAAC$  ${\tt CAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAAGAATAGACCGAGATAGG}$  $\tt GTTGAGTGTTGCCAGTTTGGAACAAGAGTCCACTATTAAAGAACGTGGACTCCA$  $\verb|ACGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCA||$  $\tt CCCTAATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTAAATCGGAACCCTAA$ AGGGAGCCCCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTGGCGAGAAAGG CTGCGCGTAACCACCACACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCA  $\tt TTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGC$ TATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACG  $\verb|CCAGGGTTTTCCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCGTAATA||$  $\tt CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCGGGCAGCTGCAGGA$  $\tt ATAAGAGGGATGTGAATGGTAATGATGGCTTCAACA\underline{TGG}CGCTTGCTCTTCATTCC$  $\tt CTGGGTGTAGTCTGCGCGTGATATGCAGCTCCAGCTTTTGTTCCCTTTAGTGAGGG$  ${\tt TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTA}$  ${\tt TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG}$  $\tt GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC$  $\tt CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG$  $\tt AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT$ CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAA  $\tt CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA$  $\tt CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC$ GACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGC TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG  $\tt CTGGGCTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA$  $\tt CTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCA$ CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT CCGCTGGTAGCGGTGGTTTTTTTTTTTTTCCAAGCAGCAGATTACGCGCAGAAAAAAA  $\tt GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA$ 

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EN1mg\_ON target:

GCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAAATGCCGCAAAAAAAGGGAATAAGG  $\tt GCGACACGGAAATGTTGAATACTCATACTCTTTCCTTTTTCAATATTATTGAAGCAT$  $\tt TTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATA$  ${\tt AACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTAAATTGTAAGCG}$ TTAATATTTTGTTAAAATTCGCGTTAAATTTTTGTTAAATCAGCTCATTTTTTAAC  ${\tt CAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAAGAATAGACCGAGATAGG}$  $\tt GTTGAGTGTTCCAGTTTGGAACAAGAGTCCACTATTAAAGAACGTGGACTCCA$ ACGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCA  $\tt CCCTAATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTAAATCGGAACCCTAA$  $\tt AGGGAGCCCCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTGGCGAGAAAGG$ AAGGGAAGAAGCGAAAGGAGCGGGCGCTAGGGCGCTGGCAAGTGTAGCGGTCACG  $\tt CTGCGCGTAACCACCACACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCA$ TTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGC TATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACG CCAGGGTTTTCCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCGTAATA CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCCTCCTTACTGCAGC  $\tt CGAAGTCCGGCCTCAGGATGTTGTCGATGAAAAAGT\underline{TGG}TGGTGCGGTGCAGCTGG$ GCCGCTGCCTGCGCGCGCTGATATGCAGCTCCAGCTTTTGTTCCCTTTAGTGAGGG TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTA TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAA GGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCCC CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA  $\tt CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC$  ${\tt GACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGC}$  $\tt TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG$  $\tt CTGGGCTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA$  $\tt CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG$  $\tt TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT$  $\tt CCGCTGGTAGCGGTGGTTTTTTTTTTTGCAAGCAGCAGATTACGCGCAGAAAAAAA$  $\tt GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA$ AAACTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGA TCCTTTTAAATTAAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACT TGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCT  $\tt AGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCG$  $\tt GCTCCAGATTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGG$  ${\tt TCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAG}$ TAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCATC  $\tt GTGGTGTCACGCTCGTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATC$  ${\tt AAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTC}$  $\tt CTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCA$  $\tt GCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGG$ TGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTT GCCCGGCGTCAATACGGGATAATACCGCGCCACATAGC (SEQ ID NO: 21)

EN1mg\_OFF target: GCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAAATGCCGCAAAAAAAGGGAATAAGG
GCGACACGGAAATGTTGAATACTCATACTCTTCTTTTTCAATATTATTGAAGCAT
TTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATA
AACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTAAATTGTAAGCG
TTAATATTTTGTTAAAATTCGCGTTAAAATTTTTGTTAAATCAGCTCATTTTTTAAC
CAATAGGCCGAAATCGGCAAAATCCCTTATAAAATCAAAAGAATAGACCGAGATAGG
GTTGAGTGTTTCCAGGTTTGGAACAAGAGATCACCATATTAAAGAACGTGGACTCCA
ACGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCA
CCCTAATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTAAATCGGAACCCTAA
AGGGAGCCCCCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTGGCCGAGAAAGG

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AAGGGAAGAAGCGAAAGGAGCGGCGCTAGGGCGCTGGCAAGTGTAGCGGTCACG CTGCGCGTAACCACCACACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCA TTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGC  ${\tt TATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACG}$  $\tt CCAGGGTTTTCCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCGTAATA$  $\tt CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCGTCCTTTCGCCGGC$  $\tt CGAACTCGGGCCGCAGGATGTTGTCGATGAAGAAGT\underline{TGG}TGATGCGGTGCGGGTGC$  $\tt TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTA$ TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG  $\tt GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC$ CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG  ${\tt AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT}$ CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA  ${\tt TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAA}$  $\tt CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA$  $\tt CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC$  $\tt TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG$  $\tt CTGGGCTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA$  $\tt CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG$  $\tt TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT$ CCGCTGGTAGCGGTGGTTTTTTTTTTTTCCAAGCAGCAGATTACGCGCAGAAAAAAA GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA AAACTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGA TCCTTTTAAATTAAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACT TGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCT ATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGG AGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCG GCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGG TCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAG TAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCATC GTGGTGTCACGCTCGTCTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATC AAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTC CTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCA GCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGG TGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTT GCCCGGCGTCAATACGGGATAATACCGCGCCACATAGC (SEQ ID NO: 22)

PCDHA4mg\_ON target:  $\tt GCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGG$ GCGACACGGAAATGTTGAATACTCATACTCTTCCTTTTTCAATATTATTGAAGCAT  $\tt TTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATA$  ${\tt AACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTAAATTGTAAGCG}$  ${\tt TTAATATTTTGTTAAAATTCGCGTTAAATTTTTGTTAAATCAGCTCATTTTTTAAC}$  ${\tt CAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAAGAATAGACCGAGATAGG}$  $\tt GTTGAGTGTTCCAGTTTGGAACAAGAGTCCACTATTAAAGAACGTGGACTCCA$  ${\tt ACGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCA}$  $\verb|CCCTAATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTAAATCGGAACCCTAA||$  $\tt AGGGAGCCCCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTGGCGAGAAAGG$ AAGGGAAGAAGCGAAAGGAGCGGGCGCTAGGGCGCTGGCAAGTGTAGCGGTCACG  $\tt CTGCGCGTAACCACCACCACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCA$  $\tt TTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGC$  ${\tt TATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACG}$  $\tt CCAGGGTTTTCCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCGTAATA$  $\tt CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCGGAACATTGGTAAT$  ${\tt TAAACTTAACGCCTCAGATTTAGACGAAGGATTGAA\underline{TGG}GGACATTGTTTATTCAT$  ${\tt TCTCGAATGATACGCGCGTGATATGCAGCTCCAGCTTTTGTTCCCTTTAGTGAGGG}$  $\tt TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTA$ TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG  $\tt GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC$  ${\tt CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG}$ AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA  ${\tt TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAA}$ CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC GACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGC  $\tt TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG$  $\tt CTGGGCTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA$ CTATCGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCA  $\tt CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG$  $\tt TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT$ CCGCTGGTAGCGGTGGTTTTTTTTTTTTCCAAGCAGCAGATTACGCGCAGAAAAAAA

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-continued

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GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA AAACTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGA TCCTTTTAAATTAAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACT  $\tt TGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCT$  ${\tt ATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGG}$ AGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCG  $\tt GCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGG$ TCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAG  ${\tt TAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCATC}$ GTGGTGTCACGCTCGTTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATC  ${\tt AAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTC}$ CTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCA GCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGG TGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTT GCCCGCCTCAATACGGGATAATACCGCCCCACATAGC (SEQ ID NO: 23)

PCDHA4mg\_ OFFtEut  $\tt GCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGG$ GCGACACGGAAATGTTGAATACTCATACTCTTTCCTTTTTCAATATTATTGAAGCAT  $\tt TTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATA$  $\verb|AACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTGCCACCTAAATTGTAAGCG|$  ${\tt TTAATATTTTGTTAAAATTCGCGTTAAATTTTTGTTAAATCAGCTCATTTTTTAAC}$ CAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAAGAATAGACCGAGATAGG GTTGAGTGTTGCCAGTTTGGAACAAGAGTCCACTATTAAAGAACGTGGACTCCA ACGTCAAAGGGCGAAAAACCGTCTATCAGGGCGATGGCCCACTACGTGAACCATCA  $\tt CCCTAATCAAGTTTTTTGGGGTCGAGGTGCCGTAAAGCACTAAATCGGAACCCTAA$ AGGGAGCCCCGATTTAGAGCTTGACGGGGAAAGCCGGCGAACGTGGCGAGAAAGG AAGGGAAGAAAGCGAAAGGAGCGGGCGCTAGGGCGCTGGCAAGTGTAGCGGTCACG CTGCGCGTAACCACCACACCCGCCGCGCTTAATGCGCCGCTACAGGGCGCGTCCCA  $\mathsf{TTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGC$ TATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACG CCAGGGTTTTCCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCGTAATA CGACTCACTATAGGGCGAATTGGGTACGATCGATGCGGCCTCGGAACGCTGGTGAT TCATCCCAATGCCTCAGATTTAGACGAAGGCTTGAATGGGGATATTATTTACTCCT TCTCCAGTGATGTGCGCGTGATATGCAGCTCCAGCTTTTGTTCCCTTTAGTGAGGG TTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTA TCCGCTCACAATTCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGG GTGCCTAATGAGTGAGCTAACTCACATTAATTGCGTTGCGCTCACTGCCCGCTTTC CAGTCGGGAAACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGAG AGGCGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTGCGCT CGGTCGTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTA TCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAA GGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTCCGCCCC  $\tt CTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGA$ CTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCC  ${\tt GACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGC}$  $\tt TTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG$  $\tt CTGGGCTGTGCACGAACCCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAA$  $\tt CTGGTAACAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAG$  ${\tt TGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT}$  $\tt CCGCTGGTAGCGGTGGTTTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAAAAAAA$ GGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGA  $\verb|AAACTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGA||$ TCCTTTTAAATTAAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACT  $\tt TGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCT$  $\tt ATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGG$  ${\tt AGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCG}$  $\tt GCTCCAGATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTGG$ TCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCTAGAG  ${\tt TAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCATC}$ GTGGTGTCACGCTCGTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATC  ${\tt AAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAAGCGGTTAGCTCCTTCGGTC}$  $\tt CTCCGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCA$ GCACTGCATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGG TGAGTACTCAACCAAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTT CCCGGCGTCAATACGGGATAATACCGCGCCACATAGC (SEQ ID NO: 24)

In a 20-uL reaction volume, 50 fmoles of linearized DNA target in the presence of 50 nM sgRNA, 39 nM recombinant purified Cas9 protein (*S. pyogenes*; Agilent) and 10 mM or 0.8 mM MgCl<sub>2</sub> at pH 7.6 was incubated at 37° C. for 30 min. Upon completion, 0.5 uL of RNace It (Agilent) was added, and incubation was continued at 37° C. for 5 min and then at 70° C. for 15 min. Subsequently 0.5 μL of Proteinase K

(Mol. Bio. grade, NEB) was added and incubated at 37° C. for 15 min. Aliquots were loaded into a DNA 1000 or DNA 7500 LabChip and were analyzed on a Bioanalyzer 2200, or alternatively were loaded into a Genomic DNA ScreenTape and were analyzed on a TapeStation. The workup steps served to release Cas9 from binding of target DNA, which was assayed for cleavage. Cleavage yields were calculated

 $[Mg^{2+}]$ 

(mM)

tcDNA

Entry

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by the formula: a/(a+b)×100 where a is the sum of the band intensities of the two cleavage products and b is the remaining uncleaved DNA if present. A cleavage percentage of 100% means that all of the target DNA construct was cleaved

A series of guide RNAs were chemically synthesized. The guide RNA oligomers were synthesized on an ABI 394 Synthesizer (Life Technologies, Carlsbad, Calif., USA) using 2'-O-thionocarbamate-protected nucleoside phosphoramidites according to procedures described in Dellinger 10 et al. (2011) J. Am. Chem. Soc., 133, 11540-56. 2'-O-methyl phosphoramidites were incorporated into RNA oligomers under the same conditions as the 2'-O-thionocarbamate protected phosphoramidites. The 2'-O-methyl-3-O-(diisopropylamino)phosphinoacetic acid-1,1-dimethylcyanoethyl 15 ester-5'-O-dimethoxytrityl nucleosides used for synthesis of thiophosphonoacetate (thioPACE)-modified RNAs were synthesized essentially according to published methods. See Dellinger et al. (2003) 1 Am. Chem. Soc., 125, 940-50; and Threlfall et al. (2012) Org. Biomol. Chem., 10, 746-54. For 20 phosphorothioate-containing oligomers, the iodine oxidation step after the coupling reaction was replaced by a sulfurization step using a 0.05 M solution of 3-((N,Ndimethylaminomethylidene)amino)-3H-1,2,4-dithiazole-5thione in a pyridine-acetonitrile (3:2) mixture for 6 min.

All the oligonucleotides were purified using reversed-phase high-performance liquid chromatography (HPLC) and analyzed by liquid chromatography-mass spectrometry (LC-MS) using an Agilent 1290 Infinity series LC system coupled to an Agilent 6520 Q-TOF (time-of-flight) mass spectrometer (Agilent Technologies, Santa Clara, Calif., USA). The yields for the synthesis and purification of the sgRNAs were estimated using deconvolution of mass spectra obtained from LC-MS-derived total ion chromatograms. The chemical synthesis of the 100-mer sgRNAs typically yielded 25-35% full-length product from a nominal 1 micromole scale synthesis. Reversed-phase HPLC purification using ion pairing buffer conditions typically gave 20% yield from the crude product with an estimated purity of the final sgRNA in the range of 90% to 95%.

The results are shown in Table 4. "% Target cleaved" indicates the percentage of the target DNA construct which was cleaved. Experiments were run with and without addition of a molar excess of targetless competitor DNA (tcDNA) which potentially competes with the target DNA, so the potential impact of the added nonspecific DNA upon the assay could be seen.

TABLE 4

Entry #	[Mg <sup>2+</sup> ] (mM)	tcDNA	% Target cleaved	% CV	% Cleaved vs. CONTROL	% CV CONTROL
			piece dual-g fied dual-gui		scaffold NA (dgRNA)	
1 2 3 4 5 6 7 8 9	0.8 0.8 0.8 0.8 0.8 0.8 0.8 0.8	N Y N Y N Y N Y	99% 99% 96% 100% 96% 0% 99% 100% 94%, 93% 88% orophore-coi	_	J. DNIA	
11	10	N	•	_	94%, 93%	
12	0.8	Y	87%	_	88%	

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TABLE 4-continued

% Cleaved vs.

CONTROL

% CV

CONTROL

% Target %

cleaved CV

_	Ħ	(IIIIVI)	ICDINA			CONTROL	CONTROL
5		2'C	Methyl,	3'Phosphorotl	nioate-	modified dgRNA	
-				•			
	13	0.8	Y	87%	_	88%	_
_		2'ON	/lethyl,3	Phosphorothi	oPACE	E-modified dgRNA	1
	14	0.8	Y	89%		88%	_
10	15	0.8	Y	86%		88%	
10				2-thioU-modi	fied d		
-							
	16	0.8	N	96%	_	99%	_
	17	0.8	Y	95%	5%	99%	5%
	18 19	0.8 0.8	N Y	95% 100%	5%	96% 100%	<u>-</u> 5%
15	20	0.8	N	97%	<i>J 70</i>	96%	
	21	0.8	Y	0%	5%	0%	5%
	22	0.8	N	98%	_	99%	_
	23	0.8	Y	99%	5%	100%	5%
	24	0.8	N	94%	_	99%	_
20	25	0.8	Y	83%	5%	99%	5%
	26 27	0.8 0.8	N Y	93% 94%	5%	96% 100%	5%
	28	0.8	N	90%	<i>J70</i>	96%	
	29	0.8	Y	0%	5%	0%	5%
	30	0.8	N	95%	_	99%	_
	31	0.8	Y	94%	5%	100%	5%
25	32	0.8	N	92%		99%	_
	33	0.8	Y	84%	5%	99%	5%
	34 35	0.8 0.8	N Y	90% 94%	5%	96% 100%	5%
	36	0.8	N	70%		96%	<i>370</i>
	37	0.8	Y	0%	5%	0%	5%
30	38	0.8	N	96%	_	99%	_
	39	0.8	Y	59%	5%	100%	5%
				Single-guid			
-			Unmod	ined single-gi	mae K	NA (sgRNA)	
	40	10	N	93%	_		
25	41	10	N	94%	_		
35	42	10	N	94%	_		
	43	10	N	92%	_		
	44	10	N	90%, 92%	_		
	45 46	10 10	N N	92% 93%	_		
	47	0.8	N	86%			
40	48	0.8	N	87%	_		
	49	0.8	Y	87%	_		
	50	0.8	N	82%	_		
	51	0.8	N	92%	_		
	52	10	N	60%	_		
45	53 54	0.8 0.8	N N	90% 90%			
10	55	0.8	Y	79%			
	56	0.8	Ň	79%	_		
	57	0.8	N	94%	_		
	58	10	N	73%	_		
	59	0.8	N	84%	_		
50	60	0.8	Y	≥85% 89%	_		
	61 62	0.8 0.8	Y N	87%, 82%			
	63	0.8	N	23%, 22%	_		
	64	0.8	N	78%	_	87%	_
	65	0.8	Y	76%	_	87%	_
55	66	0.8	N	65%	_	87%	_
	67	0.8	N	81%	_	87%	_
	68 69	0.8 0.8	N Y	85% 71%	_	87% 87%	_
	70	0.8	N	32%	_	87%	_
	71	0.8	N	84%	_	87%	_
	72	0.8	N	91%	_	87%	_
<b>CO</b>	73	0.8	Y	79%	_	87%	_
60		0.8	N	88%	_	87%	_
60	74			93%	_	87%	
60	74 75	0.8	N N			970/	
60	74 75 76	0.8 0.8	N	87%	_	87% 87%	_
60	74 75 76 77	0.8 0.8 0.8	N Y	87% 79%	_	87%	_
60	74 75 76 77 78	0.8 0.8 0.8 0.8	N Y N	87% 79% 89%		87% 87%	_
	74 75 76 77	0.8 0.8 0.8	N Y	87% 79%		87%	_

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TABLE 4-continued

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TABLE 4-continued

		-	TIDEL T	• • • • • • • • • • • • • • • • • • • •	maca					-	TIDEE T	• • • • • • • • • • • • • • • • • • • •		
Entry #	[Mg <sup>2+</sup> ] (mM)	tcDNA	% Target cleaved	% CV	% Cleaved vs. CONTROL	% CV CONTROL		Entry #	[Mg <sup>2+</sup> ] (mM)	tcDNA	% Target cleaved	% CV	% Cleaved vs. CONTROL	% CV CONTROL
81	0.8	N	5%	_	86%	_	. 5	150	0.8	N	5%		87%	_
82	0.8	N	89%		86%	_	,	151	10	N	0%, 0%		90%, 92%	_
83	0.8	N	68%		87%			152	10	N	0%, 0%		90%, 92%	
84	0.8	Y	50%		87%	_			0.8				86%	_
						_		153		N	85%			
85	0.8	N	69%	_	87%	_		154	0.8	N	87%	_	86%	_
86	0.8	N	69%	_	87%	_		155	0.8	N	89%	_	87%	_
87	0.8	N	76%	_	87%	_	10	156	0.8	Y	78%	_	87%	_
88	0.8	Y	42%	_	87%	_		157	0.8	N	84%	_	87%	_
89	0.8	N	72%	_	87%	_		158	0.8	N	93%	_	87%	_
90	0.8	N	78%	_	87%	_		159	0.8	N	90%	_	86%	_
91	0.8	N	85%	_	87%	_		160	0.8	N	90%	_	87%	_
92	0.8	Y	51%	_	87%	_		161	0.8	Y	86%	_	87%	_
93	0.8	N	82%	_	87%	_	15	162	0.8	N	90%	_	87%	_
94	0.8		83%	_	87%	_		163	0.8	N	91%	_	87%	_
			DMT-modif	ied sg	RNA			164	0.8	N	92%	_	90%	_
							•	165	0.8	N	89%	_	87%	_
95	10	N	93%	_	92%	_		166	0.8	Y	80%	_	87%	_
96	10	N	93%	_	92%	_		167	0.8	N	90%	_	87%	_
		Flu	orophore-mo	odified	l sgRNA			168	0.8	N	94%	_	87%	_
			•				20	169	0.8	N	90%	_	84%	_
97	10	N	91%, 91%	_	90%, 92%	_		170	0.8	Y	≥85%	_	≥85%	_
98	0.8	N	86%	_	87%	_		171	0.8	N	7%	_	84%	_
99	0.8	Y	77%	_	87%	_		172	0.8	Y	0%	_	≥85%	_
100	0.8	N	87%		87%	_		173	10	Ň	15%	_	73%	_
101	0.8	N	86%		87%	_		174	0.8	N	85%		84%	_
102	0.8	N	91%		87%	_	25	175	0.8	Y	75%		≥85%	_
102	0.8	Y	82%		87%	_	20	176	10	N N	86%		73%	_
103	0.8	N	90%		87%	_		177	0.8	11	0%		84%	_
						_								_
105	0.8	N	92%	_	87%	_		178	0.8	Y	0%	_	≥85%	_
106	0.8	N	91%		87%	_		179	10	N	15%		73%	_
107	0.8	Y	82%	_	87%	_				2	Deoxy-mod	пеа я	gKNA	
108	0.8	N	90%	_	87%	_	30							
109	0.8	N	91%	_	87%	_		180	10	N	27%, 19%	_	90%, 92%	_
110	0.8	N	92%	_	87%	_		181	10	N	0%, 0%	_	90%, 92%	_
111	0.8	Y	84%	_	87%	_		182	10	N	0%, 0%	_	90%, 92%	_
112	0.8	N	92%	_	87%	_				2'Deo	xy,3'PACE-	modif.	ied sgRNA	
113	0.8	N	89%	_	87%	_								
114	0.8	N	84%, 84%	_	87%, 82%	_	35	183	0.8	N	72%, 77%	_	87%, 82%	_
115	0.8	N	12%, 6%	_	23%, 22%	_	-	184	0.8	N	8%, 9%	_	23%, 22%	_
116	0.8	N	93%, 90%	_	87%, 82%	_				2'OMe	thyl,3'PACE	-modi	fied sgRNA	
117	0.8	N	8%, 9%	_	23%, 22%	_								
		3'Phos	phorothioate	-modi	fied sgRNA			185	0.8	N	82%	_	87%	_
							•	186	0.8	Y	72%	_	87%	_
118	10	N	95%	_	90%, 92%	_		187	10	Y	95%	_	93%	_
119	10	N	94%	_	90%, 92%	_	40	188	10	Y	95%	_	94%	_
120	10	N	97%	_	90%, 92%	_		189	0.8	Y	91%	_	87%	_
121	10	N	94%	_	90%, 92%	_		190	0.8	Y	84%	_	87%	_
		2'0	OMethyl-mo	dified				191	0.8	Y	85%	_	87%	_
							•	192	0.8	Y	77%	_	87%	_
122	10	N	91%	_	94%	_		193	10	Ŷ	88%	_	94%	_
123	10	N	92%		93%		45	194	0.8	Ŷ	70%	_	87%	
124	0.8	N	86%		87%	_		195	0.8	Ŷ	56%	_	87%	_
125	"	Y	77%		87%	_		196	0.8	Y	40%		87%	_
126		N	85%		87%	_		197	0.8	Y	23%		87%	
120	0.8	N	88%		87%	_		198	10	Y	88%		93%	_
				_		_								_
128	10	N	92%	_	94%	_		199	10	Y	89%	_	94%	_
129	0.8	N	83%	_	87%	_	50	200	0.8	Y	84%	_	87%	_
130	0.8	Y	78%	_	87%	_		201	0.8	Y	75%	_	87%	_
131	0.8	N	83%	_	87%	_		202	10	Y	90%	_	93%	_
132	0.8	N	85%	_	87%	_		203	10	Y	90%	_	94%	_
133	10	N	92%	_	94%	_		204	0.8	Y	86%	_	87%	_
134	0.8	N	86%	_	87%	_		205	0.8	Y	82%	_	87%	_
135	0.8	Y	78%	_	87%	_	55	206	10	Y	88%	_	93%	_
136	0.8	N	83%	_	87%	_	33	207	0.8	Y	82%	_	87%	_
137	0.8	N	88%	_	87%	_		208	0.8	Y	78%	_	87%	_
138	10	N	91%	_	94%	_		209	10	Ÿ	77%	_	93%	_
139	0.8	N	84%	_	87%	_		210	0.8	Ÿ	71%	_	87%	_
140	0.8	Y	81%		87%	_		211	"	Ÿ	69%	_	"	_
141	0.8	N	83%		87%	_		211	10	N	80%	_	93%	_
141	0.8	N	87%	_	87%	_	60	212	0.8	N N	56%		93% 87%	_
142	10		87% 89%		87% 92%	_		213	0.8	Y Y	30% 41%		8/%	_
		N		_		_						_		_
144	0.8	N	91%, 88%		87%, 82%	_		215	10	Y	78%	_	93%	_
145	0.8	N	24%, 25%		23%, 22%	_		216	0.8	Y	58%	_	87%	_
146	10	N	93%, 92%		90%, 92%	_		217	0.8	Y	44%	_	"	_
147	0.8	N	22%	_	87%	_		218	10	Y	80%	_	93%	_
148	0.8	Y	3%	_	87%	_	65	219	0.8	Y	39%	_	87%	_
149	0.8	N	12%	_	87%	_		220	0.8	Y	13%	_		_

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TABLE 4-continued

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TABLE 4-continued

	Entry #	[Mg <sup>2+</sup> ] (mM)	tcDNA			% Cleaved vs. CONTROL	% CV CONTROL
5			Z	base-modi	fied s	gRNA	
	290	10	N	19%	_	92%	_
	291	10	N	93%	_	11	_
			sgRNA n	nodified to	disfav	or misfolding	
10	292	0.8	N	93%	_	90%	_
	293	0.8	N	93%	_	86%	_
							_

The results revealed that guide RNAs containing modifications at specific positions were tolerated by active Cas protein and gRNA:Cas protein complexes, as the modifications did not prevent target-specific cleavage of the on-target polynucleotide. The modifications that were tested and found to be tolerated at specific positions include 2'-O-methylribonucleotide (=2'OMe), 2'-deoxyribonucleotide, racemic phosphorothioate internucleotide linkage, 3'-phosphonoacetate (=PACE), 3'-thiophosphonoacetate (=thioPACE), Z nucleotide, 2-thiouracil, 2-aminoadenine, 5-methyluracil, 5-aminoallyluracil coupled to Cy5 fluorophore, 2-(4-butylamidofluorescein)propane-1,3-diol bis (phosphodiester) linker, and combinations of these.

It is contemplated that the chemical modifications disclosed and tested herein, particularly at the tested positions (as listed in Tables 3 and 4), will be tolerated at equivalent 30 positions in a variety of guide RNAs.

As disclosed herein, chemically modified nucleotides were incorporated into guide RNAs in an effort to improve certain properties. Such properties include improved nuclease resistance of the guide RNA (also known as improved stability), reduced off-target effects of a gRNA:Cas protein complex (also known as improved specificity), improved efficacy of gRNA:Cas protein complex when cleaving, nicking or binding a target polynucleotide, improved transfection efficiency, and/or improved organelle localization.

The assay results in Tables 3 and 4 indicate that: (1) In guide RNAs, many positions can tolerate a variety of chemical modifications; (2) 5' and 3' ends of guide RNAs will tolerate a wide variety of end-protecting modifications, and such modifications are useful to inhibit exonucleolytic 45 RNA degradation; (3) 2-ThioU can be used to deter offtarget interactions involving G-U wobble pairings, thereby increasing the specificity of guide pairing by inhibiting off-target hybridization interactions; (4) 5' Extensions are generally well-tolerated; (5) Surface exposed regions of the 50 guide RNA (as inferred from published crystal structures) are tolerant of extensively modifying U's to 5-methylU's, which potentially make the modified RNA more likely to elude immune responses such as stimulated by unmodified RNA; and (6) For RNA folding, G-C pairs are stronger and more stable than A-U pairs. At least one guide RNA is tolerant of replacing some G-C pairs with 2'-O-methylA-2'-O-methylU pairs that are more stable thermodynamically than unmodified A-U pairs.

More particularly, the present example shows that 2'-O-methyl modifications are tolerated at the 5' and 3' ends of dual-guide RNAs (as shown by entry 12 in Tables 3 and 4) and single-guide RNAs (entries 143-146, 169-170), thus allowing end-protection to stabilize gRNAs against exonucleases. 2'-O-methyl modifications are tolerated at most but not all internal positions, thus allowing stabilization against various nucleases including endonucleases (entries 146, 153-168, 174-179). However, the present example also

		1	ABLE 4-	cont	inuea	
Entry #	[Mg <sup>2+</sup> ] (mM)	tcDNA	% Target cleaved	% CV	% Cleaved vs. CONTROL	% CV CONTROL
221	10	Y	74%	_	93%	_
222	0.8	Y	36%	_	87%	_
223	0.8	Ÿ	19%	_	87%	_
224	10	Ý	86%	_	93%	_
225	0.8	Y	84%		87%	
226	0.8	Ý	80%		0770	
227	10	Y	88%		93%	_
228	0.8	Y	83%		93% 87%	_
229	0.8	Y	82%		87%	_
					87%	_
230	0.8	N N	80%	_	87% 87%	_
231 232	0.8		84% 88%		93%	_
232	10 0.8	N N			93% 87%	_
	0.8		85%			_
234		Y	73%		87%	_
235	10	Y	82%	_	93%	_
236	0.8	Y	89%	_	87%	_
237	0.8	Y	76%	_	87%	_
238	10	Y	65%	_	93%	_
239	0.8	Y	84%	_	87%	
240	0.8	Y	56%	_	87%	
	2'C	Methyl,3	'Phosphoroth	iioate	-modified sgRNA	1
241	10	N	92%	_	92%	
242	0.8	N	84%	_	87%	_
243	0.8	Y	88%	_	87%	_
244	0.8	N	85%	_	87%	_
245	0.8	N	91%	_	87%	
246	0.8	N	91%	_	84%	
247	0.8	Y	≥85%		≥85%	_
248	0.8	N	84%		84%	_
249	0.8	Y	90%		89%	_
250	0.8	N	90%, 87%		87%, 82%	
				_		
251	0.8	N	16%, 19%		23%, 22%	_
252	0.8	N	93%	_	89%	
253	0.8	N	90%, 90%	_	87%, 82%	_
254	0.8	N	17%, 22%		23%, 22%	_
255	0.8	N	93%		89%	_
256	0.8	N	91%, 91%	_	87%, 82% 23%, 22%	_
257	0.8 2'ON	N Aethyl,3'P	13%, 16% hosphorothic	PAC	E-modified sgRN	JA
258	10	N	89%	_	92%	_
259	0.8	N	84%	_	87%	_
260	0.8	Y	80%	_	87%	_
261	0.8	N	77%	_	87%	_
262	0.8	N	83%	_	87%	_
263	0.8	N	92%	_	87%	
264	0.8	Y	79%	_	87%	
265	0.8	N	88%	_	87%	_
266	0.8	N	94%	_	87%	_
267	10	N	74%	_	93%	_
268	0.8	N	11%	_	86%	_
269	0.8	N	15%	_	0	_
270	0.8	N	49%	_	0	_
271	0.8	N	31%	_	0	_
272	0.8	N	91%	_	84%	_
273	0.8	Y	77%	_	≥85%	_
274	0.8	N	90%, 91%	_	87%, 82%	_
275	0.8	N	9%, 8%	_	23%, 22%	_
276	0.8	N	90%	_	84%	_
277	0.8	Y	≥85%	_	≥85%	_
278	0.8	N	86%, 88%	_	87%, 82%	_
279	0.8	N	11%, 7%	_	23%, 22%	_
				cludi	ng unmodified co	ontrols)
200			000/ 000:		<u></u>	
280	0.8	Y	88%, 88%	_		
281	0.8	Y	76%, 75%	_		
282	0.8	Y	87%, 91%	_	88%, 88%	_
283	0.8	Y	90%, 90%	_	76%, 75%	_
284	0.8	Y	85%, 87%	_		
285	0.8	Y	88%, 88%	_		
286	0.8	Y	93%, 96%	_	85%, 87%	_
287	0.8	Y	82%, 79%	1:6- 1	88%, 88%	_
		5-r	nethylU-moo	ппес	sgKNA	
200	n •	N	860/ 920/		870% 820%	
288	0.8	N	86%, 83%	_	87%, 82%	_
289	0.8	N	11%, 11%	_	23%, 22%	_

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demonstrates that not every position in guide RNAs will tolerate 2'-O-methyls (as shown by entries 151-152 and 171-173), suggesting that too many consecutive 2'-O-methyl modifications at the 5' end (e.g., 26 or more consecutive 2'-O-methyl-modified nucleotides), or too many 2'-O-5 methyl modifications of C and U nucleotides downstream (3') of the 5'-terminal 20mer guide sequence is not well tolerated (e.g., the inhibitory effect of one or both 2'-O-methyluracils at sequence positions +56 and +69 in entries 171-173 as revealed by the positions tested in entries 154- 10 156).

The present example shows that 2'-O-methyl modifications throughout the 20mer guide sequence are tolerated during in vitro uses in buffer containing 10 mM Mg<sup>2+</sup> (entry 146), but such extensive modification is not well tolerated 15 under physiological conditions (entries 147-150) as present in cells. Thus, in some embodiments, a gRNA comprising 15 or more, alternatively 17 or more, alternatively 18 or more, alternatively 20 2'-O-methyl modifications throughout the 20mer guide sequence is used for in vitro methods as 20 described herein, such as genomic editing to modify a DNA sequence in vitro, regulating the expression of a gene of interest in vitro, cleaving a DNA target sequence in vitro, and other uses.

The present example shows that extensive incorporation 25 of 2'-deoxy modifications is not well tolerated and can be substantially completely inhibitory (entries 180-182). However, 2'-deoxy modifications can be well-tolerated at some locations (entry 183), therefore such modification can be useful for inhibiting nucleases.

The present example also shows that fluorophore or dye labels are tolerated in every loop of the three known stemloops in CRISPR-Cas9 guide RNAs (entry 116). Such labels are also tolerated in a 5' overhang on the guide sequence (entry 114), tolerated at additional locations in sgRNAs 35 (entry 114), and tolerated in a loop in tracrRNA used in dual-guide applications (entry 11). In this example, two different types of fluorophores were tested: a phosphodiester-linked fluorophore (no ribose ring) that essentially takes the place of a nucleotide (entries 114 & 116), and a dye label 40 (Cy5) covalently coupled to 5-aminoallyIU incorporated in a guide RNA (entry 11).

The present example also shows that Z bases are tolerated in synthetic guide RNAs, particularly as modifications of synthetic guide RNAs in which some C's are replaced with 45 Z bases (entries 290-291). The present example also shows that several other bases are tolerated at various positions, as shown in Tables 3 and 4.

The present example further shows that the 5' and 3' ends of guide RNAs can tolerate a wide variety of end-protecting 50 modifications. Such modifications can be used to inhibit exonucleolytic RNA degradation. Support for the tolerance of such modifications can be found in Hendel et al., *Nat. Biotechnol.* (2015) 33:9, 985-9. Additional support for modifications at the 5' and 3' ends of guide RNAs is provided 55 by entries 143-144, 185-223, 241-257, 258-266, and 272-279 in Tables 3 and 4. In some embodiments, the guide RNA comprises 7 or fewer modified nucleotides at a 5' end or a 3' end or at each of 5' and 3' ends, alternatively 6 or fewer, alternatively 5 or fewer, alternatively 4 or fewer, alternatively 1. Dual-guide RNAs can be protected similarly (entries 12-15).

The present example further shows that 2-thioU can be used to deter off-target interactions involving G-U wobble pairings, thereby increasing the specificity of guide 65 sequence pairing by inhibiting off-target hybridization interactions (entries 16-39). One of the base pairs involved in

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hybridization between the guide RNA and CLTA1 OFF-target 3 (also referred to as "CLTA1 OFF3-target" or "CLTA1 OFF3") is a G-U wobble pair. Replacing the corresponding U in the guide RNA with a 2-thioU reduces cleavage from 100% (entry 8) to 59% (entry 39). Replacing other U's with 2-thioU's (e.g., at sequence position +3 or +9, entries 23 and 31) does not have the same effect, presumably because those U's do not involve G-U wobble pairing when fully hybridized to each of the OFF-target sites tested. Accordingly, 2-thioU can increase target specificity of guide RNAs when off-target sites involve G-U wobble pairing.

The present example also shows that 5'-overhang sequences attached to the guide sequence are generally well-tolerated (see entries 83-95, 114, and 206-223). For example, a bulky dimethoxytrityl (dmt) group at the 5' end was well tolerated (entry 95). The chromatographic properties of dmt can be used to facilitate purification of full-length synthetic RNAs from incompletely elongated byproducts which are generally produced during synthesis. Accordingly, in some embodiments, the synthetic guide RNA comprises a 5'-overhang sequence, for example, comprising a short polynucleotide sequence of 15 or fewer nucleotides which is complementary to the guide sequence and is covalently linked at its 3' end to the 5' end of the guide sequence by a polymeric linker such as a polynucleotide or similarphosphodiester-based linker, in which the linker can be 5 or more consecutive uridine nucleotides, alternatively 6 or 7.

The present example also shows that surface exposed regions of the guide RNA (as inferred from crystal structures published by others) are tolerant of extensively modifying uracils nucleotides to 5-methyluracils (5-methylU's) (entry 288), which can make the modified RNA more likely to elude immune responses such as stimulated by unmodified RNA. In particular, the 5' and 3' ends of a synthetic guide RNA are potentially immunostimulatory, and the present example shows that 5' and 3' ends are tolerant of 5-methylU modifications (entry 288).

The present example also shows that a synthetic guide RNA is tolerant of replacing some G-C pairs with 2'-O-methylA-2'-O-methylU pairs which are more stable thermodynamically than unmodified A-U pairs (see the non-terminal-2'-O-methylU and complementary-2'-O-methylA modifications in entries 292-293). This is advantageous because it is known that, for folded RNAs, G-C pairs are stronger and more stable than A-U pairs. Replacement of G-C pairs with such thermostabilized A-U pairs in synthetic guide RNAs allows for improved folding of active structures by preventing misfolded structures that involve unintended G-C pair(s), as can be predicted by RNA folding algorithms in common use.

#### EXEMPLARY EMBODIMENTS

Exemplary embodiments provided in accordance with the presently disclosed subject matter include, but are not limited to, the claims and the following embodiments:

A1. A synthetic guide RNA comprising:

- a crRNA segment comprising (i) a guide sequence capable of hybridizing to a target sequence, (ii) a stem sequence; and
- a tracrRNA segment comprising a nucleotide sequence that is partially or completely complementary to the stem sequence, wherein the synthetic guide RNA comprises at least one modified nucleotide, and wherein the synthetic guide RNA has gRNA functionality.

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- A2. The synthetic guide RNA of embodiment A1, comprising a 2'-deoxy moiety.
- A3. The synthetic guide RNA of embodiment A1 or A2, comprising a 2'-halo moiety selected from 2'-fluoro, 2'-chloro, 2'-bromo and 2'-iodo.
- A4. The synthetic guide RNA of any one of the preceding embodiments, comprising a phosphorothicate group.
- A5. The synthetic guide RNA of any one of the preceding embodiments, comprising a PACE group.
- A6. The synthetic guide RNA of any one of the preceding embodiments, comprising a thioPACE group.
- A7. The synthetic guide RNA of any one of embodiments A2-A6, comprising a 2'-O-methyl moiety.
- A8. The synthetic guide RNA of any one of the preceding 15 embodiments, comprising a 2-thiouracil.
- A9. The synthetic guide RNA of any one of the preceding embodiments, comprising a 4-thiouracil.
- A10. The synthetic guide RNA of any one of the preceding embodiments, comprising a 2-aminoadenine.
- A11. The synthetic guide RNA of any one of the preceding embodiments, comprising a hypoxanthine.
- A12. The synthetic guide RNA of any one of the preceding embodiments, comprising a 5-methylcytosine.
- A13. The synthetic guide RNA of any one of the preced- 25 ing embodiments, comprising a 5-methyluracil.
- A14. The synthetic guide RNA of any one of the preceding embodiments, comprising a 5-aminoallyl-uracil.
- A15. The synthetic guide RNA of any one of the preceding embodiments, comprising a Z ribonucleotide.
- A16. The synthetic guide RNA of any one of the preceding embodiments, comprising a Z deoxyribonucleotide.
- A17. The synthetic guide RNA of any one of the preceding embodiments, comprising a squarate conjugation.
- A18. The synthetic guide RNA of any one of the preceding embodiments, comprising a dye linker.
- A19. The synthetic guide RNA of embodiment A18, wherein the dye linker is 2-(4-butylamidofluorescein)propane-1,3-diol bis(phosphodiester) linker.
- A20. The synthetic guide RNA of any one of the preceding embodiments, comprising a nucleotide with 2'-O-methyl and 3'-phosphorothioate.
- A21. The synthetic guide RNA of any one of the preceding embodiments, comprising a nucleotide with 2'-O-methyl 45 and 3"-PACE.
- A22. The synthetic guide RNA of any one of the preceding embodiments, comprising a nucleotide with 2'-O-methyl and 3'-thioPACE.
- A23. The synthetic guide RNA of any one of the preceding embodiments, comprising a nucleotide with 2'-deoxy and 3'-PACE.
- A24. The synthetic guide RNA of any one of the preceding embodiments, comprising a 5-methylcytidine.
- A25. The synthetic guide RNA of any one of the preceding embodiments, comprising a methylphosphonate.
- A26. The synthetic guide RNA of any one of the preceding embodiments, comprising an ester of PACE, wherein the ester is optionally a methyl ester.
- A27. The synthetic guide RNA of any one of the preced- 60 ing or nicking takes place in vivo. ing embodiments, comprising a a single RNA strand comprising both the cr RNA segment and the tracr RNA seg-
- A28. The synthetic guide RNA of any one of embodiments A1-A26, comprising two RNA strands, and the cr 65 RNA segment and the tracr RNA segment are in different RNA strands.

- A29. The synthetic guide RNA of any one of the preceding embodiments, comprising a modified nucleotide at a 5' end, 3' end, or both 5' end and 3' end of each RNA strand.
- A30. The synthetic guide RNA of any one of the preceding embodiments, comprising a modified nucleotide in the guide sequence.
- A31. The synthetic guide RNA of any one of the preceding embodiments, comprising a modified nucleotide 5' to the guide sequence.
- A32. The synthetic guide RNA of any one of the preceding embodiments, comprising a modified nucleotide in the stem sequence.
- A33. The synthetic guide RNA of any one of the preceding embodiments, comprising a modified nucleotide in the scaffold region.
- A34. The synthetic guide RNA of any one of the preceding embodiments, comprising at least one unnatural, orthogonal base pair in the scaffold region, wherein the base pair is independently selected from isoG-isoC and Z base-P 20 base.
  - A35. The synthetic guide RNA of any one of the preceding embodiments, comprising a 2'-amino group.
  - A36. The synthetic guide RNA of any one of the preceding embodiments, comprising a phosphorodithioate linkage
  - A37. The synthetic guide RNA of any one of the preceding embodiments, comprising a boranophosphonate linkage group.
- A38. The synthetic guide RNA of any one of the preceding embodiments, comprising an unlocked nucleic acid modification (ULNA).
  - A39. The synthetic guide RNA of any one of the preceding embodiments, comprising a locked nucleic acid modification (LNA).
- A40. The synthetic guide RNA of any one of the preceding embodiments, comprising an unstructured nucleic acid modification (UNA).
- A41. The synthetic guide RNA of any one of the preceding embodiments, comprising a pseudoU.
- A42. The synthetic guide RNA of any one of the preceding embodiments, comprising a 2'-MOE.
- A43. The synthetic guide RNA of any one of the preceding embodiments, comprising a 2'-arabino.
- A44. The synthetic guide RNA of any one of the preceding embodiments, comprising a 4'-thioribose.
- A45. The synthetic guide RNA of any one of the preceding embodiments, comprising a squarate linkage
- A46. The synthetic guide RNA of any one of the preceding embodiments, comprising a triazaolo linkage.
- A47. A method for cleaving or nicking a target polynucleotide comprising contacting the target polynucleotide with a CRISPR-associated protein and the synthetic guide RNA of any one of the preceding embodiments, and cleaving or nicking the target polynucleotide.
- A48. The method of embodiment A47, wherein the cleaving or nicking takes place in vitro.
- A49. The method of embodiment A47, wherein the cleaving or nicking takes place in a cell.
- A50. The method of embodiment A47, wherein the cleav-
- A51. The method of any one of embodiments A47-A50, wherein the CRISPR-associated protein is Cas9.
- A52. The method of any one of embodiments A47-A51, wherein the cleaving or nicking results in gene editing.
- A53. The method of any one of embodiments A47-A52, wherein the cleaving or nicking results in alteration of gene expression.

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A54. A method for binding a target polynucleotide comprising contacting the target polynucleotide with a CRISPR-associated protein and the synthetic guide RNA of any one of the preceding embodiments.

A55. The method of embodiment A54, wherein the 5 CRISPR-associated protein comprises a mutant which does not have a cleavage or nicking activity.

A56. The method of embodiment A54 or A55, wherein the the CRISPR-associated protein is a fusion protein comprising a protein component not naturally existing in a CRISPR 10 system.

A57. The method of any one of embodiments A54 to A56, resulting in a change of expression of the target polynucleotide.

A58. The method of any one of embodiments A54 to A57 15 the guide sequence. useful to tag the target polynucleotide. B9. The synthetic

#### FURTHER EXEMPLARY EMBODIMENTS

- B1. A synthetic guide RNA comprising:
- (a) a crRNA segment comprising (i) a guide sequence capable of hybridizing to a target sequence in a polynucleotide, (ii) a stem sequence; and
- (b) a tracrRNA segment comprising a nucleotide sequence that is partially or completely complementary to the 25 stem sequence, wherein the synthetic guide RNA comprises one or more modifications, and wherein the synthetic guide RNA has gRNA functionality.
- B2. The synthetic guide RNA of embodiment 1, comprising a 2'-O-methyl moiety, a 2'-deoxy moiety, a Z base, a 30 phosphorothioate internucleotide linkage, a phosphonoacetate internucleotide linkage, a thiophosphonoacetate internucleotide linkage, or combinations thereof.
- B3. The synthetic guide RNA of embodiment 1 or 2, comprising one or more modifications selected from the 35 group consisting of a 2'-O-methyl nucleotide with a 3'-phosphorothioate group, a 2'-O-methyl nucleotide with a 3'-phosphonoacetate group, a 2'-O-methyl nucleotide with a 3'-phosphonoacetate group, a 2'-O-methyl nucleotide with a 3'-thiophosphonoacetate group, a 2'-O-methyl 40 nucleotide with a 3'-thiophosphonoacetate group, a 2'-deoxy nucleotide with a 3'-phosphonoacetate group, a 2'-deoxy nucleotide with a 3'-thiophosphonoacetate group, a 2'-deoxy nucleotide with a 3'-thiophosphonoacetate group, and a Z base.

B4. The synthetic guide RNA of embodiment 1, 2 or 3, 45 comprising one or more modifications selected from the group consisting of a 2'-fluororibosyl, a 2-thiouracil base, a 4-thiouracil base, a 2-aminoadenine base, an hypoxanthine base, a 5-methylcytosine base, a 5-methyluracil base, a methylphosphonate internucleotide linkage, a 5-aminoally-luracil base, a squarate linkage, a triazolo linkage, a dye conjugated to a nucleotide, and combinations thereof.

B5. The synthetic guide RNA of any of the preceding embodiments, comprising a modification selected from the group consisting of a 2'-MOE, 2'-amino, 2'-F-arabino, 55 2'-LNA, 2'-ULNA, 3'-methylphosphonate, 3'-boranophosphonate, 3'-phosphorodithioate, 2'-OMe-3'-P(S)<sub>2</sub>, 2'-OMe-3'-P(CH<sub>3</sub>), 2'-OMe-3'-P(BH<sub>3</sub>), 4'-thioribosyl, L-sugar, 2-thiocytosine, 6-thioguanine, 2-aminopurine, pseudouracil, 7-deazaguanine, 7-deaza-8-azaguanine, 7-deazaadenine, 60 7-deaza-8-azaadenine, 5-hydroxymethylcytosine, 5-hydroxymethyluracil, 5,6-dehydrouracil, 5-propynylcytosine, 5-propynyluracil, 5-allyluracil, 5-allyluracil, 5-allyluracil, 5-allyluracil, 5-allyluracil, 5-allyluracil, 5-bydroxymethyluracil, 5-allyluracil, 5-all

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(O,N,S)-substituted hydrocarbon linker, (keto, carboxy, amido, thionyl, carbamoyl, or thionocarbamoyl)-containing hydrocarbon linker, spermine linker, and combinations thereof.

B6. The synthetic guide RNA of any one of the preceding embodiments, comprising a stability-enhancing modification.

B7. The synthetic guide RNA of any one of the preceding embodiments, comprising at least two modifications; wherein a first modification is a stability-enhancing modification and a second modification is a specificity-altering modification.

B8. The synthetic guide RNA of embodiment 6 or 7, wherein the stability-enhancing modification is located in the guide sequence.

B9. The synthetic guide RNA of embodiment 6 or 7, wherein the stability-enhancing modification is located upstream of the guide sequence.

B10. The synthetic guide RNA of embodiment 6 or 7, wherein the stability-enhancing modification is located within the first five and/or the last five nucleotides of the crRNA segment.

B11. The synthetic guide RNA of embodiment 6 or 7, wherein the stability-enhancing modification is located in the tracrRNA segment.

B12. The synthetic guide RNA of embodiment 6 or 7, wherein the stability-enhancing modification is located within the first five and/or the last five nucleotides of the tracrRNA segment.

B13. The synthetic guide RNA of any one of embodiments 6-12, wherein the stability-enhancing modification comprises a 2'-O-methyl moiety, a 2'-fluoro moiety, or a 2'-deoxy moiety.

B14. The synthetic guide RNA of any one of embodiments 6-13, wherein the stability-enhancing modification comprises a phosphorothioate internucleotide linkage, a phosphonoacetate internucleotide linkage, a thiophosphonoacetate internucleotide linkage, a methylphosphonate internucleotide linkage, a boranophosphate internucleotide linkage, or a phosphorodithioate internucleotide linkage.

B15. The synthetic guide RNA of any one of embodiments 6-14, wherein the stability-enhancing modification comprises a 3'-phosphonoacetate or a 3'-thiophosphonoacetate

B16. The synthetic guide RNA any one of embodiments 6-15, wherein the stability-enhancing modification comprises a 2'-O-methyl-3'-phosphorothioate nucleotide, a 2'-O-methyl-3'-phosphonoacetate nucleotide, or a 2'-O-methyl-3'-thiophosphonoacetate nucleotide.

B17. The synthetic guide RNA of any one of embodiments 6-16, wherein the stability-enhancing modification comprises a 2'-fluoro-3'-phosphorothioate nucleotide, a 2'-fluoro-3'-phosphonoacetate nucleotide, or a 2'-fluoro-3'-thiophosphonoacetate nucleotide.

B18. The synthetic guide RNA of any one of the preceding embodiments, comprising a specificity-altering modification

B19. The synthetic guide RNA of embodiment 18, wherein the specificity-altering modification is located in the guide sequence.

B20. The synthetic guide RNA of any one of embodiment 18 or 19, wherein the specificity-altering modification comprises a 2-thiouracil, a 4-thiouracil or a 2-aminoadenine.

B21. The synthetic guide RNA of any one of embodiments 18-20, wherein the specificity-altering modification comprises a phosphorothioate internucleotide linkage, a phosphonoacetate internucleotide linkage, a thiophosphono-

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acetate internucleotide linkage, a methylphosphonate internucleotide linkage, a boranophosphate internucleotide linkage, or a phosphorodithioate internucleotide linkage.

B22. The synthetic guide RNA of any one of embodiments 18-21, wherein the specificity-altering modification 5 comprises a 3'-phosphonoacetate or a 3'-thiophosphonoacetate.

B23. The synthetic guide RNA of any one of the preceding embodiments, comprising a fluorescent dye or a label.

B24. The synthetic guide RNA of any one of the preceding embodiments, comprising one or more isotopic labels.

B25. The synthetic guide RNA of any one of the preceding embodiments, wherein the guide RNA is conjugated to an oligonucleotide, an aptamer, an amino acid, a peptide, a protein, a steroid, a lipid, a folic acid, a vitamin, a sugar, or 15 an oligosaccharide.

B26. The synthetic guide RNA of any one of the preceding embodiments, wherein the synthetic guide RNA is a single guide RNA, wherein the crRNA segment and the tracrRNA segment are linked through a loop L.

B27. The synthetic guide RNA of embodiment 26, wherein the loop L comprises 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 nucleotides.

B28. The synthetic guide RNA of embodiment 26 or 27, wherein the loop L comprises a nucleotide sequence of 25 GNRA, wherein N represents A, C, G, or U and R represents A or G.

B29. The synthetic guide RNA of embodiment 26, 27 or 28, wherein the loop L comprises a nucleotide sequence of GAAA

B30. The synthetic guide RNA of any one of embodiments 26-29, wherein the loop L comprises one or more modified nucleotides.

B31. The synthetic guide RNA of embodiment 30, wherein the loop L comprises a fluorescent dye.

B32. The synthetic guide RNA of embodiment 31, wherein the dye is conjugated to a 2-(4-butylamido-dye) propane-1,3-diol bis(phosphodiester) linker.

B33. The synthetic guide RNA of any one of the preceding embodiments, wherein the crRNA segment is at the 5' 40 end of the guide RNA.

B34. The synthetic guide RNA of any one of the preceding embodiments, wherein the tracrRNA segment is at the 3' end of the guide RNA.

B35. The synthetic guide RNA of any of the preceding 45 embodiments, wherein the crRNA segment comprises from 25 to 70 nucleotides.

B36. The synthetic guide RNA of any of the preceding embodiments, wherein the guide sequence comprises from 15 to 30 nucleotides.

B37. The synthetic guide RNA of any of the preceding embodiments, wherein the stem sequence comprises from 10 to 50 nucleotides.

B38. The synthetic guide RNA of any of the preceding embodiments, comprising one or more triazolo linkage(s). 55

B39. The synthetic guide RNA of any of the preceding embodiments, comprising one or more squarate linkage(s).

B40. The synthetic guide RNA of any of the preceding embodiments, wherein the guide RNA comprises a nucleotide composition of:

 $M_m N_n$ 

wherein each N independently represents an unmodified nucleotide and each M is selected from a 2'-O-methyl ribonucleotide, a 2'-O-methyl-3'-P(S) ribonucleotide, a 2'-O-methyl-3'-PACE ribonucleotide, a 2'-O-methyl-3'-thioPACE ribonucleotide, and a 2'-deoxynucleotide;

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wherein each M is at any position in the sequence of the guide RNA; and

wherein m is an integer between 1 and 220, and n is an integer between 0 and 219, and 50<m+n≤220.

B41. The synthetic guide RNA of embodiment 38, wherein m+n≤150, and each of m and n are independently selected from an integer between 0 and 150, provided that m is not 0.

B42. The synthetic guide RNA of any of the preceding embodiments, wherein the guide RNA comprises a nucleotide sequence of:

$$M_m N_n M'_m N'_n M''_{m''}$$

wherein each M, M' and M" independently represent a modified nucleotide and each N and N' independently represent an unmodified ribonucleotide;

wherein any given M is the same or different from any other M, any given N is the same or different from any other N, any given M' is the same or different from any other M', any given N' is the same or different from any other N', any given M" is the same or different from any other M"; and wherein m is an integer between 0 and 40, n is an integer between 20 and 130, m' is an integer between 0 and 10, n' is an integer between 0 and 10, provided that m+m'+m" is greater than or equal to

B43. The synthetic guide RNA of any of the preceding embodiments, wherein the crRNA segment comprises a nucleotide sequence of:

$$M_m N_n M'_m N'_n$$

wherein M and M' each represent a modified nucleotide and N and N' each represent an unmodified ribonucleotide; wherein any given M is the same or different from any other M, any given N is the same or different from any other N, any given M' is the same or different from any other M', any given N' is the same or different from any other N'; and

wherein n and n' are each independently selected from an integer between 0 and 50, and wherein m and m' are each independently selected from an integer between 0 and 25, provided that m+m' is greater than or equal to 1.

B44. The synthetic guide RNA of any of the preceding embodiments, wherein the guide sequence comprises a nucleotide sequence of:

$$M_m N_n M'_m N'_n$$

wherein M and M' each represent a modified nucleotide and N and N' each represent an unmodified ribonucleotide;

wherein any given M is the same or different from any other M, any given N is the same or different from any other N, any given M' is the same or different from any other M', any given N' is the same or different from any other N'; and

wherein m, n, m', and n' are each independently selected from an integer between 0 and 40, provided that m+m' is greater than or equal to 1.

B45. The synthetic guide RNA of any of the preceding embodiments, wherein the tracrRNA segment comprises a nucleotide sequence of:

$$N_n M_m N'_n M'_m$$

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wherein M and M' each represent a modified nucleotide and N and N' each represent an unmodified ribonucleotide; wherein any given M is the same or different from any other M, any given N is the same or different from any other N, any given M' is the same or different from any other M', any given N' is the same or different from any other N'; and

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wherein n is an integer between 0 and 130 m is an integer between 0 and 40, and n' is an integer between 0 and 130, and m' is an integer between 0 and 40, provided that m+m' is greater than or equal to 1.

B46. The synthetic guide RNA of any one of embodiments 40-43, wherein m, m', m+m', or m+m'+m" is 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20.

B47. The synthetic guide RNA of any one of embodiments 40-43, wherein m, m', m+m', or m+m'+m" is 1, 2, 3,

B48. The synthetic guide RNA of any one of embodiments 40-45, wherein n is 16, 17, 18, or 19.

B49. The synthetic guide RNA of any one of embodiments 40-45, wherein n, n', or n+n' is an integer between 75 15 and 115.

B50. The synthetic guide RNA of any one of embodiments 40-47, wherein each M is independently selected from the group consisting of a 2'-modified nucleotide, a 3'-modified nucleotide, and combinations thereof.

B51. The synthetic guide RNA of embodiment 48, wherein the 2'-modified nucleotide is selected from the group consisting of a 2'-deoxy nucleotide, a 2'-O-methyl nucleotide, a 2'-fluoro nucleotide, and a 2'-O-C<sub>1-3</sub>alkyl- $O-C_{1-3}$ alkyl nucleotide.

B52. The synthetic guide RNA of embodiment 48, wherein the 3'-modified nucleotide is selected from the group consisting of a 3'-phosphonoacetate nucleotide and a 3'-thiophosphonoacetate nucleotide.

B53. The synthetic guide RNA of embodiment 48, 30 wherein the combination of the 2'-modified nucleotide and the 3'-modified nucleotide comprises a 2'-O-methyl-3'-phosphorothioate nucleotide, a 2'-O-methyl-3'-phosphonoacetate nucleotide, or a 2'-O-methyl-3'-thiophosphonoacetate nucleotide.

B54. A method for cleaving a target polynucleotide comprising contacting the target polynucleotide with a CRISPRassociated protein and the synthetic guide RNA of any one of the preceding embodiments and cleaving the target poly-

B55. The method of embodiment 52, further comprising contacting the target polynucleotide with an exogenous CRISPR-associated protein.

B56. The method of embodiment 53, wherein the CRISPR-associated protein is Cas9.

B57. The method of any one of embodiments 52-54, wherein the cleavage results in a functional knockout of a target gene.

B58. The method of any one of embodiments 52-55, further comprising repairing the cleaved target polynucle- 50 otide by homology-directed repair with an exogenous or endogenous template polynucleotide.

B59. The method of embodiment 56, wherein the exogenous or endogenous template polynucleotide comprises at least one sequence having substantial sequence identity with 55 embodiments, having a single RNA strand or two separate a sequence on either side of the cleavage site.

B60. The method of any one of embodiments 52-57, further comprising repairing the cleaved target polynucleotide by non-homologous end joining.

B61. The method of any one of embodiments 56-58, 60 wherein the repairing step produces an insertion, deletion, or substitution of one or more nucleotides of the target polynucleotide.

B62. The method of embodiment 59, wherein the insertion, deletion, or substitution results in one or more amino 65 acid changes in a protein expressed from a gene comprising the target polynucleotide.

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B63. The method of any one of embodiments 52-60, wherein the target polynucleotide is contacted with the CRISPR-associated protein and the synthetic guide RNA in

B64. The method of any one of embodiments 52-61, wherein the target polynucleotide contacted with the CRISPR-associated protein and the synthetic guide RNA is within the genome of a cell in vitro or in vivo.

B65. The method of embodiment 62, wherein the cell is isolated from a multicellular source prior to contacting the target polynucleotide with the CRISPR-associated protein and the synthetic guide RNA.

B66. The method of embodiment 63, wherein the source is a plant, an animal, a multicellular protist, or a fungus.

B67. The method of any one of embodiments 62-64, wherein the cell, or a cell derived therefrom, is returned to the source after contacting the target polynucleotide with the CRISPR-associated protein and the synthetic guide RNA.

B68. A method of modifying a target polynucleotide in a cell comprising introducing into the cell the synthetic guide RNA of any one of embodiments 1-51 and introducing into the cell a CRISPR-associated protein or a nucleic acid that expresses a CRISPR-associated protein in the cell.

B69. The method of embodiment 66, wherein the CRISPR-associated-protein is Cas9.

B70. A method of altering expression of at least one gene product in a cell comprising introducing into the cell the synthetic guide RNA of any one of embodiments 1-51 and further introducing into the cell a CRISPR-associated-protein or a nucleic acid that expresses a CRISPR-associated protein in the cell, wherein the cell contains and expresses a DNA molecule having a target sequence and encoding the 35 gene product.

B71. The method of embodiment 68, wherein the CRISPR-associated-protein is Cas9.

B72. The method of embodiment 69, wherein the CRISPR-associated-protein cleaves the DNA molecule.

B73. A set or library of RNA molecules comprising two or more synthetic guide RNAs of any one of embodiments

B74. A kit comprising the synthetic guide RNA of any one of embodiments 1-51 or the set or library of RNA molecules of embodiment 71.

B75. The kit of embodiment 72, further comprising a CRISPR-associated protein or a nucleic acid encoding the CRISPR-associated protein.

B76. The kit of embodiment 73, wherein the CRISPRassociated-protein is Cas9.

B77. The synthetic guide RNA, method or kit of any of the preceding embodiments, wherein the synthetic guide RNA comprises an end modification.

B78. The synthetic guide RNA of any of the preceding complementary RNA strands, wherein the guide RNA comprises at least one stability-enhancing modification at both ends of each RNA strand.

C1. A synthetic guide RNA comprising:

- (a) a crRNA segment comprising (i) a guide sequence capable of hybridizing to a target sequence in a polynucleotide, (ii) a stem sequence; and
- (b) a tracrRNA segment comprising a nucleotide sequence that is partially or completely complementary to the stem sequence, wherein the synthetic guide RNA comprises one or more modifications, and wherein the synthetic guide RNA has gRNA functionality.

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- C2. The synthetic guide RNA of embodiment C1, wherein one or more of the modifications comprises a stability-enhancing modification.
- C3. The synthetic guide RNA of embodiment C2, wherein one or more of the stability-enhancing modifications is <sup>5</sup> located in the guide sequence.
- C4. The synthetic guide RNA of embodiment C2, wherein the stability-enhancing modification comprises a 2'-Omethyl moiety, a Z base, a 2'-deoxy nucleotide, a phosphorothioate internucleotide linkage, a phosphonoacetate (PACE) internucleotide linkage, or a thiophosphonoacetate (thioPACE) internucleotide linkage, or combinations thereof
- C5. The synthetic guide RNA of any of the foregoing embodiments, comprising less than 26 consecutive 2'-Omethyl modified nucleotides at a 5' end of the guide RNA.
- C6. The synthetic guide RNA of any of the foregoing embodiments, comprising a Z base replacing a cytosine in the synthetic guide RNA.
- C7. The synthetic guide RNA of any of the foregoing embodiments, comprising at least one 2-thiouracil at a position corresponding to a uridine that can engage in U-G wobble pairing with a potential off-target sequence.
- C8. The synthetic guide RNA of any of the foregoing 25 embodiments, comprising one or more modifications selected from the group consisting of a 2'-O-methyl nucleotide with a 3'-phosphorothioate group, a 2'-O-methyl nucleotide with a 3'-phosphonoacetate group, a 2'-O-methyl nucleotide with a 3'-thiophosphonoacetate group, and a 30 2'-deoxy nucleotide with a 3'-phosphonoacetate group.
- C9. The synthetic guide RNA of any of the foregoing embodiments, comprising at least two modifications.
- C10. The synthetic guide RNA any of the foregoing embodiments, comprising up to 50 modifications.
- C11. The synthetic guide RNA of any of the foregoing embodiments, comprising a single RNA strand or two separate RNA strands, and one or more modifications at a 5' end of each RNA strand, at a 3' end of each RNA strand, or at both a 5' end and a 3' end of each RNA strand.
- C12. The synthetic guide RNA of any of the foregoing embodiments, comprising 7 or fewer consecutive modified nucleotides at a 5' end or at a 3' end or at each of 5' and 3' ends.
- C13. The synthetic guide RNA of any of the foregoing 45 embodiments, comprising one or more 5-methyluridine nucleotides at one or more of a 5' end, a 3' end, or a stem-loop.
- C14. The synthetic guide RNA of any of the foregoing embodiments, wherein one or more of the modifications 50 alters base-pairing thermostability.
- C15. The synthetic guide RNA of embodiment C14, wherein said one or more modifications enhances the base-pairing thermostability.
- C16. The synthetic guide RNA of embodiment C15, 55 wherein said one or more modifications is independently selected from a 2-thiouracil (2-thioU), a 4-thiouracil (4-thioU), a 2-aminoadenine, a 2'-O-methyl, a 2'-fluoro, a 5-methyluridine, a 5-methylcytidine, and a locked nucleic acid modification (LNA).
- C17. The synthetic guide RNA of embodiment C15, wherein said one or more modifications decreases the base-pairing thermostability.
- C18. The synthetic guide RNA of embodiment C17, 65 wherein said one or more modifications is independently selected from a 2-thiouracil, a 2'-deoxy, a phosphorothioate

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linkage, a phosphorodithioate linkage, a boranophosphonate linkage, a phosphonoacetate linkage, a thiophosphonoacetate linkage, and an unlocked nucleic acid modification (ULNA).

- C19. The synthetic guide RNA of any of the foregoing embodiments, comprising one or more 2'-O-methylA-2'-O-methylU base pairs.
- C20. The synthetic guide RNA of any of the foregoing embodiments, wherein one or more of the modifications is a specificity-altering modification.
- C21. The synthetic guide RNA of embodiment C20, wherein the specificity-altering modification is located in the guide sequence.
- C22. The synthetic guide RNA of any of the foregoing embodiments, wherein the specificity-altering modification comprises a 2-thiouracil, a 4-thiouracil, a 2-aminoadenine, a 2'-O-methyl, a 2'-fluoro, a LNA, a phosphorothioate linkage, a phosphorodithioate linkage, a boranophosphonate linkage, a phosphonoacetate linkage, a thiophosphonoacetate linkage, an ULNA, a 2'-deoxy, a 5-methyluridine, a 5-methylcytidine, or combinations thereof.
- C23. The synthetic guide RNA of any of the foregoing embodiments, comprising a fluorescent dye or a label.
- C24. The synthetic guide RNA of embodiment C23, wherein the fluorescent dye or a label is bound to a stemloop of the synthetic guide RNA.
- C25. A method for genomic editing to modify a DNA sequence, or regulating the expression of a gene of interest, or cleaving a target polynucleotide comprising: contacting the DNA sequence, the gene of interest, or the target polynucleotide with a CRISPR-associated protein and the synthetic guide RNA of any of the foregoing embodiments, and editing, regulating or cleaving the DNA sequence, the gene of interest or the target polynucleotide.
  - C26. The method of embodiment C25, wherein the method is performed in vitro, and the synthetic guide RNA comprises 15 or more 2'-O-methyl modifications throughout the guide sequence.
  - C27. A set or library of RNA molecules comprising two or more synthetic guide RNAs of any of the foregoing embodiments.
  - C28. A kit comprising the synthetic guide RNA of any of the foregoing embodiments.
  - C29. An array of RNA molecules comprising two or more synthetic guide RNAs of any of the foregoing embodiments.

The foregoing description of exemplary or preferred embodiments should be taken as illustrating, rather than as limiting, the present invention as defined by the claims. As will be readily appreciated, numerous variations and combinations of the features set forth above can be utilized without departing from the present invention as set forth in the claims. Such variations are not regarded as a departure from the scope of the invention, and all such variations are intended to be included within the scope of the following claims. All references cited herein are incorporated by reference in their entireties.

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SEQUENCE LISTING

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65 65	Arg	Thr	Ala	Arg	Arg 70	Arg	Tyr	Thr	Arg	Arg 75	Lys	Asn	Arg	Ile	80 GÀa
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His	Glu	Arg 115	His	Pro	Ile	Phe	Gly 120	Asn	Ile	Val	Asp	Glu 125	Val	Ala	Tyr
His	Glu 130	Tàa	Tyr	Pro	Thr	Ile 135	Tyr	His	Leu	Arg	Lys 140	ГÀа	Leu	Val	Asp
Ser 145	Thr	Aap	Lys	Ala	Asp 150	Leu	Arg	Leu	Ile	Tyr 155	Leu	Ala	Leu	Ala	His 160
Met	Ile	Lys	Phe	Arg 165	Gly	His	Phe	Leu	Ile 170	Glu	Gly	Asp	Leu	Asn 175	Pro
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Asn	Gln	Leu 195	Phe	Glu	Glu	Asn	Pro 200	Ile	Asn	Ala	Ser	Gly 205	Val	Asp	Ala
Lys	Ala 210	Ile	Leu	Ser	Ala	Arg 215	Leu	Ser	Lys	Ser	Arg 220	Arg	Leu	Glu	Asn
Leu 225	Ile	Ala	Gln	Leu	Pro 230	Gly	Glu	Lys	ГÀа	Asn 235	Gly	Leu	Phe	Gly	Asn 240
Leu	Ile	Ala	Leu	Ser 245	Leu	Gly	Leu	Thr	Pro 250	Asn	Phe	ГÀа	Ser	Asn 255	Phe
Asp	Leu	Ala	Glu 260	Aap	Ala	Lys	Leu	Gln 265	Leu	Ser	ГÀа	Asp	Thr 270	Tyr	Asp
Asp	Asp	Leu 275	Asp	Asn	Leu	Leu	Ala 280	Gln	Ile	Gly	Asp	Gln 285	Tyr	Ala	Asp
Leu	Phe 290	Leu	Ala	Ala	Lys	Asn 295	Leu	Ser	Asp	Ala	Ile 300	Leu	Leu	Ser	Asp
Ile 305	Leu	Arg	Val	Asn	Thr 310	Glu	Ile	Thr	Lys	Ala 315	Pro	Leu	Ser	Ala	Ser 320
Met	Ile	Lys	Arg	Tyr 325	Asp	Glu	His	His	Gln 330	Asp	Leu	Thr	Leu	Leu 335	ГÀа
Ala	Leu	Val	Arg 340	Gln	Gln	Leu	Pro	Glu 345	Lys	Tyr	ГÀа	Glu	Ile 350	Phe	Phe
Asp	Gln	Ser 355	Lys	Asn	Gly	Tyr	Ala 360	Gly	Tyr	Ile	Asp	Gly 365	Gly	Ala	Ser

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Gln	Glu 370	Glu	Phe	Tyr	Lys	Phe 375	Ile	Lys	Pro	Ile	Leu 380	Glu	Lys	Met	Asp
Gly 385	Thr	Glu	Glu	Leu	Leu 390	Val	Lys	Leu	Asn	Arg 395	Glu	Asp	Leu	Leu	Arg 400
Lys	Gln	Arg	Thr	Phe 405	Asp	Asn	Gly	Ser	Ile 410	Pro	His	Gln	Ile	His 415	Leu
Gly	Glu	Leu	His 420	Ala	Ile	Leu	Arg	Arg 425	Gln	Glu	Asp	Phe	Tyr 430	Pro	Phe
Leu	Lys	Asp 435	Asn	Arg	Glu	Lys	Ile 440	Glu	Lys	Ile	Leu	Thr 445	Phe	Arg	Ile
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Glu Glu Gly Ile Lys Glu Leu Gly Ser Gln Ile Leu Lys Glu His Pro

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Lys	Lys 1130	_	Gly	/ Gly	7 Ph€	Asp		er P:	ro T	hr V		la 140	Tyr	Ser	Val
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Ser	Val 1160	_	; Glu	ı Lev	ı Lev	ι Gl <sub>Σ</sub> 116		le Tl	nr I	le M		lu 170	Arg	Ser	Ser
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<sup>&</sup>lt;220> FEATURE:

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aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
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<222> LOCATION: (1)..(6)
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<220> FEATURE:
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<222> LOCATION: (110) .. (113)
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aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
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<222> LOCATION: (1)..(2)
<220> FEATURE:
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                                                                        6.0
aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
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<220> FEATURE:
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<220> FEATURE:
<221> NAME/KEY: thiophosphonoacetate internucleotide linkage
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<400> SEQUENCE: 112
aguccucauc ucccucaagc guuuaagagc uaugcuggua acagcauagc aaguuuaaau
                                                                        60
aaggcuaguc cguuu
                                                                        75
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<220> FEATURE:
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                                                                        77
aaggcuaguc cguuauc
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<222> LOCATION: (1)..(2)
<220> FEATURE:
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<222> LOCATION: (77)..(77)
<220> FEATURE:
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<222> LOCATION: (77)..(78)
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uaaggcuagu ccguuauc
                                                                        78
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<222> LOCATION: (1)..(4)
<220> FEATURE:
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<222> LOCATION: (110) .. (112)
<220> FEATURE:
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<222> LOCATION: (110) .. (113)
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qcaqauquaq uquuuccaca quuuaaqaqc uauqcuqqaa acaqcauaqc aaquuuaaau
                                                                       60
aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
                                                                      113
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<220> FEATURE:
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<220> FEATURE:
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<222> LOCATION: (112) .. (113)
<400> SEQUENCE: 116
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gcagauguag uguuuccaca guuuaagagc uaugcuggaa acagcauagc aaguuuaaau
                                                                       113
aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
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<212> TYPE: RNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
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gauguugucg augaaaaagu guuuaagagc uaugcuggua acagcauagc aaguuuaaau
                                                                        60
                                                                       113
aaqqcuaquc cquuaucaac uuqaaaaaqu qqcaccqaqu cqquqcuuuu uuu
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<212> TYPE: RNA
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<220> FEATURE:
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<222> LOCATION: (15)..(15)
<400> SEQUENCE: 118
gauguugucg augaaaaagu guuuaagagc uaugcuggua acagcauagc aaguuuaaau
                                                                        60
aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
                                                                       113
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<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
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<400> SEQUENCE: 119
gauuuagacg aaggauugaa guuuaagagc uaugcuggua acagcauagc aaguuuaaau
                                                                        60
aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
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<400> SEQUENCE: 120
gauuuagacg aaggauugaa guuuaagagc uaugcuggua acagcauagc aaguuuaaau
aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
                                                                       113
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<210> SEQ ID NO 121

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<220> FEATURE:
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<222> LOCATION: (8) .. (8)
<220> FEATURE:
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<222> LOCATION: (11)..(11)
<220> FEATURE:
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<222> LOCATION: (13)..(15)
<220> FEATURE:
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<222> LOCATION: (31)..(31)
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<220> FEATURE:
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<222> LOCATION: (39)..(39)
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<220> FEATURE:
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<222> LOCATION: (81)..(82)
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<222> LOCATION: (90)..(90)
<220> FEATURE:
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<222> LOCATION: (104) .. (104)
<220> FEATURE:
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<222> LOCATION: (107) .. (112)
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gcagauguag uguuuccaca guuuaagagc uaugcuggua acagcauagc aaguuuaaau
                                                                       60
aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
                                                                      113
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<212> TYPE: RNA
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<223> OTHER INFORMATION: synthetic construct
<220> FEATURE:
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<222> LOCATION: (70)..(71)
<223 > OTHER INFORMATION: Z base
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (70)..(71)
<223> OTHER INFORMATION: n is a, c, g, or u, or unknown or other
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aguccucauc ucccucaagc guuuaagagc uaugcuggua acagcauagc aaguuuaaau
                                                                       60
aaggcuagun nguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
                                                                      113
<210> SEQ ID NO 123
<211> LENGTH: 100
<212> TYPE: RNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
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<222> LOCATION: (1) .. (1)
<220> FEATURE:
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<222> LOCATION: (1)..(2)
<220> FEATURE:
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<220> FEATURE:
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<222> LOCATION: (57)..(57)
<220> FEATURE:
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<222> LOCATION: (99)..(99)
<220> FEATURE:
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<222> LOCATION: (99)..(100)
<400> SEQUENCE: 123
aquecucaue ucceucaage quuuaagage uaquaauage aaguuuaaau aagguuaaue
                                                                       60
                                                                      100
cguuaucaac aagaaauugu ggcaccgagu cggugcuuuu
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<220> FEATURE:
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<222> LOCATION: (1)..(2)
<220> FEATURE:
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<220> FEATURE:
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<222> LOCATION: (67)..(67)
<220> FEATURE:
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<222> LOCATION: (112) .. (112)
<220> FEATURE:
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<222> LOCATION: (112) .. (113)
<400> SEQUENCE: 124
aguccucauc ucccucaagc guuuaagagc uaugcuggua acagcauagc aaguuuaaau
aagguuaauc cguuaucaac aagaaauugu ggcaccgagu cggugcuuuu uuu
                                                                      113
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<222> LOCATION: (95)..(96)
<223> OTHER INFORMATION: Z base
<220> FEATURE:
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<222> LOCATION: (95)..(96)
<223> OTHER INFORMATION: n is a, c, g, or u, or unknown or other
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aguccucauc ucccucaagc guuuaagagc uaugcuggua acagcauagc aaguuuaaau
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aaqqcuaquc cquuaucaac uuqaaaaaqu qqcannqaqu cqquqcuuuu uuu
                                                                      113
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<220> FEATURE:
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<220> FEATURE:
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<222> LOCATION: (73)..(73)
<220> FEATURE:
<221> NAME/KEY: thiophosphonoacetate internucleotide linkage
<222> LOCATION: (73)..(74)
<400> SEQUENCE: 126
aguccucauc ucccucaagc guuuaagagc uaugcuggua acagcauagc aaguuuaaau
                                                                        60
aaqqcuaquc cquu
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<220> FEATURE:
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<220> FEATURE:
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<222> LOCATION: (74)..(75)
<400> SEQUENCE: 127
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aaggcuaguc cguua
                                                                        75
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<212> TYPE: DNA
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<220> FEATURE:
<223 > OTHER INFORMATION: synthetic construct
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<220> FEATURE:
<223> OTHER INFORMATION: synthetic construct
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aatatacttg tattgagtta aacatttttt cccataaccc cttaagtaat
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<210> SEQ ID NO 130

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<220> FEATURE:
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auaacucaau uuguaaaaaa guuuuagagc uauagcaagu uaaaauaagg uaguccguua
                                                                        60
ucaacuugaa aaaguggcac cgagucggug cuuuuuuu
                                                                        98
<210> SEQ ID NO 131
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<212> TYPE: RNA
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<223 > OTHER INFORMATION: synthetic construct
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<221> NAME/KEY: 2'-deoxy-nucleotide
<222> LOCATION: (1) .. (20)
<400> SEQUENCE: 131
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                                                                       60
                                                                      113
aaqqcuaquc cquuaucaac uuqaaaaaqu qqcaccqaqu cqquqcuuuu uuu
<210> SEO ID NO 132
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<212> TYPE: RNA
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<223 > OTHER INFORMATION: synthetic construct
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<223> OTHER INFORMATION: Z base
<220> FEATURE:
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<222> LOCATION: (95)..(96)
<223> OTHER INFORMATION: n is a, c, g, or u, or unknown or other
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aguccucauc ucccucaagc guuuaagagc uaugcuggua acagcauagc aaguuuaaau
                                                                       60
aaggcuaguc cguuaucaac uugaaaaagu ggcanngagu cggugcuuuu uuu
                                                                      113
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<400> SEQUENCE: 133
aquecucaue ucceucaage quuuaagage uaugeuggua acageauage aaguuuaaau
                                                                       60
aaggcuaguc cguuaucaac uugaaaaagu ggcaccgagu cggugcuuuu uuu
                                                                      113
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ccagccaage gcacctaatt tee
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<223> OTHER INFORMATION: site for fluorescent dye or label attachment
ggaaauuagg ugcgcuuggc guuuuagagc uagaaauagc aaguuaaaau aaggcuaguc
cgccaucaac uugaaaaagc ggcaccga
                                                                        88
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<212> TYPE: DNA
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<220> FEATURE:
<223 > OTHER INFORMATION: synthetic construct
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agtecteate teecteaage agg
                                                                        23
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cctgcttgag ggagatgagg act
                                                                       23
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<212> TYPE: DNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: synthetic construct
<400> SEQUENCE: 138
agtecteaac teeeteaage agg
                                                                        23
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<212> TYPE: DNA
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<400> SEQUENCE: 139
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<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: synthetic construct
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<211> LENGTH: 23
<212> TYPE: DNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: synthetic construct
<400> SEQUENCE: 142
                                                                         23
actectcate eccetcaage egg
<210> SEQ ID NO 143
<211> LENGTH: 23
<212> TYPE: DNA
<213 > ORGANISM: Artificial Sequence
<220> FEATURE:
<223 > OTHER INFORMATION: synthetic construct
<400> SEQUENCE: 143
                                                                         2.3
cctgcttgag ggggatgagg agt
```

#### We claim:

- 1. A synthetic CRISPR guide RNA comprising:
- (a) a crRNA segment comprising (i) a guide sequence capable of hybridizing to a target sequence in a polynucleotide, (ii) a stem sequence; and
- (b) a tracrRNA segment comprising a nucleotide sequence that is partially or completely complementary to the stem sequence,

wherein the synthetic guide RNA has gRNA functionality comprising associating with a Cas protein and targeting the gRNA:Cas protein complex to the target sequence, and comprises one or more modifications in the guide sequence, 45 wherein the one or more modifications comprises a 2'-O-methyl.

- 2. A method for genome editing to modify a DNA sequence, or for regulating the expression of a gene of interest, or for cleaving a target polynucleotide, or for binding a target polynucleotide comprising: contacting the DNA sequence, the gene of interest, or the target polynucleotide with a CRISPR-associated (Cas) protein and the synthetic guide RNA of claim 1, and editing, regulating, cleaving, or binding the DNA sequence, the gene of interest, or the target polynucleotide.
- 3. A set or library of RNA molecules comprising two or more synthetic guide RNAs of claim 1.
- **4.** The synthetic guide RNA of claim **1** wherein the guide RNA is a single-guide RNA (sgRNA).
- 5. The synthetic guide RNA of claim 1, wherein said one or more modifications comprises a 2'-O-methyl nucleotide with a 3'-phosphorothioate.
- **6**. The synthetic guide RNA of claim **1**, wherein said one 65 or more modifications comprises a 2'-O-methyl nucleotide with a 3'-phosphonoacetate.

7. The synthetic guide RNA of claim 1, wherein said one or more modifications comprises a 2'-O-methyl nucleotide with a 3'-thiophosphonoacetate.

- **8**. The synthetic guide RNA of claim **1** further comprising one or more phosphorothioate internucleotide linkage, phosphonoacetate (PACE) internucleotide linkage, and/or thiophosphonoacetate (thioPACE) internucleotide linkage.
- **9**. The synthetic guide RNA of claim **1**, further comprising up to three phosphorothioate, PACE, and/or thioPACE internucleotide linkages in the guide sequence.
- 10. The synthetic guide RNA of claim 1, further comprising up to seven phosphorothioate, PACE, and/or thio-PACE internucleotide linkages in the guide sequence.
- 11. The synthetic guide RNA of claim 1, further comprising up to ten phosphorothioate, PACE, and/or thioPACE internucleotide linkages in the guide sequence.
- 12. The synthetic guide RNA of claim 1, comprising up to five consecutive phosphorothioate internucleotide linkages at a 5'-end of the guide RNA.
- 13. The synthetic guide RNA of claim 12, further comprising up to five consecutive phosphorothioate, PACE, and/or thioPACE internucleotide linkages at a 3'-end of the guide RNA.
- 14. The synthetic guide RNA of claim 1, further comprising a fluorophore at a 5'-end of the guide RNA.
- **15**. The synthetic guide RNA of claim 1, comprising one or more end modification.
- **16**. The synthetic guide RNA of claim **1**, comprising at least 2 consecutive 2'-O-methyl modifications.
- 17. The synthetic guide RNA of claim 1, comprising at least six 2'-O-methyl modifications.
- **18**. The synthetic guide RNA of claim **1**, comprising at least twenty 2'-O-methyl modifications.

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- 19. A synthetic CRISPR crRNA molecule comprising a guide sequence capable of hybridizing to a target sequence in a polynucleotide, wherein the synthetic crRNA molecule comprises one or more modifications in the guide sequence;
  - wherein the synthetic crRNA molecule has gRNA functionality comprising associating with a Cas protein and targeting the gRNA:Cas protein complex to the target sequence; and
  - wherein the one or more modifications comprises a 2'-O-methyl.
- **20**. The synthetic CRISPR crRNA of claim **19**, further comprising one or more phosphorothioate internucleotide linkage, phosphonoacetate (PACE) internucleotide linkage, and/or thiophosphonoacetate (thioPACE) internucleotide linkage.
- **21**. The synthetic CRISPR crRNA of claim **19**, wherein said one or more modifications comprises a 2'-O-methyl nucleotide with a 3'-phosphorothioate.
- **22**. The synthetic CRISPR crRNA of claim **19**, wherein said one or more modifications comprises a 2'-O-methyl nucleotide with a 3'-phosphonoacetate.
- 23. The synthetic CRISPR crRNA of claim 19, wherein said one or more modifications comprises a 2'-O-methyl nucleotide with a 3'-thiophosphonoacetate.
- **24**. The synthetic CRISPR crRNA of claim **19**, further comprising up to three phosphorothioate, PACE, and/or thioPACE internucleotide linkages in the guide sequence.
- **25**. The synthetic CRISPR crRNA of claim **19**, further comprising up to seven phosphorothioate, PACE, and/or thioPACE internucleotide linkages in the guide sequence.

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- **26**. The synthetic CRISPR crRNA of claim **19**, further comprising up to ten phosphorothioate, PACE, and/or thio-PACE internucleotide linkages in the guide sequence.
- 27. The synthetic CRISPR crRNA of claim 19, comprising up to five consecutive phosphorothioate internucleotide linkages at a 5'-end of the crRNA.
- **28**. The synthetic CRISPR crRNA of claim **27**, further comprising up to five consecutive phosphorothioate, PACE, and/or thioPACE internucleotide linkages at a 3'-end of the crRNA
- **29**. The synthetic CRISPR crRNA of claim **19**, further comprising a fluorophore at a 5'-end of the crRNA.
- **30**. The synthetic CRISPR crRNA of claim **19**, comprising at least 2 consecutive 2'-O-methyl modifications.
- 31. The synthetic CRISPR crRNA of claim 19, comprising at least six 2'-O-methyl modifications.
- **32**. The synthetic CRISPR crRNA of claim **19**, comprising at least twenty 2'-O-methyl modifications.
- 33. A method for genome editing to modify a DNA sequence, or for regulating the expression of a gene of interest, or for cleaving a target polynucleotide, or for binding a target polynucleotide comprising: contacting the DNA sequence, the gene of interest, or the target polynucleotide with a CRISPR-associated (Cas) protein and the CRISPR crRNA of claim 19, and editing, regulating, cleaving, or binding the DNA sequence, the gene of interest, or the target polynucleotide.

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